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(71) Applicant (for all designated States except US): NATURE TECHNOLOGY CORPORATION [US/US]; 109 South 54th Street, Omaha, NE 68132 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): HODGSON, Clague, P. [US/US]; 109 South 54th Street, Omaha, NE 68132 (US). ZINK, Mary, Ann [US/US]; 109 South 54th Street, Omaha, NE 68132 (US). XU, Guoping [CN/US]; 109 South 54th Street, Omaha, NE 68132 (US).

(74) Agent: MCCORMACK, Myra, H.; Muetting, Raasch & Gebhardt, P.A., P.O. Box 581415, Minneapolis, MN 55458-1415 (US).

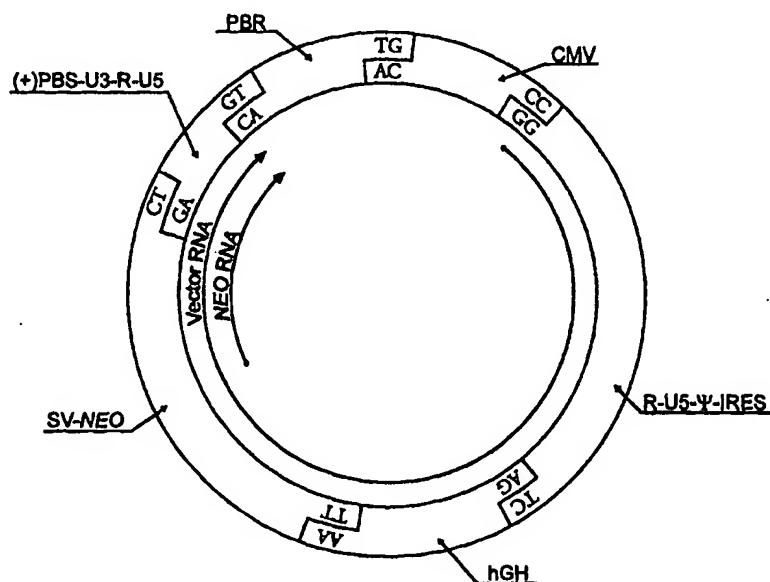
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(57) Abstract

The invention relates to a method for directing the self-assembly of a gene or gene assembly having three and preferably six or more fragments in a directionally and spatially ordered fashion to produce a gene, gene vector or large nucleic acid molecule. The method can be used to create libraries, such as combinatorial libraries. In another embodiment of the invention a vector is described for the incorporation and screening of endogenous mouse promoter elements for the identification of cell-specific promoters.

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SELF-ASSEMBLING GENES, VECTORS AND USES THEREOF

Field of the Invention

5 This invention relates to the construction and usage of synthetic genes for genetic engineering and gene therapy.

Background of the invention

 This application claims the benefit of a provisional application U.S. Serial No.
10 60/070,910, filed on February 28, 1997, entitled "Self-Assembling Genes."

 Recombination at the genetic level is important for generating diversity and adaptive change within genomes of virtually all organisms. Recombinant DNA technology is based upon simple 'cut-and-paste' methods for manipulating nucleic acid molecules *in vitro*. The pieces of genetic material, or DNA are first digested with a restriction endonuclease
15 enzyme which recognizes specific sequences within the DNA. After preparation of two or more pieces of DNA, the ends of the DNA are further manipulated, if necessary, to make them compatible for ligation or joining together. DNA ligase, together with adenosine triphosphate (ATP) is added to the genes, ligating them back together. The genetic assembly containing an origin of DNA replication and a selectable gene is then inserted into a living
20 cell, is grown up, and is positively selected to yield a pure culture capable of providing high yields of individual recombinant DNA molecules, or their products such as RNA or protein.

 Significant improvements have been made to this technology over the last two and a half decades. Numerous enzymes, end-linkers and adapter molecules have been made commercially available, which facilitate in the construction of recombinant DNA molecules.
25 By using two restriction enzymes with different single-stranded termini or blunt ends, it is possible to directionally assemble genes (forced cloning). This reduces the amount of screening required to determine orientation. Procedures have been automated for synthesis of single-stranded gene fragments up to 200 or more nucleotides in length by means of phosphoramidite chemistry, and the instrumentation is readily available through Applied
30 Biosystems, Inc., Foster City, CA. Such single-stranded fragments can be joined by annealing overlapping complimentary phosphorylated strands, and by enzymatically filling in the ends with DNA polymerase and DNA precursors. In this way, multiple, overlapping, single-stranded fragments can be assembled into a larger, double-stranded superstructure.

Whole genes have been synthesized by similar methods. However, it becomes increasingly difficult to use synthetic DNA strands when making genes larger than approximately one kilobase. Using gene amplification methods (e.g. polymerase chain reaction (PCR), Mullis *et al.*, U.S. Patent 4,683,195), together with synthetic oligonucleotides, it is possible to make
5 biologically active, synthetic retro-vectors that are capable of RNA transcription, reverse-transcription, viral packaging, and integration into genomic DNA (see for example, Hodgson, WO94/20608). Hodgson, *supra*, also disclosed methods for cloning of transcriptional promoters into such a vector using traditional recombinant DNA technology.

Modified restriction enzyme sites, linkers, and adapters can change the
10 primary or secondary structure of complex nucleic acid sequences thereby altering or obliterating a desired biological activity. For example, small mutations can drastically modify transcriptional promoters or change the reading frame of coding DNA. A logical goal of vectorology is to make exact constructs, without need of fortuitous restriction sites, adapters, or linkers.

15 Restriction endonucleases can be grouped based on similar characteristics. In general there are three major types or classes: I, II (including IIS) and III. Class I enzymes cuts at a somewhat random site from the enzyme recognition sites (see Old and Primrose, 1994. *Principles of Gene Manipulation*. Blackwell Sciences, Inc., Cambridge, MA, p.24). Most enzymes used in molecular biology are type II enzymes. These enzymes recognize a
20 particular target sequence (i.e., restriction endonuclease recognition site) and break the polynucleotide chains within or near to the recognition site. The type II recognition sequences are continuous or interrupted. Class IIS enzymes (i.e., type IIS enzymes) have asymmetric recognition sequences. Cleavage occurs at a distance from the recognition site.

These enzymes have been reviewed by Szybalski *et al.* *Gene* 100:13-26, 1991. Class
25 III restriction enzymes are rare and are not commonly used in molecular biology.

U.S. Patent No. 4,293,652 employed a linker with a class IIS enzyme recognition sequence to permit synthesized DNA to be inserted into a vector without disturbing a recognition sequence. Brousseau *et al.* (*Gene* 17:279-289, 1982) and Urdea *et al.* (*Proc. Natl. Acad. Sci. USA* 80:7461-7465, 1983) disclose the use of class IIS enzymes for
30 the production of vectors to produce recombinant insulin and epidermal growth factor respectively. Mandecki *et al.* described a method for making synthetic genes by cloning small oligonucleotides using a vector (*Gene* 68:101-107, 1988). Expansion of a population of

oligonucleotides required synthesis, cloning excision and fragment purification. The oligonucleotides were used to create a complete plasmid.

Lebedenko et al. (*Nucl. Acids Res.* 19(24):6757-6771) illustrated the class IIS enzymes and PCR for precisely joining 3 nucleic acid molecules for convention sub-cloning using BamHI. Tomic *et al.* (*Nucleic Acids Res.*, 18:1656, 1990), reported a method for site-directed mutagenesis using the polymerase chain reaction and class IIS enzymes to join two nucleic acid molecules. Two overlapping PCR primers were used where the primers included class IIS recognition sites. The primers included a region of complementarity to the template DNA and include one to a few site-directed mutations. Stemmer et al. (U.S. Patent No. 5,514,568) employed overlapping primers with class IIS enzymes to amplify a plasmid and to introduce specific mutations into DNA leaving all other positions unaltered.

There remains a need for the ordering and assembly of complex genes to overcome the problems associated with sequential sub-cloning such as multiple purification steps, the potential for sample loss, and the like. Moreover there is a need for eliminating the use of prokaryotic hosts and for minimizing or avoiding the risks associated with bacterial contamination resulting from the use of bacteria as intermediaries in the cloning process. Further, there remains a need for efficient methods to assemble large nucleic acid molecules or many-fragmented nucleic acid assemblies with precision.

Brief Description of the Figures

Fig. 1A. provides one schematic of six double stranded DNA fragments, each terminus comprising a unique overhanging two-nucleotide sequence complementary to only one other terminus

Fig. 1B. illustrates a three-piece ligation where 100% of the clones tested contained the predicted fragment order and desired fragment orientation.

Fig. 2. illustrates the use of a class IIS restriction endonuclease (as one example, *Bpm1*), restriction endonuclease recognition site and the selection of cohesive overhanging ends.

Fig. 3A. illustrates an exemplary retrotransposon-derived vector including a murine VL30 LTR (NLV-3) and packaging signal, an internal ribosome entry site (IRES) from encephalomyocarditis virus (EMCV), a gene encoding a green fluorescent protein (GFP), additional internal VL30 sequences (solid bar), SV40 early region promoter and Tn5

aminoglycosidase phosphotransferase (neo) gene, PBR322 plasmid origin of replication and a plus-strand primer binding site (VL30). An exemplary vector sequence is provided as VLBPNG (SEQ ID NO:1). Fig 3B is an illustration of an LTR with the insertion of a U3 (transcriptional promoter) region rescued by reverse transcriptase-polymerase chain reaction (RT-PCR). The promoter is amplified from the RNA of a cell expressing the VL30 U3 region. Complementary overhanging ends are created using class IIS restriction endonuclease digestion sites within the LTR and within the promoter. Fig. 3C provides the linear structure of a VL30 RNA transcript from a mouse cell with a U3 region near the 3'-terminus of the RNA molecule. PCR primers include a class IIS enzyme recognition site to amplify the U3 region from the RNA resulting in a double stranded DNA molecule. Cleavage with a class IIS enzyme (here *BpmI*), results in a double-stranded DNA molecule with end complementary to a site in the vector of Fig. 3A.

Fig. 4A. is a schematic illustrating steps for assembling a combinatorial library of *cis*- or *trans*-acting nucleic acid sequences for assembly and screening, useful for the rescue of biologically active species. Fig. 4b is a diagram of a U3 (transcriptional enhancer and promoter region of an LTR illustrating several sub-divisions of the transcriptional control region, including a distal enhancer region, an enhancer repeat region, a medial promoter and a proximal promoter. These regions have been described for other vectors in Hodgson et al. (1996. "Construction, Transmission and Expression of Synthetic VL30 Vectors" in Hodgson ed. *Retro-vectors for Human Gene Therapy*. RG Landes Company, Austin TX). Segments of these regions are amplified using primers for highly conserved sequences. Highly conserved sequences are determine based on a comparison of known VL30 sequences such as provided in Fig. 4.2 of Hodgson, 1996, *infra*). The parts are joined by annealing and ligation to provide an ordered assembly. Each construct is an allele or a representative of allelic variation in the combinatorial library.

Fig. 5 discloses two transcriptional promoters that have been rescued from mouse VL30 RNA sequences isolated from a mouse T-helper cell library. These promoters were assembled into a vector and introduced into retroviral helper cells and packaged into recombinant retrovirus for introduction into human T-cells. After transduction to human T cells, a β -galactosidase reporter gene was expressed from the T cell-derived promoters.

Fig. 6 discloses 10 biologically active mouse VL30 promoters obtained from mouse liver RNA. These promoters were introduced into the vector of SEQ ID NO:1. The vectors

were introduced into retroviral helper cells and then packaged into retrovirus where they were introduced into human liver cells. The cells expressed the green fluorescent protein.

Fig. 7 illustrates a similarity plot of nucleotide sequences found in VL30 U3 regions.

Fig. 8 illustrates a retro-vector comprising six double-stranded DNA fragments that were self-assembled into a circular structure using unique overlapping termini created using class IIS restriction endonucleases. Three templates and twelve primers were used in conjunction with three class IIS enzymes to make the six fragments that were ligated in a single step. The vector was efficiently self-assembled and was effectively transmitted by both DNA transfection as well as by retroviral transduction of the self-assembled DNA, without molecular cloning through a prokaryotic host (see Example 2).

BRIEF SUMMARY OF THE INVENTION

The invention described herein provides seamless, directional, ordered construction of complex DNA molecules, vectors and libraries. More particularly, it enables gene constructs to be assembled with greater efficiency and precision, and it enables multiple gene fragments to be assembled in the correct order and orientation without disturbing the internal structure of the gene. The method utilizes *in vitro* assembly of nucleic acid fragments and relies upon the unusual ability of certain enzymes to digest nucleic acid molecules at pre-determined sites without disrupting the structure of the gene. It is especially useful for the construction of genetic vectors for gene therapy or genetic engineering of cells and organisms. A particular application of the invention is in combinatorial, or evolutionary genetics, where it enables a large number of non-random, self-assembled constructs to be screened simultaneously for function.

In a preferred embodiment of this invention, the invention relates to a method method for assembling a gene or gene vector comprising the steps of: a) designing at least 6 primers to produce to amplify at least three fragments in at least three separate polymerase chain reactions wherein each primer comprises at least one predetermined restriction endonuclease recognition site that recognizes a restriction endonuclease that cleaves at a distance from the recognition site, a sequence complementary to a template nucleic acid for amplification, and bases positioned at the restriction endonuclease cleavage site that are selected to be complementary to only one other overhanging created from enzymatic cleavage of the fragments; b) combining the primers with template nucleic acid and performing the

polymerase chain reaction to produce multiple copies of an amplified template fragment incorporating the restriction endonuclease recognition site; c) digesting the amplified template fragments with one or more restriction endonucleases that recognize the restriction endonuclease recognition site of the primers to create overhanging termini wherein each
5 overhanging termini is complementary to only one other overhanging termini on another fragment; and d) combining the amplified and digested template fragments in a ligation reaction to produce a directionally ordered gene, nucleic acid fragment or gene vector.

In a preferred aspect of this embodiment, the restriction endonuclease is at least one class IIS restriction endonuclease and preferably, the class IIS restriction
10 endonuclease is selected from the group consisting of: *AlwI*, *Alw26I*, *BbsI*, *BbvI*, *BbvII*, *BpmI*, *BsmAI*, *BsmI*, *BsmBI*, *BspMI*, *BsrI*, *BsrDI*, *Eco57I*, *EarI*, *FokI*, *GsuI*, *HgaI*, *HphI*, *MboII*, *MnII*, *PleI*, *SapI*, *SfaNI*, *TaqII*, *Tth111III*. Still more preferably, class II restriction endonuclease recognition sites (to be distinguished from class IIS restriction endonuclease recognition sites), linkers, or adapters are not used to create the gene or gene vector. In one
15 embodiment, the product of the ligation reaction is introduced into prokaryotic or eukaryotic cells. Preferably, at least one template nucleic acid sequence is chosen from the group consisting of: transcriptional regulatory sequences; genetic vectors; introns and/or exons; viral encapsidation sequences; integration signals intended for introducing nucleic acid molecules into other nucleic acid molecules; retrotransposon(s); VL30 elements; or multiple
20 allelic forms of a sequence.

In another preferred aspect of this embodiment, the method is used to generate combinatorial libraries of a target sequence. Preferably, the target sequence is part or all of a gene. In one embodiment, the gene encodes a protein. In one embodiment, the primers amplify allelic variants of part or all of a gene.

25 In still another preferred aspect of this embodiment, the product of the ligation reaction is passed between eukaryotic cells using a virus particle, by cell fusion, or by transfection. Preferably the product of the ligation reaction is not introduced into prokaryotic cells. Moreover, the method further comprises combining at least one screening or selection step to select the products of the ligation reaction. In one embodiment, the product of the
30 ligation reaction is mutated during passage in cells in order to generate genetic diversity and preferably the product of the ligation reaction is mutated by homologous recombination during passage in cells.

In another aspect of this embodiment, the method is used to isolate and identify regulatory sequences from a cell. In another aspect of this embodiment, cells containing the product of the ligation reaction are selected for enhanced biological activity. Preferably, the cells containing the product of the ligation reaction are selected for tissue-specific, hormone-specific or developmental-specific gene expression. Also preferably, the ligation reaction is a circularized gene vector.

In another embodiment of this invention, the invention relates to a nucleic acid primer having a 5' and a 3' end to amplify a nucleic acid fragment for the ligation of at least two fragments comprising: a restriction endonuclease recognition site that recognizes a restriction endonuclease, wherein the restriction endonuclease cleaves at a distance from the recognition site and creates overhanging termini; a sequence complementary to a template sequence to be amplified to produce the nucleic acid fragment; at least two nucleic acid bases positioned at the restriction endonuclease cleavage site and that form an overhanging terminus after cleavage by the restriction endonuclease, wherein the at least two nucleic acid bases are selected to be complementary to only one other overhanging terminus on another fragment of the ligation; and an affinity handle on the 5' end of the primer. Preferably the primer further comprises an anchor to provide stability to the restriction enzyme at the restriction enzyme recognition site.

In yet another embodiment of this invention, the invention relates to a method for isolating and identifying promoters comprising the steps of: a) obtaining a vector comprising at least a portion of a promoter region from a retrovirus transposon LTR and having two non-complementary overhanging termini; b) designing at least two PCR primers to amplify at least one region of a retrovirus transposon LTR from template nucleic acid to produce at least one nucleic acid fragment wherein each primer comprises at least one predetermined restriction endonuclease recognition site that recognizes a restriction endonuclease that cleaves at a distance from the recognition site, a sequence complementary to a template sequence from a retrovirus transposon, and bases positioned at the restriction endonuclease cleavage site that are selected to be complementary to only one other overhanging terminus of the vector wherein the restriction endonuclease cleavage site is created from enzymatic cleavage of the fragments; b) combining the primers with template nucleic acid and performing a polymerase chain reaction to produce multiple copies of an amplified template fragment incorporating the restriction endonuclease recognition site; c)

digesting the amplified template fragments with one or more restriction endonuclease that recognize the restriction endonuclease recognition site of the primer to create overhanging termini; and combining the amplified and digested template fragment in a ligation reaction with the vector to produce a gene vector with an intact LTR sequence. In one embodiment of this aspect of the invention, the template nucleic acid is DNA or RNA. In another embodiment of this aspect of the invention, the method further comprises the step of sequencing the insert to identify the promoter sequence. In one embodiment promoter sequences of SEQ ID NOS:1-13 identified using the methods of claim.

10 Detailed Description of the Invention

In one embodiment of this invention, the invention relates to the seamless, oriented self-assembly of at least three DNA fragments having overlapping unique cohesive ends generated by the enzymatic cleavage of at least one restriction endonuclease that is capable of cleaving at a site distant to the restriction enzyme recognition site. Preferably the restriction endonucleases employed in this invention are class IIS restriction endonucleases. These enzymes recognize a predetermined group of nucleotides and cleave at a distance characteristic of the particular endonuclease from the recognition site. The term "unique cohesive ends" is used herein to refer to the notion that the cleavage site for the endonucleases of this invention can be manipulated to produce overhanging ends with unique termini selected by the investigator. The term "complementary" as used herein in reference to the overhanging ends of the fragments of this invention refers to standard complementarity recognized in the field of molecular biology. For example, the nucleotides sequence 5'-TAG-3' is said to be complementary to the nucleotide sequence 5'-CTA-3'. The term "PCR" is used generally to refer to the polymerase chain reaction and its variations, including RT-PCR as well as other gene amplification techniques employing primers.

In a first step for practicing one embodiment of this invention, a series of at least three overlapping fragments are created through the selection and creation of primers incorporating at least one class IIS restriction enzyme recognition sequence. The oligonucleotide primers of this invention are designed to amplify one or more nucleic acid fragments and comprise a sequence complementary to a target sequence for gene amplification, a recognition sequence for a restriction endonuclease that cleaves DNA at a distance from the recognition sequence (such as a class IIS restriction enzyme) and bases

positioned at the restriction endonuclease cleavage site that are preferably unique and complementary to only one other overhanging termini in the annealing/ligation reaction that generates the complex nucleic acid molecules. Optionally, the primers of this invention can include an "affinity handle for cleanup" at the 5' end. These sequences can be of any length, 5 preferably at least about 6 bp and the sequences extend the primer in the 5' direction from the restriction enzyme recognition site. This extra length gives many enzymes greater stability and improved activity. In addition, the sequence can be used for recognition and removal of the ends of the primers (either undigested fragments or digested ends of primers) using complementary nucleotide sequences bound to a solid support (such as cellulose, 10 nitrocellulose or silica). Incubation with, or passage over a column or support containing the complementary sequences can be used to remove the tags by allowing them to anneal or hybridize. The nucleic acid can then be eluted from the column. Adapters can also be used in this invention. For purposes of this invention, adapters refer to double stranded fragments containing an enzyme recognition site, according to this invention. The adapters are ligated 15 to double stranded DNA molecules, creating a fragment analogous to a PCR fragment with similar sites derived from a primer. The primers or adapters can be prepared using a number of methods for synthesizing oligonucleotides known in the art. For example instruments for producing oligonucleotides are available from Applied Biosystems, Inc., Foster City, CA.

In one example, for the design of an oligonucleotide primer for use in this 20 invention, the particular complementary bases that will form the site for hybridization of the primer to template (i.e., target DNA or RNA) are selected. A restriction endonuclease recognition site is selected followed by a number of nucleotides to be positioned between the recognition site and the cleavage site. The nucleotides of the cleavage site are selected to include overhanging regions formed from the restriction endonuclease cleavage that are 25 complementary to the overhanging regions of an adjacent fragment in the annealing/ligation reaction.

The length of the primer used in this invention can vary, but preferably the primer length is up to about 80 bases and preferably up to about 50 bases. In addition the primers are preferably at least about 15 bases in length and preferably at least about 25 bases 30 in length. The 5' region of the primer contains preferably at least about 6, preferably at least about 10 and still more preferably at least about 16-18 bases that are not complementary to the template DNA or RNA. Further, the primer incorporates a restriction endonuclease

recognition site preferably 5' to the region of complementarity and a restriction endonuclease digestion site preferably 5' to the region of complementarity or within the region of complementarity. There are a variety of restriction endonucleases that cleave at a distance from the restriction endonuclease recognition site of a DNA strand and a variety of enzymes that are commercially available from New England Biolabs are provided in Table 1.

Table 1. Restriction endonucleases useful in the construction of self-assembling genes

Enzyme:	Site size (bp):	Distance to overlap:	Size of overlap:	Overlap type:
<i>Alw26 I</i>	5	1-5bp	4bp	5'-Overhang
<i>BbsI</i>	6	2-6bp	4bp	5'-overhang
<i>BpmI</i>	6	16-14bp	2bp	3'-overhang
<i>BsmBI</i>	6	1-5bp	4bp	5'-overhang
<i>BspMI</i>	6	4-8bp	4bp	5'-overhang
<i>BsrDI</i>	6	0-2bp	2bp	3'-overhang
<i>Eco57I</i>	6	16-14bp	2bp	3'-overhang
<i>FokI</i>	5	9-13bp	4bp	5'-overhang
<i>HgaI</i>	5	5-10bp	5bp	5'-overhang
<i>HphI</i>	5	8-7bp	1bp	3'-overhang
<i>MnII</i>	5	7-6bp	1bp	3'-overhang
<i>PleI</i>	5	4-5bp	1bp	5'-overhang
<i>SapI</i>	7	1-4bp	3bp	5'-overhang
<i>SfaNI</i>	5	5-9bp	4bp	5'-overhang

In addition to the enzymes provided in Table 1, other restriction endonucleases that cleave at a distance from their restriction endonuclease recognition site include, but are not limited to, *AlwI*, *BbsI*, *BbvI*, *BbvII*, *BsmAI*, *BsmI*, *BsrI*, *EarI*, *GsuI*, *MboII*, *TaqII*, *Tth111II* and their respective isoschizomers. These and other enzymes are known in the art and many are available from other manufacturers. The primers can be prepared to produce either 5'-overlapping ends or 3'-overlapping ends, as long as they are both are either 5'-overlapping ends or 3'-overlapping ends and are complementary to one other set of overlapping ends.

In the case of *BpmI* (see Example 1), the enzyme digests asymmetrically, 14-16 bp from the 3'-nucleotide of the recognition site. The resulting cleavage has a 3'-overhanging end of 2 bp. A second primer is then designed with a complementary

overhanging end, and it is used to generate the adjoining fragment terminus. At the opposite ends of the two fragments that are to be joined, similar complementary, overhanging ends are designed.

The oligonucleotides are then combined with template nucleic acid (either
5 DNA or RNA, e.g., such as for reverse transcriptase polymerase chain reaction (RT-PCR))
containing bases complementary to at least a 3' portion of the primers (also referred to herein
as "templates"). In one embodiment, the fragments are gene-amplified by PCR, RT-PCR or
another gene amplification process using established PCR protocols such as those provided
with PCR amplification kits, including those available from Perkin-Elmer Corp. (Emeryville,
10 California). Preferably, the PCR products are analyzed by electrophoresis on a gel, such as
an agarose gel and still more preferably the fragments of the predicted size are purified free of
excess primers and small byproducts (such as by purification through a small column, such as
a Qiagen™ column (Qiagen, Valencia, CA)). Following amplification or purification, the
fragments are digested with the restriction endonuclease recognizing the restriction
15 endonuclease recognition site in the primers. The digested fragments are then purified from
the digested ends of the primers, preferably by preparative agarose gel electrophoresis. The
fragments are combined, annealed and are ligated using standard hybridization and ligation
conditions known for cloning (see Ausubel et al., *Current Protocols in Molecular Biology*,
John Wiley & Sons, 1994).

20 Fig. 1A illustrates an example of a self-assembling gene construct (SEQ ID
NO:1) comprising six fragments, each having unique overhanging dinucleotide ends. In this
example, the ends of the fragments prepared by the methods of this invention are constructed
using primers that include *BpmI* restriction endonuclease recognition sites. It will be
understood by those of ordinary skill in the art that one or more other restriction
25 endonucleases (such as those of Table 1) could similarly be used for the self-assembling
product of Fig. 1A. In a preferred embodiment, the primers were created as described above
and preferably the 3' ends of the primers are non-palindromic (i.e., non self-complementary)
to prevent self-annealing of such fragments. Each fragment in this example preferably joins to
only one other dinucleotide overhang in the annealing/ligation mixture, assuring ligation only
30 to the intended fragment partner. An advantage of this strategy is that the formation of
concatamers or multimers is minimal. The restriction endonuclease site is removed by

digestion with the restriction endonuclease, leaving the junction free of the extra DNA sequences associated with the site.

Using a single restriction endonuclease with a dinucleotide overhang (for example, using the enzyme *BpmI*) up to six pieces of genetic material can be joined together in a linear or circular form (such as a vector) without the need to perform sub-cloning procedures or detailed analysis of individual products because six unique combinations of dinucleotide overhangs create a directional clone with extremely high fidelity. With enzymes digesting single-base overlaps, only two fragments can be joined with positional and directional precision. With enzymes digesting three-base overlaps, $4^3/2$, or 32 fragments can be so joined in the correct order and orientation. Therefore, this invention also relates to the use of restriction endonuclease recognition sites that facilitate cleavage by restriction endonucleases with three-base overlaps and self-assembly gene constructs including 32 fragments. Alternatively, a combination of restriction endonuclease recognition sites for use with a combination of restriction enzymes that create two-base or three-base overlaps can be used. Each enzyme has its characteristic limits to self-assembly imposed by the size of the overlap. For example, there are sixteen dinucleotides, therefore *BpmI* fragments (which have two dinucleotide ends each) are limited to eight for the purpose of self-assembly; therefore in another embodiment of this invention an assembly comprising eight fragments is contemplated. However, four of the sixteen dinucleotides are palindromes. Use of these palindromic dinucleotides can create some infidelity in the annealing/ligation reaction. The enzyme *HgaI* has a five base overlap, and there are 1,024 pentanucleotide combinations, permitting 512 fragments to be ligated together directionally and in order (no palindromes). The fragments to be joined at a particular place are designed to have their cut sites aligned, so that the overlapping region fits together. In some cases, the target sequences will contain natural restriction endonuclease recognition sites for the enzyme that is being used, such as one or more internal *BpmI* sites. These sites have the potential to self-religate during vector or gene construction or they can be by passed by using a substitute enzyme in the primers (for example, *Eco 571* can substitute for *BpmI*). Alternatively, these sites can be removed by site-directed mutagenesis after consideration to the consequences of the mutagenized sequence to the gene or vector.

In addition to class IIS enzymes, class II restriction endonucleases can be used. These enzymes have intrinsic methylation activity that affects the outcome in either a

negative or a positive way, depending on the purpose for which it is used. In a preferred embodiment, the methylation activity of class II enzymes is ablated by mutation or by genetic engineering to convert the enzyme to an effective class IIS enzyme to expand the repertoire of useful enzymes for this invention.

5 In another aspect of this invention, the primer design and target fragment sequence selection can be automated (see Example 5) using a computer to assist in the selection of unique overhanging ends that have complementarity only to the overhanging end of an adjacent fragment.

 Therefore, this invention permits high-fidelity annealing and ligation of six or
10 more fragments with unique overhanging termini complementary to a single other overhanging termini. Any multitude of combinations can be created by combining the type of overhanging termini that can be created. Moreover, if one is willing to sacrifice the fidelity of the reaction, a variety of combinations can be used to anneal a variety of fragment numbers. In these cases, some selection may be necessary, such as size selection of the
15 resulting fragment based on electrophoretic migration or restriction endonuclease profiling, both methods well known to those of ordinary skill in the art

 It is also necessary to have a high per-step efficiency (e.g., each step in the process is performed with an efficiency of at least 80%) to effectively ligate large numbers of fragments without error. Where large numbers of fragments are used, the purity of the
20 fragments becomes important. This means that for large numbers of fragments, the digested DNA fragments for annealing and ligation should be substantially pure. If undigested fragments, digested ends of primers, degraded or partially degraded molecules are present, they can decrease the purity and affect the fidelity of the product. Therefore, it is particularly desirable to ensure complete digestion of both ends of each fragment and to remove all of the
25 digested ends from the fragments prior to including the fragments in an annealing and ligation reaction. The use of Qiagen columns for oligonucleotide removal prior to digestion is generally sufficient to permit efficient digestion of the fragments. Agarose gel isolation is desirable after digestion particularly where the product contains some fragments that do not appear to be full length. The use of an analytical gel before and after digestion helps in
30 determining whether both oligonucleotide tags have been removed. The isolation of fragments from agarose gels preferably avoids the use of ultraviolet light and exposure of the

DNA to ethidium bromide is also preferably avoided. These methods can be avoided by running replicate lanes and staining only a portion of the gel.

The fragments and vector are then digested to yield fully complementary ends, and the fragments are preferably again purified, as described above (such as through a Qiagen column or by gel isolation). The purified fragments are ligated together in a test tube, under standard conditions, such as using bacteriophage T4 DNA ligase and ATP. Preferred ligations include at least 20µg/ml total DNA concentration in the ligation mix to favor intermolecular interactions, and an equimolar ratio of fragments to be ligated. Where a prokaryotic intermediary is used, the ligated assemblage is transformed into a bacterium, such as an *E. coli* host, and the colonies are: selected with a drug (such as an ampicillin, tetracycline, or kanamycin marker). The colonies can then be selected either by individually selecting colonies or growing a mass culture, such as where a vector library has been created. Restriction enzyme analysis can be used to determine the identity of individual constructs or to assess the validation of the combination of plasmids. The plasmids can then be grown up and used as needed.

In one embodiment of this invention, at least a portion of a vector is used as one of the fragments for the ligation of at least three fragments according to this invention. In one example, where a vector is used as one of the starting fragments, two restriction endonuclease recognition sites recognizing an enzyme that cleaves at a distance from the recognition site, such as at least one *BpmI* site, can also be introduced into the vector. This permits the vector to be digested with the restriction endonuclease to produce a product having ends complementary to two ends of the insert DNA fragments. The vector can be made by amplifying a plasmid or portion thereof using the primers of this invention. Thus, the vector can also be constructed to include a variety of restriction endonuclease recognition sites using a variety of restriction endonucleases, including a variety of class II restriction endonucleases. In some cases, the target fragments for amplification will contain natural restriction endonuclease recognition sites for the enzyme that is being used for the self-assembly, such as for example, a fragment that includes one or more internal *BpmI* sites. Care should be taken either to utilize the complementarity of the naturally occurring site to reform the fragment as it originally existed or to eliminate the restriction endonuclease recognition site using, for example, site-directed mutagenesis. Preferably, the restriction endonuclease recognition site is substituted for a different enzyme (in the case of *BpmI*,

substituting *Eco57I* or *BsrDI*) that has an equivalent structure at its ends. Two or more fragments of insert or two or more fragments of vector with at least one insert are amplified using primers according to this invention.

The exemplary enzyme, *BpmI* digests DNA 14-16 base pairs (bp) from the 3'-
5 nucleotide of the recognition sequence (RS). Thus, by placing the RS exactly 14-16 bp from the desired dinucleotide cut site, the practitioner tags the dinucleotide for ligation with another dinucleotide that is exactly complementary to it. Such a complementary dinucleotide can be inserted by using the same enzyme and RS to make another fragment which fits the first exactly, as illustrated in Fig. 1. Because there are sixteen possible dinucleotide
10 combinations (including twelve combinations that do not have palindromic ends), it is possible to create up to six fragments with unique dinucleotides, and it is also possible to join them all together in a predetermined order and orientation (Fig 1A). In addition, the palindromic sequences (such as AT, CG, TA, and GC) could also be used, although inefficiency and incorrect ligation will result from the self-complimentarity of these
15 sequences. It is furthermore possible and desirable to have three or more fragments joined in this way, such that the construct is circular as in Fig. 1, comprising a vector that may be grown in a bacterial and/or eukaryotic host cell. If the genetic construct is to be used as a vector, the vector should be designed to include a proper origin of replication to enable it to replicate in a particular cell. For example, a prokaryotic origin of replication such as a
20 coliform plasmid origin of replication enables circular DNAs to be propagated in *E. coli* host cells. It is desirable to have at least one selectable marker, such as a neomycin marker that enables recovery of the clone through a selection process. It is also desirable, but not essential, to have two or more selectable genetic elements, to permit dual selection. For example, if one of the fragments contains a prokaryotic plasmid origin of replication, and
25 another fragment contains a selectable marker, then the two fragments are both selectable, since the construct will grow in prokaryotic cells in the presence of a selection drug (such as ampicillin) only when both fragments are present. Drug selection can be combined with the methods of directed self-assembly to assure a high percentage of correct products. Because of the unique complementarity of the fragments, each contributes a selectable element that
30 leads to recovery of a high percentage of correct products.

For prokaryotic vector construction, at least one fragment should contain a prokaryotic origin of replication and one fragment should contain a drug resistance marker

gene. However, an advantage of the methods of this invention is that the construct can be introduced directly into eukaryotic cells. Here no plasmid origin of replication is necessary and no prokaryotic selectable marker or other prokaryotic nucleic acid sequence is necessary. In cases where the vector is subject to regulatory approval or where optimal gene function is necessary, it may be undesirable to include prokaryotic sequences, such as extraneous plasmids or expressed prokaryotic fragments particularly if the sequences contain immunostimulatory sites that can lead to activation of the intracellular immune system and inactivation of a gene product (see Krieg et al., *J. Lab. Clin. Med.*, 128:128-133, 1996) or to avoid risks of endotoxin contamination. Moreover, the use of self-assembled product, according to the methods of this invention saves labor and time involved in the screening process.

Thus, in a preferred embodiment of the invention, the nucleic acid fragments are self-assembled *in vitro*, and are transferred directly into eukaryotic cells, by transfection, injection, or other methods known in the art. In one embodiment the cells receiving the assembled product of this invention are helper cells for recombinant virus assembly (including, but not limited to retroviral helper cells for retroviral or retrotransposon vectors, adenovirus helper cells for adenovirus vectors or herpes simplex virus helper cells for herpes simplex vectors). Alternatively, the assembled product can be introduced into cells along with a helper virus or the assembled product can be introduced into target cells for direct expression. The assembled product can be a vector, a minichromosome vector, a portion of a chromosome, or the like. In the preferred case of a retroviral vector, the genes are first transfected into a first helper cell line (such as ecotropic helper cells, GP+E86 (Markowitz et al. *J. Virol.* 862:1120-1124, 1988). The retrovirus-containing supernatant from these cells is then filtered (0.45mm Nalgene filters) preferably 48-72 hours after transfection and the filtrate is transferred to a second complementation retroviral helper cell line (such as PA317 retroviral helper cells, Miller et al., *Mol. Cell. Biol.* 6:2895-2902, 1986). After an additional 48 h, the second helper cell line is selected with the marker drug (such as the drug G418 for the selectable neomycin (neo) marker gene), until only drug-resistant cells remain. These cells contain stably integrated vectors that can be used to repeatedly transduce human cells. Advantageously, in the case of adenovirus vectors or other large eukaryotic -derived vectors including eukaryotic virus-derived vectors, it may be impossible to propagate them in prokaryotic hosts. The gene self-assembly method of the instant invention provides an

alternative to *in vitro* recombination method of gene construction by permitting large constructs to be constructed.

One advantage of introducing the assembled product of this invention into a helper cell line to produce recombinant virus for the introduction of a gene or nucleic acid complex into a cell is that the assembled product will be auto-selected by the cells during the packaging process. Therefore, even where the overhanging termini have palindromic sequences, where there is more than one (but preferably less than four) unique complementary matches for a particular overhanging termini, or where concatamers have formed, only the correct or functional assembled products are expressed, transmitted, and assembled into virus. When the virus is then introduced into cells, the use of a reporter gene or another selectable marker provides yet a second layer of security for the selection of cells containing a properly assembled construct. For example, where a retrovirus helper cell line is used to produce a recombinant retrovirus containing the product of this invention (for retrovirus, RNA transcribed from the DNA product of the invention becomes packaged into the virus particle), a retrovirus-derived vector is transcribed as RNA and transmitted by packaging the RNA in a retrovirus particle. In order to be properly transmitted as a virus, the construct must be: 1) transcribed as RNA in a vector producer cell; 2) packaged into viral particles; 3) reverse transcribed into double-stranded DNA (in the recipient cell); and 4) integrated into the host chromosome. Each of these steps requires specific *cis*-acting sequences that must be correctly positioned within the vector. Thus, passage via retrovirus (or by other virus) is a means of auto-selection for the essential sequences.

In one application of the methods of this invention, the methods are used to rescue expressed sequences from RNA, or genomic sequences from cell DNA without disrupting the promoter sequences. Cellular transcriptional promoters are typically difficult to identify and isolate because they are generally not included in the RNA molecule and often extend over a considerable distance in a chromosome. One application of this invention relates to a promoter rescue technique that permits the entire promoter, or a fragment of a promoter to be isolated and cloned directly in to an expression vector without disruption of the flanking sequences. Promoter rescue techniques are known and include WO 94/20608 to Hodgson.

In a preferred embodiment of the invention, transcriptional promoters are cloned in a transcriptionally active manner for the selection and identification of new and/or

of tissue or cell-specific promoters enabling them to be used, selected, or screened for activity directly. For example, Fig. 3 illustrates one example of the formation of a vector for the incorporation of promoter sequences and the ultimate identification of those sequences using an exemplary plasmid VLBPGN (SEQ ID NO:1) as provided in Example 3, with *Bpm1* sites
5 located within the locus of a retrotransposon (VL30) long terminal repeat (LTR). These methods preserve the structure and functionality of transcription factor response elements. The characteristic secondary structure of the LTR RNA remains very similar to the original LTR from which the promoter was rescued, thus preserving the important features of the original RNA/DNA molecule. Those of ordinary skill in the art will recognize that any of a
10 variety of primers can be used with a variety of vectors and that the constructs of Figs 2 and 3 are exemplary and not limiting.

Fig. 2 illustrates the primers used to amplify the promoter insert (identified at a and c in Fig.2), and the insert region of the LTR (boxed), both of which can be digested at the same nucleotide position with *Bpm1*, to ensure a proper and seamless fit. In this example,
15 after digestion of the vector, the two *Bpm1* sites leave non-complementary ends (a 3'-CC overhang on one end, and a 3'-GC overhang on the other). Thus, the ends will not efficiently anneal or ligate to one another. However, the complementary termini of the insert serves as linkage, enabling the plasmid to be completed by ligation.

In the example illustrated in Fig. 2, the terminus on the 3'-side (GC) is
20 palindromic. Palindromic termini are self-complementary and can therefore ligate to themselves or to an identical terminus facing the opposite way (forming concatamers in the opposite direction). Despite the presence of palindromic termini and despite the potential for reduced fidelity in the self-assembling process, a large percentage of clones obtained by inserting promoter sequences into VLBPGN were assembled correctly (20/23). These levels
25 are reduced somewhat when three or more fragments are combined for self-assembly, according to this invention and preferably, the use of palindromic termini are avoided when even numbers of nucleotides are exposed as overhanging termini because with even numbers of nucleotides there is an axis of symmetry. As noted above, where five base overhangs are used there are 1024 possible combinations of five nucleotides $[(4)^5]$, yet none of them is
30 palindromic.

The vector of Fig. 3 is an example of a particular type of vector that is known as a retrotransposon vector. Retrotransposon vectors are described and reviewed in Hodgson

et al., 1996 *Retro-Vectors for Human Gene Therapy*. RG Landes Company, Austin TX, chapter 5 and see US Patent 5,354,674 to Hodgson. This type of vector is derived from a mouse cellular retro-transposon element that has no essential viral or cellular genes, and that has little sequence similarity to a retrovirus. However, this RNA (known as VL30 [virus-like, 5 30S]) has all the necessary *cis*-acting structural elements (such as LTRs and primer binding sites) required for efficient transmission by a type C murine or primate retrovirus. Thus, it is a parasite transmitted by retroviruses that is also expressed as a cellular RNA in most mouse cells and tissues. This RNA becomes packaged into retroviral particles when the mouse cells become infected by retrovirus. The retrovirus then transmits the VL30 (or a VL30 vector) to 10 the next infected cell (which can be a human cell). The RNA is then reverse transcribed and integrated into the DNA of the host cell.

Some advantages of VL30 vectors (over retrovirus-derived vectors) are: 1) lack of viral genes and other sequence homology that could lead to replication competent retrovirus (RCR); 2) ability to be expressed long-term *in vivo*; 3) a variety of LTR 15 transcriptional promoters that can be expressed in various tissues and under the influence of various hormones and other stimuli; and 4) the ability to express genes in a number of cell types that are targets of gene therapy. An additional advantage is that VL30 parts can be switched with those of classical retrovirus-derived vectors. For example, the LTR or packaging signal of VL30 can be used in place of the equivalent retroviral signal. The ability 20 to make mixed, or chimeric retro-vectors is a special application of gene self assembly technology.

Using a specific primer set, such as that shown in Fig. 2, or others, as taught in this invention, it is possible to amplify the U3 sequences expressed in the RNA of many different types of mouse cells. This is done using standard RNA isolation methods (Ausubel 25 *et al., supra*), coupled with extensive digestion with ribonuclease-free dextroxyribonuclease, to eliminate residual DNA. Thus, to obtain a promoter that is expressed in the liver, one isolates RNA from liver and uses an RT-PCR procedure, such as those known in the art, with the primers to amplify the desired promoters. Fig. 6 illustrates liver RNA-derived promoters obtained using the methods of this invention. However, the promoters can also be derived by 30 conventional PCR from cDNA libraries (Fig. 5 illustrates T cell-derived promoters that were obtained in this manner). It is also possible to use the well-known hormonal and pharmacological inducibility of VL30 LTRs to find LTRs that are responsive to peptides,

hormones, and cytokines (for a table and description of VL30 pharmacologic responses (see Hodgson et al., 1996 *Retro-Vectors for Human Gene Therapy*. RG Landes Company, Austin TX, chapter 4, and Fig. 4.2). Examples of substances inducing various VL30 promoters to high levels include: epidermal growth factor, basic fibroblast growth factor, insulin, erythropoietin, glucocorticoid hormones, activators of cyclic 3'-5'AMP, and others. To rescue promoters with pharmacological responsiveness, cells or animals stimulated with the desired pharmacological agent are subjected to the RT-PCR procedure and the resulting U3 regions are cloned into a vector, (such as the exemplary VLBPGN) and are tested for inducibility. Standard RNA blotting procedures can be used before isolating VL30 promoters, to determine whether a particular drug or hormone causes induction of VL30 RNA expression in a particular mouse cell or tissue. After the promoter has been rescued, the vector is transmitted via retrovirus to the target cell (possibly a human equivalent of the mouse cell from which the promoter was rescued). After selection with the drug G418 (400-700 µg/ml, for 7-10 days) to select against cells not containing the vector, the target cell population is challenged with the pharmacological agent of choice. Reporter gene expression (in the example, GFP) or RNA expression, as determined by RNA blotting, can be used as an assay of gene inducibility by the agent (for exemplary gene expression methods, see Chakraborty et al., *Biochem. Biophys Res. Commun.* 209:677-683, 1995).

Using any specific primer set designed for use with VL30 retro-elements and using total cellular RNA from a particular mouse cell type as a template for RT-PCR, (using commercially available kits and methods therein) candidate promoter elements can be amplified. This method is useful for the identification of mouse-derived promoters and in particular the method is useful for the identification of cell-type specific or tissue-specific promoters from a mouse and for the selection of these promoters and the identification of tissue-specific or cell-specific promoters that function in human cells. Thus, these types of vectors and the methods for using these vectors permits the identification of promoters to permit controlled transcription of a foreign gene. The promoters, originally obtained from the mouse, can be used to effect tissue-specific or cell-specific expression in a human or animal liver cell such as a hepatocyte, or in a human blood cell such as a T-helper cell or in an erythrocyte (red blood cell). Methods are disclosed in Example 2 for the screening and selection of the promoters from a library of amplified promoter sequences. Other methods are well known to those of ordinary skill in the art. The specificity of the selected promoter

can be assessed, for example, by introducing a selectable marker under the control of the test promoter in question and introducing this construct into various cells to assess the ability of the promoter to selectively regulate expression.

The amplified fragments represent U3 promoter regions from any RNA species expressed in the originating cells and their abundance will be in approximate proportion to the number of expressed copies of RNA in the original mixture. Example 3 illustrates one example using a mouse T-helper cell cDNA library to produce amplified fragments representing U3 regions expressed in T cells. The vectors were efficiently expressed as RNA and protein in PA317 helper cells, and were transmitted by retrovirus into human T-helper cells, where they were integrated and expressed as protein in the form of a β -galactosidase reporter gene, as visualized by X-gal staining. The products of this experiment are provided in Fig. 5 and as SEQ ID NOS: 2 and 3 from T-helper RNA. The products of another experiment are shown in Fig. 6 as SEQ ID NOS: 4-13 from mouse liver RNA (by RT-PCR).

Examination of the different U3 sequences isolated from T cells and from liver revealed several things. First, the T cell U3 sequences were related to each other, as were the liver sequences. However, the two types of U3 sequences were quite different between the two sources (T-cell, Figure 5 and liver, Figure 6). Specifically, the liver sequences (Figure 6) appeared to be a closely related group, differing mostly by single point mutations, some of which may affect transcription factor binding sites. Some of the polymorphic sites included: a phorbol ester response element (VLTRE); a Rel/NF κ b binding region, and a possible glucocorticoid response element (GRE). Some of these polymorphisms are illustrated in Fig. 6. The T cell-derived sequences (Fig. 5, SEQ ID NO:2 and 3), on the other hand, differed significantly in length, with SEQ ID NO:3 missing more than 120 bases (compared with SEQ ID NO:2) including putative binding sites for retinoids (RAR/RXR) and several elements contained within the enhancer repeat region (including a cAMP response element (VLCRE, or CREB/jun binding site), and putative serum response element (SRE, CARG, and NF1/IL6). SEQ ID NO:3 represented one out of five clones sequenced, while SEQ ID NO:2 represented four out of five. Possible sites of interactions between transcription factors and DNA can be observed by comparing the experimentally derived U3 sequences with those in Hodgson et al. ,(Retro-Vectors for Human Gene Therapy, 1996 Fig. 4.2 *supra*). In addition

to the deleted sequences of SEQ ID NO:2, there are a number of single base differences within the conserved regions of the two T cell-derived sequences.

Advantageously, a number of new VL30 promoter sequences (SEQ ID NOS: 2-13, *supra*) were identified using these methods despite the fact that VL30 RNA comprises only about 0.3% of cell mRNA represented in a cDNA library. Moreover, in each case, the cloned insert was isolated without the need to use linkers, adapters, or multiple cloning sequences such as those that are typically use for other library construction methods. The promoter sequences can be used in the vectors disclosed here to express inserted foreign genes or the promoter sequences can be substituted into other retroviral vectors, such as MoMLV-derived vectors or other VL30-derived vectors. Further, vectors containing the promoter sequences can be propagated in retroviral helper cells, such as PA317 (U.S. Patent 4,861,719 to Miller) or introduced into cells by chemical or physical transfection.

In another application of the methods of this invention, libraries of amplified sequences can be incorporated into vectors using two or more fragments and using the restriction endonucleases cleaving at a distance from their recognition sites. Preferably the vectors are created using six or more fragments and preferably greater than 10 or more fragments. For example, as applied to VL30 promoter sequences, because there are over a hundred VL30 retro-elements in the mouse genome, it is possible to amplify all of the promoter sequences *en masse*, and propagate them *en masse*, enabling screening by serial passage through helper cells (such as the PA317 helper cell line) or by means of a replication competent retrovirus, as illustrated in Examples 3 and 4. Conversely, the promoter region may be broken down into several sub-domains and permutations of each could be combined and screened to enhance the chances of generating a superior construct (Fig. 4B).

As an example of breaking a promoter region down into several sub-domains, Fig. 7 illustrates a similarity plot of nucleotide sequences found in VL30 U3 regions. Plot similarity was performed using the Plot Similarity program (Wisconsin Sequence Analysis Package, release 8.1, Genetics Computer Group, Madison, WI). This program plots the running average of the similarity among the sequences in a multiple sequence alignment. The sequences compared were those found in Fig. 4.2 of Hodgson, 1996, chapter 4 (*infra*). That is, the plot discloses the degree of conservation of VL30 promoter sequences among known VL30 promoters. From the figure, it can be seen that conserved sequences (close to 100% conserved) can be used as primer binding sites to amplify the adjacent sequences by PCR.

An allelic mixture of three fragment sets is then created to make a combinatorial library of promoters that can be positively selected, such as by using retroviral amplification of the active sequences. This, used in combination with the Fig. 4.2 (Hodgson, 1996, chapter 4 *supra*) can be used to determine regions of high similarity. Regions of high similarity within the U3 region can be replaced with one another. Therefore, a library of permutations of these sections can be made by combining allelic pools obtained by amplifying the sequences from individual subsections, followed by ligating the subsections in the correct order using the methods of the instant invention for gene self-assembly. For example, sub-section 1 can include the distal enhancer (from the LTR 5'-end to the site of insert primer 2, see for example the region defined by the insert primers 1 and 2 (SEQ ID NOS 55 and 56 of Example 4). In this way, using a plot similarity (such as Fig. 7), within each sub-section, the primers position fragments within a region of nearly 100% identity. Degenerate primers can also be used in these experiments to account for multiple nucleic acid base combinations along a particular sequence. In each case, the primers preferably are designed to have a melting temperature that is compatible with the RT-PCR conditions being used, and the conditions should be those recommended by the manufacturer (preferably Perkin Elmer Corp., Emeryville, CA). In Example 4, a set of primers is given that can be used to amplify different U3 subsections, together with directions for assembling a combinatorial library.

It will be appreciated by persons of ordinary skill in the art that the methods of the instant invention can thus be used to make allelic libraries of a variety of genes. For example, different allelic portions of a gene can be combined in a predetermined order and orientation to produce combinatorial libraries, without the need for fortuitous restriction sites separating the parts in the original construct, and without perturbing the important sequences joining the parts using the methods of this invention.

In this invention primers are constructed as described above. However, for the generation of allelic libraries or more complex library constructs it may be helpful to include 5'tags into the 5' end of the primer. The purposes of the tag sequence are: 1) to provide extra nucleotides on both sides of the restriction endonuclease recognition sites (for more efficient digestion); and 2) to enable recovery of sequence tags or undigested fragments by means of an affinity reagent (such as silica, magnetic beads, or nitro-cellulose containing the complementary sequences) for purification. The use of an affinity reagent permits the digested ends to be purified away from the digested fragments. Furthermore, if any

undigested ends remain after thorough digestion, the affinity reagent will remove them, further aiding in the purification. In one embodiment, affinity purification of the digested fragments is used in place of gel isolation, eliminating possible damage caused by ultraviolet light as well as possible damage caused by dye (e.g., ethidium bromide) binding to the DNA.

5 It will also be appreciated that a number of other variations to the primer sequences can be employed. For example, as discussed above, the enzyme recognition site for an enzyme that digests outside of its recognition sequence is included in the primer, so that the DNA digest creates an overlapping end that is complementary to one other terminus to which it will be joined. The enzyme recognition site can be moved to any location within
10 the primer so as to digest the DNA at the exact location desired. The primer can also be programmed with a novel enzyme recognition sequence to add any desired sequences between the two sequences to be joined or to incorporate a linker or adapter if desired. If the sequences to be amplified contain the enzyme recognition site of the primers, it may be necessary to switch to a different enzyme usage. The use of several different enzymes is
15 possible and has been discussed above. As with other PCR procedures, after the initial primer selections have been made the primers are assessed for their ability to fold back on themselves or to create internal secondary structure. The primers are preferably modified to avoid palindromic sequences or the potential for self folding within a primer. Nucleic acid analytical software (such as the Wisconsin GCG package, Oxford Biomolecular, Oxford, UK)
20 is available to perform this analysis and aid in the selection of alternative primers.

 In addition, as with all PCR processes, it is necessary to determine the melting temperatures (T_m), and to adjust the annealing temperature of the PCR reactions to compensate for such temperatures. Finally, it is important to perform a sequence redundancy search, to determine whether the target sequence (the sequence complementary to the primer)
25 is found more than once in the region to be amplified. If the sequence is repeated, it will be necessary to use a different primer in order to establish the single, correct priming site. Preferably, no more than 6-8 bases of incorrect target complementarity at the 3'-end of the complementary region is used and to allow a difference of at least 10° C between the T_m s of the correct and the incorrect target. The annealing temperature should always be at least 5°C
30 lower than the T_m of the correct target and 5°C above the T_m of the incorrect target. Again, the necessary software and instructions are readily available from the cited sources (Wisconsin Gene Computer Group and Oxford Biomolecular, *supra*)

Next, a vector is constructed to include the appropriate elements for expression in the desired cell type. For example, the plasmid of Fig. 3A can be used for the creation of a promoter library or a vector can be created using a commercially available vector and primers to create a three or more fragment annealing and ligation reaction as provided above.

- 5 Preferably, the inclusion of a dominant negative selectable marker on the vector (e.g., the neomycin phosphotransferase gene, conferring G418 drug resistance) can be used to reduce the likelihood that cells without the vector are being maintained in culture.

Multiple allelic copies of DNA (cell derived or cDNA) can be amplified in separate reactions as a set of potential inserts with each set having its own unique overlap
10 sequence following digestion with a restriction endonuclease, according to this invention. The fragments can then be ligated into an existing vector or in a single reaction of three or more fragments to form a combinatorial collection of potential alleles. For example, if six adjacent regions are amplified from five separate alleles, the number of combinations would be 5^6 , or 15,625 potential combinations. The combinations can then be grown *en masse*, and
15 selected *in vitro* or *in vivo*. A variety of screening strategies can be used in this invention and those of ordinary skill in the art will appreciate that the type of screen will match the type of library being generation. Therefore, for the promoter library, introducing members of the library into particular cell types to assess for expression in one or more cell types versus the absence of expression in another cell type is evidence of tissue-specific or cell-specific
20 expression. For screening purposes, the libraries of this invention function like other libraries created through other methods. A variety of screening methods for a variety of libraries have been described in the art. For example, selective screens are reviewed by Hodgson et al. (1996, RG Landes Company, *supra*). Reporter protein production is well known in the art as is dominant selectable marker (e.g. drug) selection and selection by fluorescence activated
25 cell sorting, antibody affinity selection, phage display selection (such as commercially available from Amersham, Milwaukee, WI), and the like can be used without detracting from this invention.

In this way, it is possible to isolate multiple forms of genes, gene fragments or regulatory regions such as transcriptional promoters or packaging signals (for example, in a
30 retro-vector system). The individual constructs may then be tested *in vitro* or *in vivo* to further characterize a particular phenotype.

In one example the method is used to create a library of complementarity determining regions (e.g., allelic variations that give rise to antibody diversity) of antibodies or from receptors, including T-cell receptors, epitopes, antigens, ligands and the like. For example, where a library of T-cell receptors is created, the introduction of a vector designed to create a functioning T-cell receptor can be introduced into T cells or T-cell progenitors and the cells can be tested for their ability to bind to a particular test ligand. The ligand-recognizing cells can then be isolated from the ligand and grown in the presence of cytokines to produce specialized T cell clones. Where a library of antibodies or antibody fragments is created, the antigen reactive portions, for example, can be recombined in a vector containing the remaining portions of an antibody molecule to generate antibodies or antibody fragments in a cell. In other examples, the methods of this invention can be used to create allelic domains of receptor families (such as the steroid receptor super-family); libraries with related regions from peptide hormones; cytochromes P450; or other protein families that have shared domains or sub-sections with similar structures. The methods of the instant invention allow the joining of allelic sub-sections in an ordered fashion. In each case, it will be necessary to design primers, and to keep track of the uniqueness of joining overlaps and the presence of internal restriction sites as described above. While these will be different in each case, here are listed some general guidelines that are incorporated into the method of the instant invention.

As discussed above, although described as it relates to promoter libraries, libraries of other nucleic acid sequences can be created using the methods of this invention. These libraries include, introns and/or exons and/or functional domains libraries, libraries of potential alleles for a particular gene sequence, and the like. These sequences can be amplified from cell DNA or RNA using the primers of this invention and incorporated into a variety of vectors. For example, one vector of this invention, VLBPGN, has a portion of LTR removed and can be used to create a variety of libraries following digestion with *Bpm*1.

Selected or screened products of the combinatorial library can be used for gene expression, such as the promoters of Figs. 5 and 6. In addition, the exploitation of these sequences for the expression of a variety of genes, the LTR fragment containing the promoter can be joined to one or more functional retroviral packaging signals, internal ribosome entry sites, additional promoters, coding regions, processing sites, and the like.

Advantageously, there are almost no spatial constraints upon the joining of molecules by the method of the instant invention and other methods have not taken advantage of the combination of PCR to isolate genes or gene fragments; enzymes cleaving at a site distant from their restriction endonuclease recognition site to combine three or more fragments with precision; and, the use of unique overlapping non-palindromic termini to ensure fidelity of multi-fragment ligations. This combination permits the artisan to prepare complex gene constructions in one ligation step and does not require sequential sub-cloning into a vector or propagation in a prokaryotic host. Added to this the combination by these methods of fragment pools facilitates recombinatorial genetics.

The ability to recombine (in the correct order and direction) and screen a large number of allelic variants (whether as a simple library or as a combinatorial library), resulting in increased abundance (by amplification in the RNA, and subsequently in the DNA) is a special characteristic of this invention. Particular advantages of this system are obtained when the methods of this invention are combined with retrovirus vector technology or other virus vector technology. For example, the combination provides a form of *in vitro* evolution whereby the passage of the library through virus and through cells selects functioning sequences and increases the abundance of the surviving RNA and DNA molecules.

For example, consider the consequences of screening several different promoters expressing RNA in a donor cell (*i.e.*, a cell producing virus particles), but at differing levels of RNA abundance. In the following example, the least abundant RNA species is expressed at 0.1 copy of RNA per cell, while six others are expressed at 1 copy, 10 copies, 100 copies, 1,000 copies, or 10,000 copies, or 100,000 copies/cell, respectively. After a single passage, the DNA copy number in the recipient cells now reflects the approximate RNA copy number in the donor cells. These numbers are further amplified in the relative abundance of RNA species produced in the recipient cells. Disallowing for factors such as position effects, transcription factor depletion, etc., (which may be considerable), the same relative ratios of expression would be expected. Taking into consideration position effects, the disparity between abundance caused by changing insertion loci should average out. The most abundant RNA species after two passages is then many orders of magnitude more abundant than the least abundant.

Species:	RNA abundance: P=0	DNA copy no. P=1	RNA abun. P=1	DNA copy no. P=2	RNA abun. P=2
A	0.1 copy/cell	0.1	0.01	0.01	0.001
B	1	1	1	1	1
C	10	10	100	100	1,000
D	100	100	10,000	10,000	10 ⁶
E	1,000	1,000	10 ⁶	10 ⁶	10 ⁹
F	10,000	10,000	10 ⁸	10 ⁸	10 ¹²
G	100,000	100,000	10 ¹⁰	10 ¹⁰	10 ¹⁵

Table 2. Enhancement of DNA and RNA copy number as a result of different RNA expression levels, after retroviral passage. P= (no. of passages). Numbers are interpreted as relative ratios within a column.

5

The present invention is able to efficiently create a library of RNA or DNA sequences whether or not they are in low abundance. The kinetics of screening for RNA abundance of a promoter can be appreciated best in the following discussion. For the purposes of this discussion, position effects have been ignored. An equation describing the

10 kinetics of screening for RNA abundance is:

$$(1) R_{rel\chi} = A_{\chi} / \sum A_{n=\infty}$$

The above equation (1) can be stated in plain English: The relative abundance of an RNA species χ ($[R_{rel\chi}]$) within a population of RNA molecules expressed in a single cell or within a population of cells) is equal to the RNA copy number of RNA species χ (A_{χ}) divided by the sum of the RNA copies of all RNA species present, including χ .

The relative abundance number of any given species changes as the number of passages change, according to the following approximation:

20

$$(2) R_{\chi py} = D_{\chi p0} R^{p+1}$$

In the simplest of terms, equation two (2) can be expressed as: The abundance of RNA species χ after Y passages ($R_{\chi py}$) is equal to the initial abundance of the DNA for species χ at passage=0 ($D_{\chi p0}$), multiplied by the RNA abundance/DNA copy, raised to the power of the number of passages plus one. Thus, a typical RNA species that starts out as a

25

single copy of DNA, after zero passages (*i.e.*, in the donor cell) expresses 10 copies of RNA/cell. After one passage it is amplified at the DNA level to a relative ten copies (the same as the RNA abundance at $P=0$), and at the RNA level to 100 copies (10 copies per DNA copy). The reason for the amplification is that viral packaging and passage is based upon the number of RNA copies present in the donor cell. These calculations can be used to arrive at approximate abundance determinations for any given passage. The actual results of any given experiment, of course, will be biological rather than physical or mathematical. This means that other variables such as RNA efficiency of transmission and longevity, availability of transcription factors, experimental variation, *etc.* also come into play. The underlying purpose of the approximating equations, however, is to illustrate that RNA is amplified in DNA in proportion to the abundance of the template (RNA) within the cell.

The abundance of mRNA in cells can vary continuously from less than a copy per cell to nearly 100,000 copies/cell in actively transcribing, highly-specialized cells such as reticulocytes, the chicken oviduct, the silk moth silk gland, *etc.* Therefore, the spectrum of RNA abundance from $0-10^5$ /cell is within the biological window of interest. For most practical purposes, such as biotechnological expression of genes in specific cells, only the higher end of this abundance range is desired. Therefore, using a viral selection system, as disclosed in this invention, it may be possible to disregard those species with less than a threshold level, such as <0.1 copies per cell. The selection through virus will lead to the recovery of the more abundant species. Furthermore, because the vector is likely to be the only considered sequence, it may be considered as a proportion of the whole of RNAs expressed in the target cell. The situation is more complex when a large number of permutations and combinations is generated, for example by self-assembling thousands or millions of fragments in a predetermined order using the self-assembly technique of the instant invention. Consider the assembly of allelic variants of four promoter subregions: distal enhancer, proximal enhancer, distal promoter and proximal promoter. If 100 varieties of each of the four groups were amplified and combined using the instant process along with a single vector, 10^8 resultant combinations could occur. However, a sufficient number of molecules to start out a combinatorial screening program might be a million. The problem can be simplified by considering these in groups as follows:

Table 3. Grouped abundance of RNA molecules derived from combinations.

No. of species in group:	RNA abundance:	Total No. RNA molec. at P=0:	RNA at P=1	RNA at P=2	RNA at P=3
9×10^5	1	9×10^5	9×10^5	9×10^5	9×10^5
2×10^5	10	2×10^6	2×10^7	2×10^8	2×10^9
2×10^4	1,00	2×10^6	2×10^8	2×10^{10}	2×10^{12}
1×10^3	1000	1×10^6	1×10^9	2×10^{12}	2×10^{15}
1×10^1	10,000	1×10^5	1×10^9	1×10^{13}	1×10^{17}
1	100,000	1×10^5	1×10^{10}	1×10^{15}	1×10^{20}
Sum Total:		6.6×10^6	1.11×10^{10}	1.01×10^{15}	1×10^{20}

Thus, it follows that in the example population (Table 3) of over a million constructs (equally represented in the DNA), a single construct expressing 10^5 copies of RNA per DNA copy will increase to approximately 99% of the total expressed RNA sequences in two passages. Using similar procedures in combination with drug and/or hormonal stimulation, and after consideration of the possible transcription factor binding sites within the sequence family (Figs. 5 & 6), it is within the intended scope of the invention to select for hormonal or pharmacological controls of transcription such as have been described herein. The factors contributing to the outcome are not only the input constructs, but recombinants and mutants as well. These secondary contributors to molecular diversity will be enhanced if multiple rounds of infections are allowed to occur, as oftentimes the difference between a particular transcription factor being able to bind (or not) may depend upon a single base change. Because viral infection is progressive and competitive, molecular evolution can be used to generate gene constructs *de novo* in the tissue culture dish in short time periods. Advantageously, the use primers to generate amplified fragments with uniquely complementary cohesive ends (i.e., that the ends will preferably only hybridize with the intended 5' and 3' fragments) to ligate three or more fragments as taught in this invention improves the potential for obtaining a diverse library.

Although the examples particularly point out a transcriptional promoter as the product of the process, the skilled artisan can appreciate that a particular selection technique can be applied to other *cis*- and *trans*-acting genetic sequences as well. Although a virus is used to propagate the selective advantage of a preferred embodiment, it can also be appreciated that any selective screen, such as drug selection, cell survival, phenotypic selection, cell sorting, antibody selection, and the like (see Ausuble et al., *supra*) could be

substituted without changing the intended scope of the invention. Likewise, transfection or cell fusion could be used in place of viral infection. Furthermore, substitution of different viruses, retrotransposons, or functional groups are likewise within the intended scope of the invention. The described embodiments are to be considered only as illustrative and not
5 restrictive, and the scope of the invention is indicated by the claims rather than by the narrative description. All references and publications, cited herein, are incorporated by reference into this disclosure.

Like the embodiments detailed above, the method of library production is also conducive to assembly and transfer of genetic material directly into eukaryotic cells, saving
10 the step of propagation in bacteria that is standard in bacteria. An advantage of direct transfer of the libraries of this invention to eukaryotic cells, including the exemplary retroviral vector producer cells, is that certain essential *cis*-acting structural features will be under positive selection (i.e., if they are not present, the molecule will be lost due to its non-functionality). As discussed above, it is often advantageous to eliminate bacterial and plasmid DNA
15 sequences, endotoxin, and other bacterial contaminants by introducing the constructs directly into eukaryotic cells.

In addition to providing a method for constructing complex DNA molecules efficiently (as in the examples of three piece and six piece constructs), the methods of this invention permit the assembly of constructs that are larger than those conventionally
20 propagated in *E. coli*. Examples of these types of vectors include adenovirus vectors, herpes simplex vectors and artificial minichromosomes. In order to insert genes into such vectors that are too large for conventional molecular cloning procedures, in the past it was often necessary to resort to *in vivo* recombination, wherein the genes of interest are cloned into a suitable vector and the flanking homologous regions are used to target the foreign genes to a
25 homologous site within the larger viral or minichromosome vector. However, the methods of this invention permit PCR fragments of any size (up to the limits of PCR capability, 20-30 kb per fragment) to be joined together. Thus, it is feasible to precisely construct adenovirus vectors by amplifying larger sequences, and combining them by ligation. For example, several sections of adenovirus (5-10 kb each) can be ligated using the methods of this
30 invention, up to for example, about 37 kb, and then transformed directly into human cells. Only the correctly recombined vectors are capable of replicating. Hence, the DNA is autoselecting. A similar procedure is used for generating herpes virus vectors, which are

approximately 150 kb. The precision of the methods of this invention permit non-essential-viral genes to be more easily eliminated from the construct. After transfection into appropriate cells, the DNA replicates and virus particles are formed.

Some special considerations apply to larger vectors, however. First, it is
5 desirable to use enzymes that do not cut within the large DNA fragments. To prevent excessive fragmentation of the DNA by internal sites, it is desirable to use enzymes that cut rarely or infrequently, such as CpG-containing enzymes recognizing six bases, or enzymes such as *Sap1*, recognizing seven bases and digesting a three bp overhang (thus permitting up to 32 fragments to be joined in order). It is also desirable to avoid shearing the DNA once
10 large segments have been joined by ligation. One method of avoiding shear is to add the transfection agent, such as Superfect[™] reagent (dendrimers, Qiagen) or Lipofectamine[™] (liposomes, Life Technologies, Gaithersburg, MD) directly to the ligation reaction, and then add the cells to be transfected to the mixture. This, or a similar method avoids the need to physically move the ligated DNA, and thus prevents shearing. Another method is to add a
15 DNA condensing reagent (dendrimers, polycations [such as polyethyleneamine] histones or liposomes) directly to the DNA ligation reaction, and then move the DNA by pipette after it has condensed (thus reducing shearing of the DNA). Once inside the cell, viral DNA can replicate (as in the examples of partially replication-competent adenovirus and herpes simplex virus vectors).

20 Artificial minichromosomes have been under development for years. True artificial chromosomes require a centromere, at least one origin of DNA replication, and in the case of linear molecules, telomeric repeats at the chromosomal termini. In addition, to be very effective it is desirable to have a selectable marker gene, one or more therapeutic genes, and/or reporter genes.

25 In reality, the use of minichromosomes has been delayed by the inability to effectively manipulate the larger DNA molecules *in vitro*. Yeast and bacterial artificial chromosomes have been used with little success in mammals, and the addition of telomeres to the ends of linear chromosomes is also a special problem, as there is no prokaryotic host that can tolerate large linear DNA. The methods of this invention offers the opportunity to
30 assemble human or mammalian minichromosomes *in vitro*, by using large segments (10-30 kb) of synthetic, gene-amplified DNA as ligation starting materials. For example, up to 32 *Sap1* fragments (up to 30 kb each, containing the essential *cis*- and *trans*-acting sequences),

or 512 shorter *Hga*1 fragments can be combined using these methods. As with the other examples, several enzymes suitable for this invention (e.g., such as class IIS enzymes) can be combined (possibly with different termini lengths) to simplify the task. The methods of this invention also facilitate construction of telomeric repeats, because the constructs of this invention do not need to be circular. Thus, the methods of this invention can be used to make telomeres of any length, by adding additional segments onto the ends of molecules. One way to do this is using self assembling genes that employ a repeating overhang sequence (self-complementary molecule, such as AG-3' at one end, and CT-3' at the other end), permitting the telomeres to be lengthened to the extent desired by adding the required molar excess of the telomeric repeat-containing fragment. This technique gives the investigator some control over the relative length of the telomeres, although the self-complementarity indicates that many repeats will be lost due to self-ligation. This can be alleviated by using higher starting concentrations of DNA to favor inter-molecular ligations over intra-molecular ligations (e.g., >20 µg/ml starting concentration of DNA).

A two fold molar excess of telomeric fragments gives approximately twice the average length of telomere as a strictly 1:1 molar ratio of all fragments. By using a higher molar ratio of shorter telomeric repeats it is possible to give greater uniformity to the overall length of the molecules, which will vary from one terminus to the other. Thus, in addition to providing a way to build large molecules with precision, the methods of this invention provides for a way to control the telomere length (or potential life-span) of the artificial chromosome. To prevent damage during handling, the minichromosome DNA can be condensed with polycations, adenovirus particles, dendrimers, histones, or liposomes prior to transfection, similar to larger viral vectors.

The methods of this invention can be used to create recombinant virus. One example of this is an adenovirus vector self-assembling gene system. This system can include three parts: 1) vector; 2) helper virus; and 3) helper cells. The vector part is a self-assembling fragment set of at least three fragments comprising the essential cis-acting sequences (left and right inverted terminal repeats, which are the 103 bp at both ends of the genome that are required for replication [ITRs] and packaging sequences [Y, base pairs 194-358) and central 'baggage' area, comprising one or more self-assembling fragments including therapeutic genes, marker genes, and reporter genes. The baggage area is thus flanked by the cis-acting sequences in the vector. Because the synthetic oligonucleotide sequences

comprising the 5' and 3' termini of the helper virus are not phosphorylated, they will not ligate together creating multimers. Thus, the Ad5 vector region will assemble only into monomers. The helper virus part comprises all Ad5 trans-acting genes except for the E1A and E1B genes. The helper virus part has no cis-acting sequences, and it is amplified in several sections. In this preferred embodiment, the virus is amplified using primers that exclude the ITRs, packaging region and E1A&B genes. The helper virus is digested by *Sap1* digestion, creating seven uniquely terminated fragments comprising the trans-acting viral genome, with dephosphorylated, blunt 5' and 3' ends on the terminating fragments. The primers are designed so as to amplify the internal virus sequences without changing them, except for the 5' and 3' ends of the virus. The PCR-amplified fragments are digested with *Sap1* and are religated in their natural order after gel isolation and Qiagen column purification. The 5' end of the helper virus genome starts at 3.2 kb (in the E1A gene) so as not to overlap the vector sequences, which could otherwise cause replication competent adenovirus (RCA). Because the 5' and 3' ends of the helper virus do not contain *Sap1* sites, they remain intact after digestion with *Sap1*. Because the synthetic oligonucleotide sequences comprising the 5' and 3' termini of the helper virus are not phosphorylated, they will not ligate. Thus, the Ad5 helper virus genome assembles only into preferred monomers during ligation.

In a preferred embodiment, non-essential genes are deleted from the Ad5 genome by means of the method of self-assembling genes. In another preferred embodiment, the helper virus genome is approximately 30 kb after deletion of E1A, E1B and E3 gene sequences from the helper virus, and it is amplified as a single long fragment using the eLONGase Amplification System (Life Technologies or a similar strategy for creating long PCR fragments with high fidelity). It is not of great importance that occasional PCR errors may occur, because multiple copies of the Ad5 helper virus are transfected into target cells, thus providing trans-complementation. The helper cells are preferably 293 cells, a human kidney cell line expressing E1A and E1B genes (ATCC). The vector part and the helper virus part are combined in equimolar ratios after ligation has been performed separately on each fragment set. The Superfect protocol (Qiagen) is used to transfect the vector part and the helper part into the helper cells. The helper cells lyse, releasing high-titer adenovirus particles that are capable of infecting a variety of human cells. The resulting defective virus is incapable of forming RCA, and it transmits up to 34 kb of foreign genes in the baggage area. Unlike conventional Ad5 vectors that require separate constructs for *E. coli* propagation of

insert genes, and recombination in vivo, the present vectors are relatively easy to make and provide a precise, safe alternative to first generation and second generation adenovirus vectors.

Exemplary methods for producing self-assembling vectors and genes are provided below. Further, the Examples provide methods for producing libraries of nucleic acid sequences using the methods of this invention. A number of nucleic acid sequences identified using the methods of this invention are described. The examples provided below are exemplary and not limiting. All references and publications provided herein are incorporated by reference into this disclosure.

10

Example 1 **Three-Piece Gene Self-Assembly with 100% efficiency**

Using 6 primers (SEQ ID NOS:24 and 63-67), three PCR fragments were amplified from templates VLMG (SEQ ID NO:22) and VLBPGN (SEQ ID NO: 1). PCR reactions were carried out using the hot start technique, according to the manufacturer's instructions (Perkin Elmer) using *Pfu* DNA polymerase (Stratagene). To amplify specific portions of the above templates, each primer contained a class IIS enzyme site capable of digesting a unique overhanging end that was complementary to only one other terminus in the subsequent ligation. The class IIS enzymes used were *Bpm1* and *Eco 57I* (the latter was used to copy a fragment that contained an internal *Bpm1* site). The reactions were carried out as follows: 1) the lower reaction was assembled according to the protocol for PCR Gems (Perkin Elmer); 2) the lower reaction was heated to 80°C, 5 min, then cooled to 4°C for 5 min; 3) the upper reaction was prepared according to PCR Gems protocol and was added to the lower reaction (separated by cooled wax). The primer concentration was 0.3 µM (final). The dNTP concentration was 200µM (final). 5 Units of *Pfu* polymerase was used. All fragments were amplified using the following conditions: 96°C, 45 sec; (then followed by 30 cycles of the following) 96°C 45 sec, 52°C 45 sec, 72°C, 6 min; then followed by a single incubation at 72°C for 10 min; then hold at 4°C. All fragments were successfully amplified. The PCR fragments were purified using the Qiaquick PCR purification protocol (Qiagen). The fragments were digested with an excess of the appropriate restriction enzyme (*Bpm1* or *Eco57I*). The digested fragments were run on a 1% agarose gel and were excised using minimal irradiation from a hand-held 365 nm ultraviolet light. The fragments were purified

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using the Qiagen Qiaquick Gel Purification Protocol. The fragments were ligated at an equimolar ratio at a concentration of $>20\mu\text{g/ml}$ with T4 DNA ligase (Boehringer Mannheim) overnight at 4°C . Competent *E. coli* SCS110 cells (Stratagene) were transformed with the ligated DNA. Eight colonies were characterized by restriction enzyme analysis, and all eight
5 contained the correct order and orientation of the three fragments. The experiment was repeated independently by another investigator, and the same result was obtained ($8/8=100\%$). Thus, the procedure resulted in a high percentage of correctly assembled vectors.

This three-piece vector was VLΔBP. The deletion extended from the distal
10 enhancer region to the TATA box near the start of transcription. The deletion region was a pair of *Bpm1* sites that permitted U3 sequences to be cloned into the insert.

One validated *E. coli* clone of VLΔBP was transfected into retroviral helper cells. After 48 h, the vector was transduced into amphotropic helper cells. After selection for two weeks with the drug G418, drug resistant colonies were grown up in a mass culture and
15 the vector was transduced from the amphotropic helper cells into a human HT1080 cell line (ATCC, Rockville, MD). Surprisingly, even with a large deletion in the LTR promoter, the basal TATA box-containing VLΔBP was transmitted as a retrovector and was permanently inserted into the human cell line, thus establishing the validity of the self-assembly technique for the construction of functional eukaryotic vectors.

20

Example 2

Production of a Six Piece Self-Assembling Expression Vector

Due to the high efficiency of the gene self assembly process for the three piece
25 assembly, a complex vector containing six fragments was constructed. The results here were extended to determine whether such a self-assembled vector would also have biological activity in human cells without being cloned and grown in a prokaryotic cell.

Six fragments were individually constructed by PCR using three different templates and twelve primers (as illustrated in Fig.8). The primers used three different class
30 IIS enzymes. The enzymes were chosen so as to give 2 base pair, 3'-overhanging ends. Three enzymes were used in order to avoid the use of enzymes that had additional sites internal to the fragments being amplified. Thus, *Bpm1* was used unless there was an internal *Bpm1* site. If such a site existed, *Eco57I* was used. If there was also an internal *Eco57I* site, then *BsrD1*

was used. However, it is alternatively possible to use an enzyme such as *Eam*11041, where the *Eam*11041 sites in the primers are unmethylated (therefore susceptible to digestion by the enzyme), and wherein the ^ms dCTP analog of dCTP is used in the PCR reaction, methylating all internal sites (and protecting them from digestion by *Eam*11041), as suggested by Padgett
5 and Sorge, 1996, *supra*, and incorporated herein by reference.

Using 12 primers, 6 fragments were amplified from 3 templates: pBK-CMV (SEQ ID NO:26), pVLMB (SEQ ID NO:23) and pVLOVhGH-900 (SEQ ID NO:21).

Fragment 1 was amplified from pBK-CMV using primers 1 and 2 (SEQ ID NOS:31 and 32).

Fragment 2 was amplified from pVLMB using primers 3 and 4 (SEQ ID NOS:33 and 34).

10 Fragment 3 was amplified from pVLOVhGH-900 using primers 5 and 6 (SEQ ID NOS:35 and 36). Fragment 4 was amplified from pVLMB using primers 7 and 8 (SEQ ID NOS:37 and 38). Fragment 5 was amplified from pVLMB using primers 9 and 10 (SEQ ID NOS:39 and 40). Fragment 6 was amplified from pVLMB using primers 11 and 12 (SEQ ID NOS:41 and 42). PCR reactions were carried out using the hot start technique, according to the
15 manufacturer's instructions (Perkin Elmer Ampliwax PCR GEMS 100). The lower reaction was heated to 80 °C for 5 min, then cooled to 20 °C for 5 min. The upper reaction was prepared according to PCR gems protocol and was added to the lower reaction (separated by cooled wax). The primer concentration was 0.3 micromolar (final). The dNTP concentration was 200 μM (final). 5 U of *Pfu* polymerase (Stratagene) was used per reaction. 100 ng of
20 template was used for each reaction. 14 rounds of PCR amplification were used to reduce mutagenesis of the templates. The PCR cycling protocol was 96 °C 45 sec; then two cycles of (96°C 45 sec, 52°C 45 sec, 72°C 6 min); then 12 cycles of (96°C 45 sec, 58°C 45 sec, 72°C 6 min) followed by a 72° C soak for 10 min, then to 4°C hold.

The six PCR fragments were designed to self-assemble into a retro-vector after
25 digestion with the correct class IIS restriction enzyme (Fig. 8). After transfection into retroviral helper cells, the vector DNA is transcribed as RNA by means of the cytomegalovirus immediate early promoter (fragment 1). This promoter replaces the retroviral or VL30 LTR in this vector. The RNA transcript region begins with the R and U5 regions of the Moloney murine leukemia virus (MoMLV) LTR, the viral packaging signals (Ψ) region of MoMLV, the packaging enhancer (Ψ+) region of mouse VL30 and the IRES
30 region of EMCV fragment 2. Fragment 3 consists of the human growth hormone (hGH) cDNA sequence. Fragment 4 consists of the SV40 virus early region promoter driving

expression of the neomycin phosphotransferase (neo) gene. Fragment five consists of the (+)-strand primer binding site of the MoMLV LTR, the U3 region of the MoMLV LTR, the repeat (or R) region, and a portion of the U5 region. Fragment 6 consists of the PBR322 plasmid origin of replication.

5

Fragment 1: CMV early region promoter

Template: pBK-CMV plasmid DNA (Stratagene, LaJolla, CA) *Bpm1* (SEQ ID NO:26)

PCR primer 1 (SEQ ID NO:31)

10 GACTAACCTTGATTCCACTGGAGCCGTATTACCGCCATGCATTAGTTATTAATAG

PCR primer 2 (SEQ ID NO:32)

GACTAACCTTGATTCCACTGGAGTAATTGCGGCTAGCGGATCTGACG

Fragment 2: R-U5-Psi-Psi(+)-IRES *Bpm1*

15 Template: pVLMB plasmid DNA (SEQ ID NO:23)

PCR primer 3: SEQ ID NO:33

GACTAACCTTGATTCCACTGGAGACACTTGACCTCTACCGCGCCAGTCCTCCGAT
TGACTGAGTCG

PCR primer 4: SEQ ID NO:34

20 GACTAACCTTGATTCCACTGGAGGGATCCGCGCCCATGATTATTATCG

Fragment 3: human growth hormone (hGH) *Bsr* DI

Template: pVLCNOVhGH plasmid DNA (SEQ ID NO:21)

PCR primer 5: SEQ ID NO:35

GACTAACCTTGATTCCAGCAATGTCGGTTAGCTTGTTCCTTTACTGTTTGTC

25 PCR primer 6: SEQ ID NO:36

GACTAACCTTGATTCCAGCAATGTTAGGACAAGGCTGGTGGGCACTGG

Fragment 4: SV40 early promoter-neomycin phosphotransferase

Template: VLMB plasmid (SEQ ID NO:23)

30 PCR primer 7: SEQ ID NO:37

GACTAACCTTGATTCCACTGGAGGGTCGACCCTGTGGAATGTGTGTCAG

PCR primer 8: SEQ ID NO:38

GACTAACCTTGATTCCACTGGAGAATCTCGTGATGGCAGGTTGGGCGT

Fragment 5: MLV(+)PBS-U3-R-U5

5 Template: VLMB plasmid (SEQ ID NO:23)

PCR primer 9: SEQ ID NO:39

GACTAACCTTGATTCCACTGAAGAGATTTTATTAGTCTCCAGAAAAAGGGGGG

PCR primer 10: SEQ ID NO:40

GACTAACCTTGATTCCACTGAAGCCCCCAAATGAAAGACCCCCGCTGACG

10

Fragment 6: PBR322 origin of replication

Template: VLMB plasmid (SEQ ID NO:23)

PCR primer 11: SEQ ID NO:41

GACTAACCTTGATTCCACTGGAGCCGGGACGGAATTCGTAATCTGCTGC

15 PCR primer 12: SEQ ID NO:42

GACTAACCTTGATTCCACTGGAGTTCTCGAGGCGGCGCATCTCGGCG

Procedure: The twelve primers were prepared by the following procedure: 1)
oligonucleotides were synthesized with trityls off. After deprotection and lyophilization, the
20 samples were resuspended in 5 microliters deionized formamide and loaded onto a
polyacrylamide gel (12% polyacrylamide, 250V). The samples were excised under short
wave UV irradiation and eluted overnight in 600 microliters of sample elution buffer (0.5 M
ammonium acetate, 10 mM Mg acetate, 1 mM EDTA, 0.1% SDS). The contents were loaded
onto a BioRad Chromatography column (Cat. # 732-6008) and centrifuged into an Eppendorf
25 tube at low speed (2000 RPM, 5 min). After washing the column with 500 microliters TE
buffer (10 mM Tris, 1 mM EDTA), pH 8.0 and recentrifugation (2000 RPM, 5 min), the
pooled eluate was ethanol precipitated, washed with 100% ethanol, resuspended in TE buffer
and quantitated by spectrophotometry of a small sample, which was then discarded.

Fragments were cleaned using the Qiaquick PCR cleanup procedure. The
30 fragments were digested with their respective class IIS restriction enzyme. The digested
fragments were run on 1% agarose gels, and the fragments were excised and cleaned using
the Qiaquick gel cleanup procedure. Fragments were combined in an equimolar mixture and

ligated overnight at 4° C with T4 ligase and ATP. An analytical gel was run with the ligated DNA, as well as with controls including unligated fragments and ligated fragments with a single fragment missing. As opposed to the controls, the complete ligation included bands equivalent to the full-length supercoiled monomer (referred to as GENSA 981, SEQ ID NO:29), as well as bands possibly representing multimers (up to six bands were observed).

In order to assess the efficiency of the method, eleven nanograms of DNA were transfected into SCS1 supercompetent cells. Thirteen kanamycin resistant colonies were harvested, and plasmid DNA preps indicated 10 out of thirteen that appeared to be the correct length. All ten gave the expected bands when digested with *Pst*I, *Sna*BI, and *Bam*HI. 1.35 µg of the ligated DNA was purified by phenol-chloroform-isoamyl alcohol extraction, followed by two extractions with chloroform-isoamyl alcohol, and was precipitated in ethanol. The DNA was washed in 70% ethanol and re-suspended in 50 µl of sterile phosphate buffered saline (for transfection). The DNA was transfected (using the Qiagen Superfect protocol) into HTam1 (amphotropic human helper cells). 24 h after transfection, the target cells were washed and fresh culture media was added. 48 h after transfection, the supernatant from the vector producer cells was filtered (0.45 µm, Nalgene) and transferred to PG13 helper cells (ATCC) and HT1080 human fibrosarcoma cells. This procedure was repeated after 72 h. 48 h after transduction, recipient cells were started on G418 drug selection (500 µg/ml). The appearance of G418 drug-resistant colonies on transduced PG13 and HT 1080 cells after 6 days of selection indicated successful transmission via retrovirus particles. The transfect HTam cells were also selected with G418. After six days of drug treatment, 45 colonies of resistant cells were counted. Thus, the six fragment gene assembly was effectively transmitted and expressed as either a DNA (transfection) vector or a retro-vector.

25

Example 3 Design and Construction of Single LTR Vectors

Background: In order to manipulate the interior of the VL30 LTR sequences using a promoter rescue technique, single LTR vectors were constructed. The mouse VL30 element NVL-3 was used as the starting material as it is constitutively and abundantly expressed in most mouse tissues. Single LTR vectors are circular and behave as if they contained two LTRs. Thus, in these vectors RNA transcription begins at the start of the R region (see Fig.

30

3B), and continues through the polyadenylation site after completing the second round of transcription of the R sequences (Fig. 3A). In previous studies, these vectors were expressed transiently in vector producer cells and the DNA did not integrate into cell DNA as a standard two LTR vector. Therefore, the vectors were usually passed to a second complementation helper cell line via retroviral transduction of the vector RNA transcribed in the first helper cell. This process resulted in the vector regenerating a correct (two LTR) structure upon integration into the recipient cell DNA.

Experimental method: The plasmid pNVL-3 (SEQ ID NO:25, kindly provided by Dr. J. Nortonm Manchester, UK), containing a complete copy of the NVL-3 (mouse VL30) genome (Adams *et al.*, 1989), was digested with *Xho*I (which cuts in the LTRs), releasing the 4.27 kb VL30 genome with one copy of the LTR. This fragment was circularized using T4 DNA ligase and ATP. The circular DNA was linearized by digestion with *Sna*BI, 187 bp from the 3'-LTR. A 2.3 kb fragment containing the SV40 virus early region promoter and the aminoglycoside phosphotransferase (*neo*) gene, together with the PBR322 plasmid origin of replication, was excised from the BAG retrovirus vector (Price *et al.*, *Proc. Natl. Acad. Sci.* 84:156-160, 1987, kindly provided by C. Cepko, Cambridge, MA). BAG is also obtainable in a retrovirus helper cell line from American Type Culture Collection (ATCC), Rockville, MD by digestion with *Xho*I and *Bam*HI. This fragment was blunted with T4 DNA polymerase and dephosphorylated with calf intestinal alkaline phosphatase (CIP). The fragment was then ligated to the linearized *Sna*BI fragment of NVL-3. The resulting plasmid (containing a circularly permuted NVL-3 genome with the SV-*neo-ori* region) was designated VLSNO2 (SEQ ID NO:30).

In order to facilitate the switching of LTR sequences by means of the class IIS enzyme *Bpm*I, VLSNO2 was digested with *Bpm*I (six sites). The region containing four *Bpm*I sites was removed and replaced with a 19 bp linker (SEQ ID NOS: 1 and 52, see below), 921 bp beyond the LTR. The linker contained *Sna* BI, *Cla*I and *Bam* HI cloning sites.

Linker (top strand): 5'-TACGTATCGATGGATCCGA-3' (SEQ ID NO:51)

Linker (bottom strand): 5'-GGATCCATCGATACGTAAG-3' (SEQ ID NO:52)

The remaining two of the *Bpm*1 sites had complementary ends, which permitted their ligation and resulted in eradication of all *Bpm*1 sites within the resulting vector VLSNO3 (SEQ ID NO:20).

5 In order to facilitate reporter/therapeutic gene function, a 3.7 kb fragment containing the internal ribosome entry site (IRES) from encephalomyocarditis virus, together with the β -galactosidase reporter gene, was excised from the plasmid pVLSAIBAG (kindly provided by Mr. James Grunkemeyer, Omaha, NE) by means of a partial digestion of the plasmid with *Bam* HI. This region was inserted into the *Bam* HI site of VLSNO3, resulting in the vector VLSNOSIB (SEQ ID NO:14).

10 A second reporter construct, pVLSNOG (5774 bp, SEQ ID NO:19) contained the green fluorescent protein (GFP, Clontech, Palo Alto, CA) gene was constructed by inserting a *Bgl*2-*Bcl*1 fragment (800 bp) from plasmid pGFP-N1. This sequence, containing the GFP gene, was treated with mung bean exonuclease and inserted into the unique *Sna* B1 site of pVLSNO3.

15 In order to enhance GFP fluorescence from the reporter plasmid pVLSNOG, the serine-65 codon in the GFP gene was mutated into threonine by a site-directed mutagenesis procedure with the Transformer™ Site-Directed Mutagenesis kit from Clontech. A *Bpm*1 site in the GFP gene (threonine-9) was mutated at the same time without changing the amino acid (ACT to ACA). The resulting plasmid was pVLSNOGM (SEQ ID NO:18).

20 An *Nco*1-*Xho*1 fragment (585 bp) from plasmid pG1IL2EN (kindly provided by Dr. Steven Rosenberg, Bethesda, MD), containing the internal ribosome entry site (IRES) from encephalomyocarditis virus (EMCV) was inserted into the *Apa*1 site upstream of the GFP gene in pVLSNOGM, resulting in pVLSNOGMI (SEQ ID NO:17). Both insert and plasmid fragments were blunted with mung bean exonuclease. One variant version of
25 pVLSNOGMI with an IRES tandem dimer was also constructed and designated pVLSNOGMI2 (SEQ ID NO:16).

Oligonucleotides (SEQ ID NO:53 and 54) containing a splice acceptor (SA) of AKV virus (in bold) was inserted into pVLSNOGMI at the unique *Sac* 2 site just before the IRES, resulting in pVLSNOGMIS (SEQ ID NO:15).

30 Oligo: (SEQ ID NO:53)

5' -GGCCGCT**AACTAATAGCCCATTTCTCCAAGGTACGTAGC**-3'

3' -CGCCGGCGATTGATTATCGGGTAAGAGGTTCCATGCAT-5'

(SEQ ID NO:54, bottom Oligo)

Recovery of LTR promoter sequences from mouse CD4⁺ T-helper cells

5 In order to facilitate the recovery of VL30 promoter sequences expressed in mouse T-helper cells, a mouse CD4⁺ T-helper cell cDNA library (Stratagene, San Diego, CA, Catalog # 937311) was screened by plaque hybridization. Approximately 2×10^4 bacteriophage λ -ZAP clones were plated on a lawn of *E. coli* cells according to the manufacturer's instructions. Two nylon filters were sequentially layered onto the lawn of *E.*
10 *coli* cells and bacteriophage. The filters were hybridized to a ³²P-labelled (Prime-It RmT Random Primer Labeling Kit, Stratagene), 4.2 kb internal *Xho*I fragment of NVL-3 (containing the NVL-3 genome). 55 plaques (or approximately 0.3% of the total phage) reacted positively on both filters. 18 VL30 cDNA sequences were cloned from the plate, which was used to identify U3 promoters that are actively expressed in the RNA of mouse T-
15 cells. Five of the 18 clones contained intact U3 sequences, representing four of one molecular species, named TH1 (SEQ ID NO: 2) and one of another species, named TH2 (SEQ ID NO: 3) also provided in Fig. 5. TH1 contained approximately 120 bp more DNA than did TH2. Because TH1 was more abundant (4 out of 5 clones), the additional sequences in the enhancer region were implicated to be a possible reason for the stronger expression in
20 mouse T cells. Examination of the known and putative transcription factor binding sites in the VL30 LTR (Hodgson, 1996, chapter 4, Fig. 4.2 *supra*) revealed several interesting features of TH1 and TH2. First, the extra sequences of TH1 that were missing in TH2 included an extra copy of the enhancer repeat region as well as a potential retinoid (RAR/RXR) binding site. Several transcription factor binding sites in the enhancer repeat
25 region that differed between the two elements included: a cyclic 3'-5'AMP response element (VLCRE, a potential CREB/jun binding site), a serum response element (SRE), and a potential NF1/IL6 binding site (although there were additional sites for these factors in other enhancer repeats). These factors could possibly explain why VLTH1 appeared to be expressed at higher levels, both in the source cells and into transduced cells. Together, the
30 VL30 sequences represented 0.3% of the mRNA expressed in the T cells, and TH1 appeared to be most abundant VL30.

Sequencing Primers:

(SK, SEQ ID NO:49) 5'-CGCTCTAGAACTAGTGGATC (20 mers, T_m 60°C).(T7, SEQ ID NO:50) 5'-GTAATACGACTCACTATAGGG (21 mers, T_m 60°C).5 **Seamless Rescue of T cell promoters using class IIS restriction enzymes**

Two sets of primers containing offset *Bpm*I restriction sites were designed and synthesized. One set was for amplification of the plasmid sequences, and another was for the amplification of the inserts.

10 **Insert Primers: (*Bpm*I site bold)**ITA (43 mer, T_m: 67.2 °C, SEQ ID NO:45)

CGATCCACTGGAGCTCGGAGCCCACCCCCTCCCATCTAGAGGT

15 ITB (43 mers, T_m: 66.3 °C, SEQ ID NO:46)

CGTCCTCCTGGAGAGCACAGGGTAGAGGAGTCTCGACGGTCAG

Vector primers: (*Bpm*I site bold)VLA (43 mers, T_m: 68.2 °C, SEQ ID NO:47)

CGCAACCCTGGAGACCTCTAGATGGGAGGGGGTGGGCTCCGAG

20 VLB (43 mers, T_m: 66.3 °C, SEQ ID NO:48)

GCAGGACCTGGAGCTGACCGTCGAGACTCCTCTACCCTGTGCT

To amplify vector sequences more efficiently, vector templates were shortened by deleting marker genes from vectors. pVLSNOSIB (SEQ ID NO:14) was cut with *Kpn* I and a 4201 bp fragment containing β-gal gene was removed. The remaining vector has 3923
25 bp.

The U3-promoter inserts (357 bp for TH1 and 240 bp for TH2) were PCR-amplified from TH1 and TH2 promoters with primers ITA and ITB. The vector cassettes (~4.2 kb for pVLSNOSIB and ~3.7 kb for pVLSNOGMIS) were PCR-amplified from the shortened vector templates using primers VLA and VLB, (*supra*). The PCR-amplification
30 was done with high-fidelity *Pfu* DNA polymerase from Stratagene (La Jolla, CA). The amplified products were gel-purified (1% agarose gel). The inserts were then cut with *Bpm* I to produce complementary ends. The vector cassette products were phosphorylated with

PNK, then circularized with T4 ligase, and transformed into SCS 110 cells. Recovered plasmids were then digested with *Bpm* 1 and treated with CIP to produce complementary ends. *Bpm* 1 treated inserts and vector cassettes were ligated, and T-cell tissue-specific VL 30 vectors VLTH1 and VLTH2 were produced. The marker β -gal gene and GFP gene were put
5 back into those vectors at the original unique sites *Kpn* 1 and *Sal* 1 respectively.

Transmission and expression of single LTR vectors and T cell U3 sequences

Vector DNA constructs were transfected into GP+E86 retroviral helper cells (Markowitz et al, 1988, *supra*) using the Lipofectamine protocol (Life Technologies,
10 Gaithersburg, MD). The culture media from these cells (supernatant), containing defective transducing particles (72 h post-transfection), was transmitted to PA317 (Miller, US Patent, cited *supra*) amphotropic helper cells, using Lipofectamine to enhance transduction efficiency (Hodgson *et al.*, 1996. Synthetic Retrotransposon Vectors and Gene Targeting pp. 3-14, in : Felgner et al., eds. *Artificial Self-Assembling Systems for Gene Delivery*. American Chemical
15 Soc. Books, Washington, D.C.). A similar procedure was used to transmit VLTH1 and VLTH2 to the PG13 helper cell line (Miller *et al.*, 1991. *J. Virol.* 65:2220-2224). 24 h post-transfection, the recipient cells were selected with the drug G418 (500 μ g/ml, 2 weeks) to enrich for stably transduced cell populations.

All of the single LTR vectors, including VLTH1 and VLTH2 were transmitted
20 by this method, indicating that single LTR vectors can be used for promoter switching and yet revert to dual LTR vectors after a single passage. Vectors VLSNO2, VLSNO3, and VLSNOSIB were then titered on NIH 3T3 cells (using the PA317 vector producer cell lines). VLTH1 and VLTH2 vectors were titered on human HT1080 cells (PG13 cell lines). Surprisingly, all of the single LTR vectors were transmitted effectively. However the titers of
25 stably transduced TH1 and TH2 cell lines were 5.5×10^2 - 1.1×10^3 TU/ml, compared to 0.4 - 3.0×10^4 TU/ml for the VLSNO2, VLSNO3 and VLSNOSIB cell lines. Thus, switching from the NVL-3 transcriptional promoter (originally isolated from NIH 3T3 fibroblast cells) to VL30 promoters derived from T helper cells, appeared to have a negative effect on RNA expression in fibroblast cells, as determined by the transmissibility of the RNA.

30 In order to study the usefulness of rescued promoters as DNA transfection vectors (as opposed to retro-vectors), VLSNOSIB, VLTH1 and VLTH2 were also transfected into a number of cell lines (using Lipofectamine), including NIH 3T3, PA317, GP+E86,

PG13, HT1080, SW480 and HeLa (available from ATCC). RNA expression in these cell lines is shown in Table 4, wherein gene expression from the LTR promoter (as determined by β -gal staining) is normalized to VLSNOSIB (100).

Cell line:	NIH	PA317	GP+E86	PG13	HT1080	SW480	HeLa
	3T3						
Vector:							
VLSNOSIB	100	100	100	100	100	100	100
VLTH1	39.3	18.7	0.1	21	25.5	156	156
VLTH2	28.6	7.1	5.5	11.5	46.8	82	156

Table 4. Transient expression of a β -gal marker gene by three VL30 promoters: NVL-3 (VLSNOSIB), VLTH1 and VLTH2. Cells were transfected using the Lipofectamine procedure. Total blue cells were counted from each well in 6-well plates, and the number of blue cells from VLSNOSIB was normalized to 100%.

The expression of both the VLTH1 and VLTH2 promoters was significantly reduced compared to VLSNOSIB in cell lines of fibroblastic origin, whereas in SW480 colorectal cancer cells and HeLa cells, it was comparable to or better than VLSNOSIB (the NVL-3 promoter). However, VLSNOSIB was expressed poorly in the non-fibroblastic cell lines, so a direct comparison was difficult to interpret. Unfortunately, the human T cell lines (Jurkat and MOLT4 [obtained from ATCC]) were not transfected by Lipofectamine, and they were poorly transduced by VLTH1 and VLTH2 retro-vectors. In the Jurkat and MOLT4 cells transduced with VLTH1 and VLTH2, only a small percentage (1-10%) of cells that were stably transduced by the vectors stained positively for β -gal expression. However, the marker gene (neo) continued to be expressed from an internal promoter, as evidenced by drug selection.

Taken together, the results demonstrated: 1) the ability of the promoter rescue technique to seamlessly capture functional transcriptional promoters from specialized cells; 2) the ability of single LTR vectors to introduce the rescued promoters into standard transducing vectors; 3) the ability of the rescued promoters to be expressed at differing levels in several different cell types, including T cells; and 4) screening and selection established the efficacy, or lack thereof, of individual promoter sequences.

Although the general method of promoter rescue was demonstrated by the foregoing experiments, the titers obtained from the sLTR VL30 vectors may not be useful where selection systems are not available.

Additional experimentation led to the development of a chimeric packaging signal, combining the essential packaging signal from Moloney murine leukemia virus (Ψ), and the enhanced packaging signal (Ψ^+) from a mouse VL30 element. A vector embodiment of this packaging system is VLMB (SEQ ID NO:23). One advantage of the chimeric packaging system was the elimination of retroviral *gag* gene sequences that were present in previous high-titer MLV-based vectors (viral *gag* sequences contribute to the generation of replication competent retrovirus outbreaks). The titers of VLMB-based vectors ranged from approximately 1×10^5 to 4×10^6 TU/ml.

Construction of a cloning vector for promoter rescue

Using pVLSNOGMIS as a template, and primers (SEQ ID NOS:28 and 68), a 6.4 kb plasmid fragment was PCR amplified (Using Hot Start Ampliwax PCR Gems 100, Perkin Elmer). 30 cycles of PCR were performed by following the manufacturer's instructions, with the following input conditions: lower reaction, 80°C , 5 min., then add upper reaction and template, 96°C , 1 min. Each reaction vial contained 50 ng template, 0.5 μM each primer, 200 μM dNTPs and 5U (2 μl) *Pfu* polymerase (Stratagene, LaJolla, CA). 30 repeating cycles of: 96°C , 45 sec; 50°C , 45 sec; 75°C , 1 min. A final incubation of 75°C , 10 min, then hold at 4°C . After amplification, the reactions were purified using Qiaquick PCR Purification Kits (Qiagen). The PCR products were digested with *Pac1*, heat inactivated (65°C , 20 min) and ligated together using T4 DNA ligase (overnight at 4°C in a 5 μl vol). The ligated DNA was transfected into SCS110 *E. coli* cells (Stratagene) with kanamycin (50 $\mu\text{g/ml}$) antibiotic added to the agar plates. The cells were *dcm*⁻, *dam*⁻ (to prevent methylation of *Bpm1* sites). The resulting plasmid, pVLBPGN (SEQ ID NO:1, Figs 2 &3) has a deletion in the U3 region of the LTR. A linker containing a central *Pac1* site flanked by two outwardly-digesting *Bpm1* sites occupies the site of the deleted U3 sequences. The *Bpm1* sites enable the plasmid to be digested with *Bpm1*, resulting in two 2 bp 3'-overhanging ends that are complementary to the U3-derived RT-PCR inserts described below. The digested plasmid was purified free from the intervening linker sequences from an agarose gel after digestion with *Bpm1*, using the Qiaquick gel purification kit (Qiagen).

Procedure for amplification of liver U3 promoter region

Purified mouse liver total tissue RNA was purchased from Ambion, Inc., (Austin, TX). Total liver RNA was treated with RQ1 Rnase-free (Promega, Madison, WI). Using Perkin Elmer Gene Amp thermostable rTth reverse transcriptase RNA PCR kit (P/N N808-0069), the following conditions for RT-PCR were used: RT-PCR A 70° (hot start); RT-PCR B, 95°C, 60 sec, then 35 cycles (95°C 10 sec, 58°C, 15 sec) then a final 58°C incubation for 7 min, then 4°C and hold. Additional conditions were: primer concentration 0.15 micromolar, template 100 ng/reaction, dNTPs 200 micromolar (final) and MgCL₂ 3.5 mM(final). The primers for insert amplification were SEQ ID NOS:28 and 68)

The amplified U3 sequences were purified using Qiaquick. The pVLBPGN plasmid was digested with *Bpm*1, isolated from a 1% agarose gel and purified using the Qiaquick method. The purified U3 sequences were ligated at 1:2, 1:4 and 1:6 molar ratios of VLBPGN plasmid:insert using T4 DNA ligase and a 5 microliter reaction volume overnight at 4°C (100 ng plasmid: 16 ng insert = 1:1 molar ratio). 1 microliter of each ligation reaction was transformed into *E. coli* SCS 110 competent cells (Stratagene). 26 colonies were recovered in total. Out of 23 clones grown overnight in the presence of kanamycin, 20 had sequences that appeared to be mouse VL30 sequences, representing 10 different VL30 species (Fig. 6, SEQ ID NOS: 4-13). One of these (Hep 10, SEQ ID NO: 13) was transiently transfected into Hep G2 liver hepatocellular carcinoma cells. 48 h after transfection, intense GFP fluorescence was observed, indicating strong expression of the Hep 10 U3 promoter region.

Example 4

Creating a combinatorial library of mouse VL30 U3 sub-regions.

Using Fig. 7 and Hodgson, 1996, supra, Fig. 4.2 as a guide, the following three sub-regions of the VL30 U3 region were empirically established: Distal (1); medial (2); and proximal (3). Peaks of similarity were used to guide the following choice of primers: (+) primer binding site-5'-LTR boundary; ~80 bp (defines sub-region 1); ~80-210 bp (sub-region 2); ~210-430 (sub-region 3). The following primers were selected to amplify the vector VLBPGN or a similar VL30, NVL-3 LTR-containing vector:

P1 (going left from the 5'-end of the LTR to amplify the plasmid)

(SEQ ID NO:55)

GACTAACCTTGATTCCACTGGAGTTTT(CT)(CT)ATTCTTCATTCCCCACTTC
TTCTT

P2 (going right from the 3'-end of the promoter region to amplify the plasmid)

5 (SEQ ID NO:56)

GACTAACCTTGATTCCACTGGAGAATCTGGACCAATTCTATATAAGCCTG
TGAAAAATTT

The six primers selected to amplify the inserts are as follows:

10 Fragment 1, primer 1 (going right from the LTR terminus into U3) (SEQ ID NO:57)

GACTAACCTTGATTCCACTGGAGAAGAAGAAGTGGGGAATGAAGAA

Fragment 1, primer 2 (going left from the end of fragment 1) (SEQ ID NO:58)

GACTAACCTTGATTCCACTGGAGATCTCTAGATGGGAGGGG(GT)(CT)GGG
CTC

15 Fragment 2, primer 1 (going right from the left end of fragment 2) (SEQ ID NO:59)

GACTAACCTTGATTCCACTGGAGCTCGGAGCCCACCCCTCCCATCT

Fragment 2, primer 2 (going left from the right end of fragment 2) (SEQ ID NO:60)

GACTAACCTTGATTCCACTGGAGGGAGGCCCTTATCTCAAAAATGTT

Fragment 3, primer 1 (going right from the left end of fragment 3) (SEQ ID NO:61)

20 GACTAACCTTGATTCCACTGGAGTCTAAGAACATTTTGTAGATAAGGGCC
T

Fragment 3, primer 2 (going left from the right end of fragment 3) (SEQ ID NO:62)

GACTAACCTTGATTCCACTGGAGTCACAGGCTTATATAG(TG)AAA

25 100 ng of genomic DNA from *Mus musculus* is used as a template (the mouse genome bears 100-200 copies of VL30 elements). Standard PCR procedures for *Pfu* polymerase are used. Fragments are amplified 35 rounds of PCR to obtain single-copy genomic DNA amplification. Samples of Qiagen column purified DNA are examined on analytical agarose gels to determine the approximate size. The remainder of each reaction is digested with the
30 appropriate enzyme and run on an acrylamide or agarose gel. The digested fragments are purified by standard gel purification procedures and are ligated to the plasmid fragment at an equimolar ratio of the four PCR fragments (three inserts and one plasmid). The ligation mix

is transformed into *E. coli* SCS1 and is grown on kanamycin. The number of colonies is used to establish the size of the combinatorial library, and the pooled colonies are grown in *E. coli* and the DNA is harvested *en masse*. A dozen or more colonies are characterized by DNA sequencing to determine the approximate fidelity of the reaction. A library of 1,000 or more, but preferably 100,000 or more members is used for combinatorial screening procedures.

Screening the combinatorial libraries for expression in specific cell types using a replication defective helper virus

The U3 library DNA is transfected into the desired target cells in which expression is desired. Along with the library, approximately 25% of the total DNA should include retroviral helper sequences. The latter sequences can be a helper plasmid (such as pPAM3, Miller *et al.*, US Patent 4,861,719). The virus is amphotropic, permitting it to infect most human cells. The RNA from individual clones that are transcribed in the target cells will be packaged into retroviral virions made by the helper virus, and the virions can be harvested as the cell free filtrate (0.45 mm) from the vector producer cells. This virus (containing the expressed sequences) can be transmitted to fresh target cells that do not contain helper virus. 48 hours after passage, the DNA form of the transcriptionally active clones will be integrated in the recipient cells, and these transcriptionally active loci will produce more RNA, and protein. After G418 drug selection to increase the proportion of cells expressing the vector sequences, helper virus DNA is again transfected into the recipient cells, transforming them into vector producer cells. The virus from these cells should contain increased amounts of the RNA from clones that are transcriptionally active in those cells. Passage of the virus is continued for two or three rounds to permit recombination and mutation to take place, enhancing the effect of *in vitro* evolution of promoters. The actual degree of enhancement attainable at each step is illustrated in Table 2 (*supra*). After several passages, the actual level of RNA expressed by several clones is determined by RNA blotting, or by the amount of a reporter gene expressed as protein (determined visually or by the appropriate assay). Because human cells do not naturally contain VL30 DNA or RNA, the sequences that remain in the human cells are those with the most transcriptionally active promoters. These sequences can be amplified and re-cloned using the methods of the instant invention, or they can be rescued by virus packaging, reverse transcribed by the endogenous reverse

transcriptase reaction, and grown as plasmids (due to their plasmid origin of replication and the selectable kanamycin marker gene).

In addition to using a replication defective helper virus, such as the clone pPAM3, it is also possible to use a replication competent retrovirus, such as Moloney murine leukemia virus to passage the library. For use in human cells, however, the virus should have a tropism that is compatible with human cells (gibbon ape leukemia virus and amphotropic [4070A] murine retroviruses are acceptable).

In addition to being useful for generating active transcriptional promoters *de novo*, a small variation on the above procedures may enable the isolation of hormone responsive promoters. In it, the cells are treated with the hormone (which could be a steroid, a peptide hormone known to affect the cells, a drug, a drug agonist or antagonist, etc.) during passage. After isolation of surviving VL30 vector-containing cells, individual clones of drug resistant cells are tested for reporter gene expression with and without drug treatment to determine relative protein expression. Likewise, RNA expression can be determined by blot analysis or a similar method. A useful list of known VL30 responses to pharmacological agents is listed in Fig. 4.2 of Hodgson, 1996, *supra*, and can be used as a guide to help assess the potential agents known to have an effect on VL30 transcription.

Once the transcriptional promoters with the known specificity have been obtained, they can be used to obtain expression of genes from a variety of types of vectors. For example, in addition to retrovirus particles, the promoters can be incorporated into all other major groups of vectors: adenoviruses, herpes simplex virus vectors, DNA transfection vectors, etc. It will be apparent to persons of ordinary skill in the art that similar combinatorial libraries can also be used to screen for other characteristics than transcription activity in a particular cell. For example, combinatorial libraries of complementarity determining regions (CDRs) of antibodies or T cell receptors can be so screened using antibody screening methods, such as the phage display screening method (Pharmacia, Milwaukee, WI). Thus, the methods of this invention, particularly the combinatorial simplicity of this invention is a significant improvement over many *in vivo* recombination methods including those of (Stemmer, US Patent 5,605,793; 1997) that have described for the production of CDR combinatorial libraries.

Example 5 Gene Assembly Line

5 From the above examples of 3 and 6 fragment gene self-assemblies, it is evident that assembly of genes by means of gene amplification, the use of offset restriction enzymes and incorporating unique, non-palindromic ends is a highly efficient process compared to conventional cloning methods. However, in addition to the considerations already discussed, it will be apparent to a person of ordinary skill in the art that the various
10 procedures, protocols, methods and material of the instant invention become more difficult to use as the number of fragments increases. For example, if the efficiency of combining each fragment in an assemblage is 99%, then the overall efficiency of combining ten fragments will be 90%, the efficiency of combining 100 fragments will be 37%, etc. Therefore, a small drop in efficiency of any step or fragment, or a large increase in the complexity of the project,
15 will be sufficient to reduce the overall efficiency. Fastidious procedures permit one to achieve success with more complex projects.

 Foremost in its potential for inducing failure is human error in primer design where large numbers of fragments are used. Fortunately, the instant invention is suited to automation of most of the steps. This allows human input to be focused on design, analysis,
20 and quality control. For the purposes of generating large vectors or chromosomes, it is desirable to provide an automated environment. One method to achieve this goal is a gene assembly line.

 In a gene assembly line, multiple tasks are controlled by a machine or machines working together to increase speed and efficiency and to reduce human error. For
25 example, computer aided design (CAD) and computer aided manufacturing (CAM) are incorporated and combined with the methods of this invention. The computers accept inputs in the form of template and primer sequences, together with preferences of regions to be copied and joined. The preferences include at least the sequences of the primer regions and information about the known restriction sites and maps of the sequences to be assembled, but
30 ideally include the entire sequence. The preferences also include the number of sequences to be joined, the desired T_m for the primers, the list of potential restriction enzymes capable of offset digestion that are potential candidates for use in the assembly process, the desired end structures for each fragment terminus, a tag sequence (if any), whether circular or linear ends

are desired, and additional design considerations. The computer algorithm then searches the sequences to determine the candidate enzymes and specific primers that match the criteria of the input. Candidates for selection of unique non-palindromic overlaps are selected. The computer then posts selections or preferences for the type and order of end structures, the primer binding sites, their potential for primer-dimer and intra-molecular interaction artifacts, and the potential conflicts with repeat sequences within the templates that could lead to incorrect polymerization. Based upon the selections made by the operator, the computer then determine the T_m for each primer, and makes adjustments (with suitable inputs from the investigator) to achieve a suitable T_m for the appropriate DNA synthesis or gene amplification reaction. Ideally, the primers should have similar T_m s so that all amplification reactions can be performed at once with one set of amplification instructions. In reality, it may be difficult to do this with complex projects. The output of this portion of the program, which can be in a generic format, such as a Microsoft Excel spreadsheet is then downloaded to a computerized oligonucleotide synthesizer, such as the Applied Biosystems 3928 nucleic acid synthesizer. One advantage of using a computerized synthesizer is its robotic capability to de-protect and purify the oligonucleotides automatically. In addition this synthesizer can accept computerized input.

The quantity of individual oligos recovered is then determined spectrophotometrically. It is desirable to purify the oligonucleotides by high performance liquid chromatography or by polyacrylamide gel. In a preferred embodiment, the oligonucleotides and templates are then assembled robotically using an automated nucleic acid handling system such as the Qiagen BioRobot 9600. The BioRobot is capable of accepting input from a computer and can combine the gene amplification reactions based upon the assignments of templates, primer and reagents provided in the input. The assembled reactions are then amplified for example by PCR. In a preferred embodiment, the PCR heat block is incorporated into the robotic workspace and genes are assembled robotically but with minimal human intervention to change buffers, rearrange the platform, change programs, and the like. The resulting amplified products are also purified by the BioRobot or a similar robotic device. In a preferred embodiment, the robotic device uses Qiaquick cleanup procedures, or a similar method and then assembles restriction endonuclease reactions to digest the purified gene amplification products. The gene amplification products are loaded onto a gel and electrophoresed. Human intervention may be necessary to analyze the

products and excise the correct fragments from the gel. At this point, the results are assessed and missing or incorrect sized fragments are resynthesized. The robotic device is preferably used to purify the gel fragments using Quiagen or similar cleanup procedures. After spectrophotometric quantitation of the purified fragments, the robotic device is preferably
5 used to assemble the ligation. Ideally the fragments are combined in an equimolar ratio of 1:1. However it is not necessary to use equimolar ratios in order to achieve gene self-assembly. For automated gene assembly, it may be desirable not to use equimolar ratios of input fragments, particularly if it simplified the task of quantitation. After ligation, the assemblies can be purified and ethanol precipitated or they can be added to the appropriate
10 host cells. Automation aids in maintaining the sterility of the reaction.

Several additional considerations can assist in the construction of long genes using gene assembly. First the number of fragments and the length of constructs are limiting factors. In addition to maintaining high standards of purify of both the oligonucleotide primers and gene amplification products, it is important to keep the error rate low during
15 copying. Thus, one can optimally start with 100 ng of template use only five rounds of gene amplification and finish with nearly 2 micrograms of product. This is more desirable for reducing errors than using a large number of amplification steps. It is also desirable to use a special copying enzyme such as *Pfu* DNA polymerase that has a low intrinsic error rate. Further it is desirable to use *in vivo* selection (in eukaryotic cells or tissues) rather than *E. coli*
20 cloning to reduce the incorporation of errors into the vectors. For example, a viral vector such as an adenoviral vector or the retro-vectors of the preceding examples are auto-selecting. A single correctly-assembled adenovirus vector molecule, for example, leads to a lytic infection (the viral products of which are cloned by limiting dilution on the appropriate eukaryotic cells), even though it may be combined in a ligation mix with a large excess of
25 incorrectly assembled molecules that are non-functional. Thus, it is not necessary to have a high efficiency, although high efficiency has been demonstrated in this system, in order to achieve success in making, for example gene therapy vectors.

For long fragments (3-30 kb), it is desirable to use enzymes and procedures that are designed or facilitate replication of long fragments, one such example is the
30 eLONGase system (Life Technologies). This system can copy up to 30 kb on a fragment with proofreading. Considerations for long PCR are reviewed in Beck, 1998. (The Scientist 6 Janary, 1998, pp. 16-18).

Internal restriction sites are a potential problem, particularly with large constructs and can be overcome in a number of ways. Use of alternate enzymes, methylation of internal restriction sites (such as by using methylated DNA precursors during synthesis to leave the sites in primers unaffected, incorporation of the internal sites into the construct (if they are non-palindromic), or mutagenesis of internal sites, are exemplary ways to deal with some of these issues.

For very large constructs, it is desirable to use enzymes such as *SapI* (recognizing 7 nucleotides and leaving a 3 bp overhang). This enzyme digests every 16,384 bp on average. There are 64 nucleotide triplet combinations, meaning that up to 32 fragments can be ligated in a circle using *SapI*. *FokI* and *HgaI* are other examples of class IIS enzymes that are useful for making large constructs. *HgaI* has 5 bp overhangs, permitting more than 500 *HgaI* fragments to be ligated. *FokI* includes a Kozak ATG start codon. In a preferred embodiment, a *FokI* site is inserted at the PuXXATG start site of a cDNA encoding region. The cDNA is inserted in frame, providing a site for inserting and switching coding sequences within a vector.

It will be readily understood by those skilled in the art that the foregoing description has been for purposes of illustration only and that a variety of embodiments can be envisioned without departing from the scope of the invention. Therefore, it is intended that the invention not be limited except by the claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5

(i) APPLICANT: NATURE TECHNOLOGY CORPORATION, ET AL.

(ii) TITLE OF INVENTION: SELF-ASSEMBLING GENES, VECTORS AND USES THEREOF

10

(iii) NUMBER OF SEQUENCES: 68

(iv) CORRESPONDENCE ADDRESS:

15

(A) ADDRESSEE: MUETING, RAASCH & GEBHARDT, P.A.
(B) STREET: 119 NORTH FOURTH STREET, SUITE 203
(C) CITY: MINNEAPOLIS
(D) STATE: MINNESOTA
(E) COUNTRY: USA
(F) ZIP: 55401

20

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: Not Assigned
(B) FILING DATE: 28-FEB-1998
(C) CLASSIFICATION:

30

(vii) PRIORITY APPLICATION DATA:

(A) APPLICATION NUMBER: 60/070,910
(B) FILING DATE: 28-FEB-1997
(C) CLASSIFICATION:

35

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: MCCORMACK, MYRA M.
(B) REGISTRATION NUMBER: 36,602
(C) REFERENCE/DOCKET NUMBER: 228.00010201

40

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 612-305-1225
(B) TELEFAX: 612-305-1228

45

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

50

(A) LENGTH: 6225 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

55

(ii) MOLECULE TYPE: DNA (genomic)

60

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TGAAGAATAA AAAATTACTG GCCTCTTGTG AGAACATGAA CTTTCACCTC GGAGCCCACC 60

CCCTCCCATC TGGAAAACTC CAGTTATAAC TGGAGTTTTT CCTTTAAAAG CTTGTGAAAA 120
65 ATTTGAGTCG TCCTCGAGAC TCCTCTACCC TGTGCAAAGG TGTATGAGTT TCGACCCCAG 180

	AGCTCTGTGT GCTTTCTGTT GCTGCTTTAT TTCGACCCCA GAGCTCTGGT CTGTGTGCTT	240
	TCATGTGCGT GCTTTATTAA ATCTTACCTT CTACATTTTA TGTATGGTCT CAGTGTCTTC	300
5	TTGGGTACGC GGCTGTCCCG GGACTTGAGT GTCTGAGTGA GGGTCTTCCC TCGAGGGTCT	360
	TTCATTTGGT ACATGGGCCG GGAATTGCAG AATCTTTCAT TTGGTGCATT GGCCGGGAAT	420
10	TCGAAAATCT TTCATTTGGT GCATTGGCCG GGAAACAGCG CGACCACCCA GAGGTCCTAG	480
	ACCCACTTAG AGGTAAGATT CTTTGTCTG TTTTGGTCTG ATGTCTGTGT TCTGATGTCT	540
	GTGTTCTGTT TCTAAGTCTG GTGCGATCGC AGTTTCAGTT TTGCGGACGC TCAGTGAGAC	600
15	CGCGCTCCGA GAGGGAGTGC GGGGTGGATA AGGATAGACG TGTCCAGGTG TCCACCGTCC	660
	GTTCCGCCCTG GGAGACGTCC CAGGAGGAAC AGGGGAGGAT CAGGGACGCC TGGTGGACCC	720
20	CTTTGAAGGC CAAGAGACCA TTTGGGGTTG CGAGATCGTG GGTTCGAGTC CCACCTCGTG	780
	CCCAGTTGCG AGATCGTGGG TTCGAGTCCC ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT	840
	CGAGTCCAC CTCGCGTCTG GTCACGGGAT CGTGGGTTTC AGTCCCACCT CGTGTTTTGT	900
25	TGCGAGATCG TGGGTTTCGAG TCCCACCTCG CGTCTGGTCA CGGGATCGTG GGTTCGAGTC	960
	CCACCTCGTG CAGAGGGTCT CAATTGGCCG GCCTTAGAGA GGCCATCTGA TTCTTCTGGT	1020
30	TTCTCTTTTT GTCTTAGTCT CGTGTCGCT CTTGTTGTGA CTA CTGTTTT TCTAAAAATG	1080
	GGACAATCTG TGTCCACTCC CCTTCTCTG ACTCTGGTTC TGTCGCTTGG TAATTTTGT	1140
	TGTTTACGTT TGTTTTGTG AGTCGTCTAT GTTGTCTGTT ACTATCTTGT TTTTGTGTT	1200
35	GGTTTACGGT TTCTGTGTGT GTCTTGTGTG TCTCTTTGTG TTCAGACTTG GACTGATGAC	1260
	TGACGACTGT TTTTAAGTTA TGCCTTCTAA AATAAGCCTA AAAATCCTGT CAGATCCCTA	1320
40	TGCTGACCAC TTCCTTTCAG ATCAACAGCT GCCCTTACTC GAGCTCAAGC TTCGAATTCT	1380
	GCAGTCGACG GTACCGCGGC CGCTAACTAA TAGCCCATTG TCCAAGGTAC GTAGCGGGGA	1440
	TCAATTCCGC CCCCCCCTA ACGTTACTGG CCGAAGCCGC TTGGAATAAG GCCGGTGTGC	1500
45	GTTTGTCTAT ATGTTATTTT CCACCATATT GCCGTCTTTT GGCAATGTGA GGGCCCGGAA	1560
	ACCTGGCCCT GTCTTCTTGA CGAGCATTCC TAGGGGTCTT TCCCCTCTCG CCAAAGGAAT	1620
50	GCAAGGTCTG TTGAATGTCG TGAAGGAAGC AGTTCCTCTG GAAGCTTCTT GAAGACAAAC	1680
	AACGTCTGTA GCGACCCTTT GCAGGCAGCG GAACCCCCCA CCTGGCGACA GGTGCCTCTG	1740
	CGGCCAAAAG CCACGTGTAT AAGATACACC TGCAAAGGCG GCACAACCCC AGTGCCACGT	1800
55	TGTGAGTTGG ATAGTTGTGG AAAGAGTCAA ATGGCTCTCC TCAAGCGTAT TCAACAAGGG	1860
	GCTGAAGGAT GCCCAGAAGG TACCCCATTG TATGGGATCT GATCTGGGGC CTCGGTGCAC	1920
60	ATGCTTTACA TGTGTTTAGT CGAGGTTAAA AAAACGTCTA GGCCCCCGA ACCACGGGGA	1980
	CGTGGTTTTT CTTTGAAAAA CACGATACGG GATCCACCGG TCGCCACCAT GGGTAAAGGA	2040
	GAAGAACTTT TCACAGGAGT TGTCCCAATT CTTGTTGAAT TAGATGGTGA TGTTAATGGG	2100
65	CACAAATTTT CTGTCAGTGG AGAGGGTGAA GGTGATGCAA CATACGGAAA ACTTACCCTT	2160

	AAATTTATTT	GCACTACTGG	AAAACCTACCT	GTTCCATGGC	CAACACTTGT	CACTACTTTC	2220
	ACTTATGGTG	TTCAATGCTT	TTCAAGATAC	CCAGATCATA	TGAAACGGCA	TGACTTTTTTC	2280
5	AAGAGTGCCA	TGCCCCAAGG	TTATGTACAG	GAAAGAACTA	TATTTTTC	AAAGATGACGGG	2340
	AACTACAAGA	CACGTGCTGA	AGTCAAGTTT	GAAGGTGATA	CCCTTGTTAA	TAGAATCGAG	2400
10	TTAAAAGGTA	TTGATTTTAA	AGAAGATGGA	AACATTCTTG	GACACAAATT	GGAATACAAC	2460
	TATAACTCAC	ACAATGTATA	CATCATGGCA	GACAAACAAA	AGAATGGAAC	CAAAGTTAAC	2520
	TTCAAAATTA	GACACAACAT	TGAAGATGGA	AGCGTTCAAC	TAGCAGACCA	TTATCAACAA	2580
15	AATACTCCAA	TTGGCGATGG	CCCTGTCCTT	TTACCAGACA	ACCATTACCT	GTCCACACAA	2640
	TCTGCCCTTT	CGAAAGATCC	CAACGAAAAG	AGAGACCACA	TGGTCCTTCT	TGAGTTTGT	2700
20	ACAGCTGCTG	GGATTACACA	TGGCATGGAT	GAAGTATACA	AGTCCGGATC	TAGATAACTG	2760
	TATCGATGGA	TCCGAAGGCG	GGGACAGCAG	TGCAGTGGTG	GACAGAAAGC	AAGTGATCTA	2820
	GGCCAGCAGC	CTCCCTAAAG	GGACTTCAGC	CCACAAAGCC	AAACTTGTGG	CTTTAATACA	2880
25	AGCTCTGTAA	ATGGTAAAA	AAAAAAGTC	TACACGGACA	GCAGGTATGC	TCTTGCCACT	2940
	GTACAGAGCA	ATATACAGAC	AAAGAGAACT	GTTGACATCT	GCAGAGAAAG	ACCTAAGATG	3000
30	CTGTGGCTAA	AAGAAATCAG	ATGGCAAATC	TAACCGCCCA	GGCATCCTAA	AGAGCAATGA	3060
	TCCTGACAGT	CTGAAGACTA	TCAAGTTATA	GACAAATTAA	GACTGGTAAA	AAAAACCCTG	3120
	TATAAAATAG	TAAAACTGA	AAAAAGAAAA	CTAGTCCTCT	CATGAGAAGA	CAGACCTGAC	3180
35	ATCTACTGAA	AAATAGACTT	TACTGGAAAA	AATATGTGTA	TGAATACCTT	CTAGTTTTTG	3240
	TGAACGTTCT	CAAGATGGAT	AAAAGCTTTT	CCTTGTAATA	CGAGACTGAT	CAGATAGTCA	3300
40	TCAAGAAGAT	TGTTAAAGAA	AATTTTCCAA	GGTTCGGAGT	GCCAAAAGCA	ATAGTGTCAG	3360
	ATAATGGTCC	TGCCTTTGTT	GCCCAGGTAA	GTCAGGGTGT	GGCCAAGTAT	TTAGAGGTCA	3420
	AATGAAAATT	CCATTGTGTG	TACAGACCTC	AGAGCTCAGG	AAAGATAAAA	AAGAATAAAT	3480
45	AAAACCTCTA	ACAGACCTTG	ACAAAATTAA	TCCTAGAGAC	TGGCAGAGAC	TTACTTGGTA	3540
	CTCCTTCCCC	TTGCCCTATT	TAGAACTGAG	AATACTCCCT	CTTGATTCGG	TTTTACTCTT	3600
50	TTTAAGATCC	TTTATGGGGC	TCCTATGCCA	TCACTGTCTT	AAATGATGTG	TTTAAACCTA	3660
	TGTTGTTATA	ATAATGATCT	ATATGTTAAG	TTAAAAGGCT	TGCAGGTGGT	GCAGAAAGAA	3720
	GTCTGGTCAC	AACTGGCTAC	AGTGAACAAG	CTGGGTACCC	CAAGGACATC	TTACCAGTTC	3780
55	CAGCCAGAGA	TCTGATCTAC	GATCCCCGGG	TCGACCCGGG	TCGACCCTGT	GGAATGTGTG	3840
	TCAGTTAGGG	TGTGGAAGT	CCCCAGGCTC	CCCAGCAGGC	AGAAGTATGC	AAAGCATGCA	3900
60	TCTCAATTAG	TCAGCAACCA	GGTGTGGA	GTCCCCAGGC	TCCCCAGCAG	GCAGAAGTAT	3960
	GCAAAGCATG	CATCTCAATT	AGTCAGCAAC	CATAGTCCCG	CCCCTAACTC	CGCCCATCCC	4020
	GCCCCTAACT	CCGCCAGTT	CCGCCATTTC	TCCGCCCAT	GGCTGACTAA	TTTTTTTTAT	4080
65	TTATGCAGAG	GCCGAGGCCG	CCTCGGCCCTC	TGAGCTATTC	CAGAAGTAGT	GAGGAGGCTT	4140
	TTTTGGAGGC	CTAGGCTTTT	GCAAAAAGCT	TCACGCTGCC	GCAAGCACTC	AGGGCGCAAG	4200

	GGCTGCTAAA GGAAGCGGAA CACGTAGAAA GCCAGTCCGC AGAAACGGTG CTGACCCCGG	4260
	ATGAATGTCA GCTACTGGGC TATCTGGACA AGGGAAAACG CAAGCGCAAA GAGAAAGCAG	4320
5	GTAGCTTGCA GTGGGCTTAC ATGGCGATAG CTAGACTGGG CGGTTTTATG GACAGCAAGC	4380
	GAACCGGAAT TGCCAGCTGG GGCGCCCTCT GGTAAGGTTG GGAAGCCCTG CAAAGTAAAC	4440
10	TGGATGGCTT TCTTGCCGCC AAGGATCTGA TGGCGCAGGG GATCAAGATC TGATCAAGAG	4500
	ACAGGATGAG GATCGTTTCG CATGATTGAA CAAGATGGAT TGCACGCAGG TTCTCCGGCC	4560
	GCTTGGGTGG AGAGGCTATT CGGCTATGAC TGGGCACAAC AGACAATCGG CTGCTCTGAT	4620
15	GCCGCCGTGT TCCGGCTGTC AGCGCAGGGG CGCCCGGTTT TTTTGTCAA GACCGACCTG	4680
	TCCGGTGCCC TGAATGAACT GCAGGACGAG GCAGCGCGGC TATCGTGGCT GGCCACGACG	4740
20	GGCGTTCCTT GCGCAGCTGT GCTCGACGTT GTCAGTGAAG CGGGAAGGGA CTGGCTGCTA	4800
	TTGGGCGAAG TGCCGGGGCA GGATCTCCTG TCATCTCACC TTGCTCCTGC CGAGAAAGTA	4860
	TCCATCATGG CTGATGCAAT GCGGCGGCTG CATACGCTTG ATCCGGCTAC CTGCCCATTG	4920
25	GACCACCAAG CGAAACATCG CATCGAGCGA GCACGTAATC GGATGGAAGC CGGTCTTGTC	4980
	GATCAGGATG ATCTGGACGA AGAGCATCAG GGGCTCGCGC CAGCCGAATC GTTCGCCAGG	5040
30	CTCAAGGCGC GCATGCCCGA CGGCGAGGAT CTCGTCGTGA CCCATGGCGA TGCCTGCTTG	5100
	CCGAATATCA TGGTGGAAAA TGGCCGCTTT TCTGGATTCA TCGACTGTGG CCGGCTGGGT	5160
	GTGGCGGACC GCTATCAGGA CATAGCGTTG GCTACCCGTG ATATTGCTGA AGAGCTTGGC	5220
35	GGCGAATGGG CTGACCGCTT CCTCGTGCTT TACGGTATCG CCGCTCCCGA TTCGCAGCGC	5280
	ATCGCCTTCT ATCGCCTTCT TGACGAGTTC TTCTGAGCGG GACTCTGGGG TTCGAAATGA	5340
40	CCGACCAAGC GACGCCCAAC CTGCCATCAC GAGATTTCTG TTCCACCGCC GCCTTCTATG	5400
	AAAGGTTGGG CTTGCGAATC GTTTTCCGGG ACGGAATTCT TAATCTGCTG CTTGCAAACA	5460
	AAAAAACCAC CGCTACCAGC GGTGGTTTGT TTGCCGGATC AAGAGCTACC AACTCTTTTT	5520
45	CCGAAGGTAA CTGGCTTCAG CAGAGCGCAG ATACCAAATA CTGTCCTTCT AGTGAGCCG	5580
	TAGTTAGGCC ACCACTTCAA GAACTCTGTA GCACCGCCTA CATACTCGC TCTGCTAATC	5640
50	CTGTTACCAG TGGCTGCTGC CAGTGGCGAT AAGTCGTGTC TTACCGGGTT GGACTCAAGA	5700
	CGATAGTTAC CGGATAAGGC GCAGCGGTCG GGCTGAACGG GGGGTTCGTG CACACAGCCC	5760
	AGCTTGAGC GAACGACCTA CACCGAATG AGATACCTAC AGCGTGAGCA TTGAGAAAGC	5820
55	GCCACGCTTC CCGAAGGGAG AAAGGCGGAC AGGTATCCGG TAAGCGGCAG GGTGGAACA	5880
	GGAGAGCGCA CGAGGGAGCT TCCAGGGGGA AACGCCTGGT ATCTTTATAG TCCTGTCGGG	5940
60	TTTCGCCACC TCTGACTTGA GCGTCGATTT TTGTGATGCT CGTCAGGGGG GCGGAGCCTA	6000
	TGGA AAAACG CCAGCAACGC CGAGATGCGC CGCCTCGAGT ACACCTGCGT CATGCTGAGA	6060
	CCCTCAAGCC TCACTAAAAG GGTCCCTGCC TAGTTCTGTT TACTAATCTG CCTTATCTG	6120
65	TTTTGTTC CATGTTAAAG ATAGAGTAAA TGCAGTATTC TCCACATAGA GATATAGACT	6180

TCTGAAATTC TAAGATTAGA ATTATTTACA AGAAGAAGTG GGGAA

6225-

(2) INFORMATION FOR SEQ ID NO:2:

- 5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 487 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
10 (D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

CCTCCCATCT AGAGGTTGTT CTCGGAACAC TCCTAAACTT TTCACCCCAA AACTCCTCAC 60
20 CCTAAAGTTC GAAAAAAGTCT TTCCAAGAAC ATTTTGTAGA TAAAGGCCTC CTAGAACAAC 120
CTCAAAATGA CATTGCCAAA TGATAAGACA TGACTCCTTA GTTACGTAGG TTCCTTGATA 180
GGACATGACT CCTTAGTTAC GTAGGTTCTT TGATAGGACA TGACTCCTTA GTTACGTAGA 240
25 TTCCTTTGGT AGAACTCCCT AGTGATGTAA ACTTGTACTT TCCCTGCCCA GTTCTCCCCC 300
TTTGAGTTTT ACTATATAAG CCTGTAAAAA ATTTTGTCTG ACCGTGCGAGA CTCCTCTACC 360
30 CTGTGCTAAG GTGTATGAGT TTCGACCCCA GAGCTCTGTG TGCTTCCATG TTGCTGCTTT 420
ATTTCGACCC CAGAGCTCTG GTCTGTGTGC TTTCATGTGC CTGCTTTATT AAATCTTGCC 480
TTCTACA 487

(2) INFORMATION FOR SEQ ID NO:3:

- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 366 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

50 CCTCCCATCT AGAAAACATT TTTGAGATAA AGGCTTCCTG GAACAACCTC AAAATGAACC 60
AGGTACTCCT TAGTTACGTA GGTTCCTTGA TAGGACATGA CTCCTTAGTT ACATAGATTC 120
55 CTTTGGCAGA ACTCCCTAGT GATGTAAACT TGTACTTTCC CTGCCCAGTT CTCCCCCTTT 180
GAGTTTTACT ATATAAGCCT GTGAAAAATT TTGGCTGACC GTCGAGACTC CTCTACCCTG 240
TGCTAAGGTG TATGAGTTTC GACCCAGAG CTCTGTGTGC TTCCATGTTG CTGCTTTATT 300
60 TCGACCCAG AGCTCTGGTC TGTGTGCTTT CATGTTGCTG CTTATTATAA TCTTGCCCTC 360
TACATT 366

(2) INFORMATION FOR SEQ ID NO:4:

- 65 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CCTCCCATCT AGAGATTGTT CCCAGAACAC TCCTGAACTC TTCACCCAG AATGCATGCC 60
15 TGAAGTCCTC ACCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCGGG TACATTGCCA AATAATAGGA 180
CATGACCCCT TAGTTACGTA AAATCCCTTG GCAGAACCCC TTGTCCCTTG GCAGAACCCC 240
20 TTAGTTATGT AAAGTTGTAC TTTCCCTACC CCGCTCTCCC CCCTTGAGTT TTTCTATAT 300
AAGC 304

25 (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 base pairs
(B) TYPE: nucleic acid
30 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

40 CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTGAACTC TTCACCCAG AATGCATTCC 60
TGAAGTCCTC ACCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCGGA TACATTGCCA AATAATAGGA 180
45 CATGACCCCT TAGTTACGTA GAATCCCTTG GCAGAACCCC TTGTCCCTTG GCAGAACCCC 240
TTAGTTATGT AAAGTTGTAC TTTCCCTACC CCGCTCTCCC CCCTTGAGTT TTTCTATAT 300
50 AAGC 304

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 base pairs
55 (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

60 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

65 CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTGAACTC TTCACCCAG AATGCATTCC 60

5 TGAACTCCTC ATCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCTGG TACATTGCCA AATAATAGGA 180
CATGACCCTT TAGTTACGTA GAATCCCTTG GCAGAACCCC TTGTCCCTTG GCAGAACCCC 240
TTAGTTATGC AAACCTTGTA TTTCTCTGCC CCGCTCTCCC CCCTTGAGTT TTTCTATAT 300
10 AAGC 304

(2) INFORMATION FOR SEQ ID NO:7:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 304 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
20 (ii) MOLECULE TYPE: DNA (genomic)

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTGAACTC TTCACCTCAA AATGCATTCC 60
30 TGAACTCCTC ACCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCAGG TACATTGCCA AATAATAGGA 180
CATGACCCTT TAGTTACGTA GAATCCCTTG GCAGAACCCC TTGTCCCTTG GCAGAACCCC 240
35 TTAGTTATGC AAACCTTGTA TTTCTCTGCC CCGCTCTCCC CCCTTGAGTT TTTCTATAT 300
AAGC 304

40 (2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 305 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
45 (D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

55 CCTCCCATCT AGAGATTGTT CCCAGAACAC TCCTGAACTC TTCACCCAG AATGCATTCC 60
TGAACTCCTC ACCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCGGA TACATTGCCA AATAATAGGA 180
60 CATGACCCTT TAGTTACGTA GAATCCCTT GGCAGAACCC CTTGTCCCTT GGCAGAACCC 240
CTTAGTTATG CAAACTTGTA CTTTCCTGC CCGCTCTCC CCCCTTGAGG TTTCTATA 300
TAAGC 305

65

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 305 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

15 CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTGAACTC TTCACCCCAG AATGCATTCC 60
TGAACCCCTC ACCCTAGAGT TCGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCAGG TACATTGCCA AATAATAGGA 180
20 CATGACCCCT TAGTTACGTA GAATTCCCTT GGCAGAACCC CTTGTCCCTT GGCAGAACCC 240
CTTAGTTATG CGAACTTGTA CTTTCCCTGC CCCGCTCTCC CCCCTTGAGT TTTTCCTATA 300
25 TAAGC 305

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 306 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

40 CCCTCCCATC TAGAGAGTGT TCCCAGAACA CTCCTGAACT CTTTCATCCA GAATGCATTC 60
CTGAACTCCT CACCCATAG TTCGAACCCT CCCAACTAAA GACTGTTCCA AGAACATTTT 120
45 TGAGATAAGG GCCTCCTGGA ACAACCTCAG AATGAACCGG GTACATTGCC AAATAATAGG 180
ACATGACCCC TTAGTTACGT AGAATTCCCT TGGCAGAACC CCTTGTCGCT TGGCAGAACC 240
50 CCTTAGTTAT GTAAACTTGT ACTTTCCTG CCCGCTCTC CCCCTTGAG TTTTACTAT 300
ATAAGC 306

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 305 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CCTCCCATCT AGAGAGTGTT CCCAAAACAC TCCTGAACTC TTCACCCCAG AATGCATTCC 60
 5 TGAAGTCCTC ACCCTAAAGT TCAAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
 GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCGGG TACATTGCCA AATAATAGGA 180
 CATGACCCCT TAGTTACACA GAATTCCTT GGCAAAACCC CTTGTCCCTT GGCAGAACCC 240
 10 CTTAGTTATG CAACTTGTA CTTTCCCTGC CCAGCTCTCC CCCCTTGAGT TTTTCCTATA 300
 TAAGC 305

15 (2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 304 base pairs
 (B) TYPE: nucleic acid
 20 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

30 CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTGAACTC TTCACCCCAG AATGCATTCC 60
 TGAAGTCCTC ACCCTAGAGT TTGAACCCCTC CCAACTAAAG ACTGTTCCAA GAACATCTTT 120
 GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAACCGGG TACATTGCCA AATAATAGGA 180
 35 CATGACCCCT TAGTTACGTA GAATTCCTT GGCAGAACCC CTTGTCGCTT GGCAGAACCC 240
 CTTAGTTATG CAACTTGTA CTTTCCCTGC CCCGCTCTCC CCCTTGAGTT TTTCTATAT 300
 40 AAGC 304

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

45 (A) LENGTH: 303 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: DNA (genomic)

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CCTCCCATCT AGAGAGTGTT CCCAGAACAC TCCTAAACTC TTCACCCCAG AATGCATTCC 60
 TGAAGTCCTC ACCCTAGAGT TCGAACCTT CCAACTAAAG ACTGTTCCAA GAACATTTTT 120
 60 GAGATAAGGG CCTCCTGGAA CAACCTCAA ATGAACCGGG TACATTGCCA AATGATAGGA 180
 CATGACCCCT TAGTTACGTA GATTCCCTT GCAGAACCCC TTGTCCCTTG GCAGAACCCC 240
 CTAGTGATGT AACTTGTAC TTTCCCTGCC CAGCTCTCCC CCCTTGAGTT TTCCTATATA 300
 65 AGC 303

(2) INFORMATION FOR SEQ ID NO:14:

5 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8657 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

	TGAAGAATAA AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC	60
	CCCTCCCATC TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT	120
20	CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG	180
	TTGTAAACT TCCCCTATTC CCTCCCCATT CCCCTCCCA GTTTGTGGTT TTTTCCTTTA	240
25	AAAGCTTGTG AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG	300
	AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTGAC CCCAGAGCTC	360
	TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TTAAATCTTA CCTTCTACAT TTTATGTATG	420
30	GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT	480
	TCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG	540
35	CATTGGCCGG GAATTCGAAA ATCTTTCATT TGGTGCATTG GCCGGGAAAC AGCGCGACCA	600
	CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTTGG TCTGATGTCT	660
	GTGTTCTGAT GTCTGTGTTC TGTTTCTAAG TCTGGTGC GA TCGCAGTTTC AGTTTTGCGG	720
40	ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
	GGTGTCCACC GTCCGTTCGC CCTGGGAGAC GTCCCAGGAG GAACAGGGGA GGATCAGGGA	840
45	CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCAGAT CGTGGGTTCG	900
	AGTCCCACCT CGTGCCAGT TGCGAGATCG TGGGTCGAG TCCCACCTCG TGTTTTGTG	960
	CGAGATCGTG GGTTGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
50	ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
	CGTGGGTTCG AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
55	CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTGCG	1260
	TTGGTAATTT TGTTTGTTTA CGTTTGTTTT TGTGAGTCGT CTATGTTGTC TGTTACTATC	1320
60	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTGAGA	1380
	CTTGACTGA TGA CTGACGA CTGTTTTTAA GTTATGCCTT CTAAAATAAG CCTAAAAATC	1440
65	CTGTCAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACGTATCGAT	1500

	GGATCCCTCG	ACTAACTAAT	AGCCCATTCT	CCAAGGTCGA	GCGGGATCAA	TTCCGCCCCC	1560
	CCCCTAACGT	TACTGGCCGA	AGCCGCTTGG	AATAAGGCCG	GTGTGCGTTT	GTCTATATGT	1620
5	TATTTTCCAC	CATATTGCCG	TCTTTTGCCA	ATGTGAGGGC	CCGGAACCT	GGCCCTGTCT	1680
	TCTTGACGAG	CATTCCTAGG	GGTCTTTCCC	CTCTCGCCAA	AGGAATGCAA	GGTCTGTTGA	1740
	ATGTCGTGAA	GGAAGCAGTT	CCTCTGGAAG	CTTCTTGAAG	ACAAACAACG	TCTGTAGCGA	1800
10	CCCTTTGCAG	GCAGCGGAAC	CCCCACCTG	GCGACAGGTG	CCTCTGCGGC	CAAAAGCCAC	1860
	GTGTATAAGA	TACACCTGCA	AAGGCGGCAC	AACCCAGTG	CCACGTTGTG	AGTTGGATAG	1920
15	TTGTGGAAAG	AGTCAAATGG	CTCTCCTCAA	GCGTATTCAA	CAAGGGGCTG	AAGGATGCCC	1980
	AGAAGGTACC	CCATTGTATG	GGATCTGATC	TGGGGCCTCG	GTGCACATGC	TTTACATGTG	2040
	TTTAGTCGAG	GTTAAAAAAA	CGTCTAGGCC	CCCCGAACCA	CGGGGACGTG	GTTTTCTTTT	2100
20	GAAAAACACG	ATAATAATCA	TGGGCGCGGA	TCCCGTCGTT	TTACAACGTC	GTGACTGGGA	2160
	AAACCCTGGC	GTTACCCAAC	TTAATCGCCT	TGCAGCACAT	CCCCCTTTCG	CCAGCTGGCG	2220
25	TAATAGCGAA	GAGGCCCGCA	CCGATCGCCC	TTCCCAACAG	TTGCGCAGCC	TGAATGGCGA	2280
	ATGGCGCTTT	GCCTGGTTTC	CGGCACCAGA	AGCGGTGCCG	GAAAGCTGGC	TGGAGTGCGA	2340
	TCTTCTGAG	GCCGATACTG	TCGTCGTCCC	CTCAAACCTG	CAGATGCACG	GTTACGATGC	2400
30	GCCCATCTAC	ACCAACGTAA	CCTATCCCAT	TACGGTCAAT	CCGCCGTTTG	TTCCACGGA	2460
	GAATCCGACG	GGTTGTTACT	CGCTCACATT	TAATGTTGAT	GAAAGCTGGC	TACAGGAAGG	2520
35	CCAGACGCGA	ATTATTTTTG	ATGGCGTTAA	CTCGGCGTTT	CATCTGTGGT	GCAACGGGCG	2580
	CTGGGTCGGT	TACGGCCAGG	ACAGTCGTTT	GCCGTCTGAA	TTTGACCTGA	GCGCATTTTT	2640
	ACGCGCCGGA	GAAAACCGCC	TCGCGGTGAT	GGTGCTGCGT	TGGAGTGACG	GCAGTTATCT	2700
40	GGAAGATCAG	GATATGTGGC	GGATGAGCGG	CATTTTCCGT	GACGTCTCGT	TGTCGCATAA	2760
	ACCGACTACA	CAAATCAGCG	ATTTCCATGT	TGCCACTCGC	TTTAATGATG	ATTTACGCCG	2820
45	CGCTGTACTG	GAGGCTGAAG	TTCAGATGTG	CGGCGAGTTG	CGTGACTION	TACGGGTAAC	2880
	AGTTTCTTTA	TGGCAGGGTG	AAACGCAGGT	CGCCAGCGGC	ACCGCGCCTT	TCGGCGGTGA	2940
	AATTATCGAT	GAGCGTGGTG	GTTATGCCGA	TCGCGTCACA	CTACGTCTGA	ACGTCGAAAA	3000
50	CCCGAAACTG	TGGAGCGCCG	AAATCCCGAA	TCTCTATCGT	GCGGTGGTTG	AACTGCACAC	3060
	CGCCGACGGC	ACGCTGATTG	AAGCAGAAGC	CTGCGATGTC	GGTTTCCGCG	AGGTGCGGAT	3120
55	TGAAAATGGT	CTGCTGCTGC	TGAACGGCAA	GCCGTTGCTG	ATTTCGAGCG	TTAACCGTCA	3180
	CGAGCATCAT	CCTCTGCATG	GTCAGGTCAT	GGATGAGCAG	ACGATGGTGC	AGGATATCCT	3240
	GCTGATGAAG	CAGAACAAC	TTAACGCCGT	GCGCTGTTTC	CATTATCCGA	ACCATCCGCT	3300
60	GTGGTACACG	CTGTGCGACC	GCTACGGCCT	GTATGTGGTG	GATGAAGCCA	ATATTGAAAC	3360
	CCACGGCATG	GTGCCAATGA	ATCGTCTGAC	CGATGATCCG	CGCTGGCTAC	CGGCGATGAG	3420
	CGAACGCGTA	ACGCGAATGG	TGCAGCGCGA	TCGTAATCAC	CCGAGTGTGA	TCATCTGGTC	3480
65	GCTGGGGAAT	GAATCAGGCC	ACGGCGCTAA	TCACGACGCG	CTGTATCGCT	GGATCAAATC	3540

	TGTCGATCCT	TCCCGCCCGG	TGCAGTATGA	AGGCGGCGGA	GCCGACACCA	CGGCCACCGA	3600
	TATTATTTCG	CCGATGTACG	CGCGCGTGGA	TGAAGACCAG	CCCTTCCCGG	CTGTGCCGAA	3660
5	ATGGTCCATC	AAAAAATGGC	TTTCGCTACC	TGGAGAGACG	CGCCCGCTGA	TCCTTTGCGA	3720
	ATACGCCAC	GCGATGGGTA	ACAGTCTTGG	CGGTTTCGCT	AAATACTGGC	AGGCGTTTCG	3780
10	TCAGTATCCC	CGTTTACAGG	GCGGCTTCGT	CTGGGACTGG	GTGGATCAGT	CGCTGATTAA	3840
	ATATGATGAA	AACGGCAACC	CGTGGTCGGC	TTACGGCGGT	GATTTTGGCG	ATACGCCGAA	3900
	CGATCGCCAG	TTCTGTATGA	ACGGTCTGGT	CTTTGCCGAC	CGCACGCCGC	ATCCAGCGCT	3960
15	GACGGAAGCA	AAACACCAGC	AGCAGTTTTT	CCAGTTCCTG	TTATCCGGGC	AAACCATCGA	4020
	AGTGACCAGC	GAATACCTGT	TCCGTCATAG	CGATAACGAG	CTCCTGCACT	GGATGGTGGC	4080
20	GCTGGATGGT	AAGCCGCTGG	CAAGCGGTGA	AGTGCCTCTG	GATGTCGCTC	CACAAGGTAA	4140
	ACAGTTGATT	GAAGTGCCTG	AACTACCGCA	GCCGGAGAGC	GCCGGGCAAC	TCTGGCTCAC	4200
	AGTACGCGTA	GTGCAACCGA	ACGCGACCGC	ATGGTCAGAA	GCCGGGCACA	TCAGCGCCTG	4260
25	GCAGCAGTGG	CGTCTGGCGG	AAAACCTCAG	TGTGACGCTC	CCCGCCGCGT	CCCACGCCAT	4320
	CCCGCATCTG	ACCACCAGCG	AAATGGATTT	TTGCATCGAG	CTGGGTAATA	AGCGTTGGCA	4380
30	ATTTAACCGC	CAGTCAGGCT	TTCTTTCACA	GATGTGGATT	GGCGATAAAA	AACAACCTGCT	4440
	GACGCCGCTG	CGCGATCAGT	TCACCCGTGC	ACCGCTGGAT	AACGACATTG	GCGTAAGTGA	4500
	AGCGACCCGC	ATTGACCCTA	ACGCCTGGGT	CGAACGCTGG	AAGGCGGCGG	GCCATTACCA	4560
35	GGCCGAAGCA	GCGTTGTTGC	AGTGCACGGC	AGATACACTT	GCTGATGCGG	TGCTGATTAC	4620
	GACCGCTCAC	GCGTGGCAGC	ATCAGGGGAA	AACCTTATTT	ATCAGCCGGA	AAACCTACCG	4680
40	GATTGATGGT	AGTGGTCAAA	TGGCGATTAC	CGTTGATGTT	GAAGTGGCGA	GCGATACACC	4740
	GCATCCGGCG	CGGATTGGCC	TGAACTGCCA	GCTGGCGCAG	GTAGCAGAGC	GGGTAAACTG	4800
	GCTCGGATTA	GGGCCGCAAG	AAAACCTATCC	CGACCGCCTT	ACTGCCGCCT	GTTTTGACCG	4860
45	CTGGGATCTG	CCATTGTCAG	ACATGTATAC	CCCGTACGTC	TTCCCGAGCG	AAAACGGTCT	4920
	GCGCTGCGGG	ACGCGCGAAT	TGAATTATGG	CCCACACCAG	TGGCGCGGCG	ACTTCCAGTT	4980
50	CAACATCAGC	CGCTACAGTC	AACAGCAACT	GATGGAACC	AGCCATCGCC	ATCTGCTGCA	5040
	CGCGGAAGAA	GGCACATGGC	TGAATATCGA	CGGTTTCCAT	ATGGGGATTG	GTGGCGACGA	5100
	CTCCTGGAGC	CCGTCAGTAT	CGGCGGAATT	CCAGCTGAGC	GCCGGTCGCT	ACCATTACCA	5160
55	GTTGGTCTGG	TGTCAAAAAT	AATAATAACC	GGGCAGGGGG	GATCCGAAGG	CGGGGACAGC	5220
	AGTGCAGTGG	TGGACAGAAA	GCAAGTGATC	TAGGCCAGCA	GCCTCCCTAA	AGGGACTTCA	5280
	GCCCACAAAG	CCAACTTGT	GGCTTTAATA	CAAGCTCTGT	AAATGGTAAA	AAAAAAAAAAG	5340
60	TCTACACGGA	CAGCAGGTAT	GCTCTTGCCA	CTGTACAGAG	CAATATACAG	ACAAAGAGAA	5400
	CTGTTGACAT	CTGCAGAGAA	AGACCTAAGA	TGCTGTGGCT	AAAAGAAATC	AGATGGCAAA	5460
65	TCTAACCGCC	CAGGCATCCT	AAAGAGCAAT	GATCCTGACA	GTCTGAAGAC	TATCAAGTTA	5520

	TAGACAAATT AAGACTGGTA AAAAAAACCC TGTATAAAAT AGTAAAAACT GAAAAAGAA	5580
	AAC TAGTCCT CTCATGAGAA GACAGACCTG ACATCTACTG AAAAATAGAC TTTACTGGAA	5640
5	AAAATATGTG TATGAATACC TTCTAGTTTT TGTGAACGTT CTCAAGATGG ATAAAAGCTT	5700
	TTCC TTGTAA AACGAGACTG ATCAGATAGT CATCAAGAAG ATTGTTAAAG AAAATTTTCC	5760
10	AAGGTT CGGA GTGCCAAAAG CAATAGTGTC AGATAATGGT CCTGCCTTTG TTGCCCAGGT	5820
	AAGTCAGGGT GTGGCCAAGT ATTTAGAGGT CAAATGAAAA TTCCATTGTG TGTACAGACC	5880
	TCAGAGCTCA GGAAAGATAA AAAAGAATAA ATAAAAC TCT AAACAGACCT TGACAAAATT	5940
15	AATCCTAGAG ACTGGCACAG ACTTACTTGG TACTCCTTCC CCTTGCCCTA TTTAGAACTG	6000
	AGAATACTCC CTCTTGATTG GGT TTTACTC TTTTAAAGAT CCTTTATGGG GCTCCTATGC	6060
20	CATCACTGTC TTAAATGATG TGTTTAAACC TATGTTGTTA TAATAATGAT CTATATGTTA	6120
	AGTTAAAAGG CTTGCAGGTG GTGCAGAAAG AAGTCTGGTC ACAACTGGCT ACAGTGAACA	6180
	AGCTGGGTAC CCAAGGACA TCTTACCAGT TCCAGCCAGA GATCTGATCT ACGATCCCCG	6240
25	GGTCGACCCG GGTGACCCCT GTGGAATGTG TGTCAAGTAG GGTGTGGAAG GTCCCCAGGC	6300
	TCCCCAGCAG GCAGAAGTAT GCAAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA	6360
30	AAGTCCCCAG GCTCCCCAGC AGGCAGAAGT ATGCAAAGCA TGCATCTCAA TTAGTCAGCA	6420
	ACCATAGTCC CGCCCCAAC TCCGCCCATC CCGCCCC TAA CTCCGCCAG TTCCGCCAT	6480
	TCTCCGCCCC ATGGCTGACT AATTTTTTTT ATTTATGCAG AGGCCGAGGC CGCCTCGGCC	6540
35	TCTGAGCTAT TCCAGAAGTA GTGAGGAGGC TTTT TGGAG GCCTAGGCTT TTGCAAAAAG	6600
	CTTCACGCTG CCGCAAGCAC TCAGGGCGCA AGGGCTGCTA AAGGAAGCGG AACACGTAGA	6660
40	AAGCCAGTCC GCAGAAACGG TGCTGACCCC GGATGAATGT CAGCTACTGG GCTATCTGGA	6720
	CAAGGGAAAA CGCAAGCGCA AAGAGAAAGC AGGTAGCTTG CAGTGGGCTT ACATGGCGAT	6780
	AGCTAGACTG GGCGGTTTTA TGGACAGCAA GCGAACC GGA ATTGCCAGCT GGGGCGCCCT	6840
45	CTGGTAAGGT TGGGAAGCCC TGCAAAGTAA ACTGGATGGC TTTCTTGCCG CCAAGGATCT	6900
	GATGGCGCAG GGGATCAAGA TCTGATCAAG AGACAGGATG AGGATCGTTT CGCATGATTG	6960
50	AACAAGATGG ATTGCACGCA GGTTCTCCGG CCGCTTGGGT GGAGAGGCTA TTCGGCTATG	7020
	ACTGGGCACA ACAGACAATC GGCTGCTCTG ATGCCGCCGT GTTCCGGCTG TCAGCGCAGG	7080
55	GGCGCCCGGT TCTTTTGTG AAGACCGACC TGTCCGGTGC CCTGAATGAA CTGCAGGACG	7140
	AGGCAGCGCG GCTATCGTGG CTGGCCACGA CGGGCGTTCC TTGCGCAGCT GTGCTCGACG	7200
	TTGTCACTGA AGCGGGAAGG GACTGGCTGC TATTGGGCGA AGTGCCGGGG CAGGATCTCC	7260
	TGTCATCTCA CCTTGCTCCT GCCGAGAAAG TATCCATCAT GGCTGATGCA ATGCGGCGGC	7320
60	TGCATACGCT TGATCCGGCT ACCTGCCCAT TCGACCACCA AGCGAAACAT CGCATCGAGC	7380
	GAGCACGTAC TCGGATGGAA GCCGGTCTTG TCGATCAGGA TGATCTGGAC GAAGAGCATC	7440
65	AGGGGCTCGC GCCAGCCGAA CTGTTGCGCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG	7500
	ATCTCGTCGT GACCCATGGC GATGCCTGCT TGCCGAATAT CATGGTGGAA AATGGCCGCT	7560

5 TTTCTGGATT CATCGACTGT GGCCGGCTGG GTGTGGCGGA CCGCTATCAG GACATAGCGT 7620
 TGGCTACCCG TGATATTGCT GAAGAGCTTG GCGGCGAATG GGCTGACCGC TTCCTCGTGC 7680
 TTTACGGTAT CGCCGCTCCC GATTCGCAGC GCATCGCCTT CTATCGCCTT CTTGACGAGT 7740
 TCTTCTGAGC GGGACTCTGG GGTTCGAAAT GACCGACCAA GCGACGCCCA ACCTGCCATC 7800
 10 ACGAGATTTC GATTCCACCG CCGCCTTCTA TGAAAGGTTG GGCTTCGGAA TCGTTTTCCG 7860
 GGACGGAATT CGTAATCTGC TGCTTGCAA CAAAAAACC ACCGCTACCA GCGGTGGTTT 7920
 GTTTGCCGGA TCAAGAGCTA CCAACTCTTT TTCCGAAGGT AACTGGCTTC AGCAGAGCGC 7980
 15 AGATACCAA TACTGTCCTT CTAGTGTAGC CGTAGTTAGG CCACCACTTC AAGAACTCTG 8040
 TAGCACCGCC TACATACCTC GCTCTGCTAA TCCTGTTACC AGTGGCTGCT GCCAGTGGCG 8100
 20 ATAAGTCGTG TCTTACCGGG TTGGACTCAA GACGATAGTT ACCGGATAAG GCGCAGCGGT 8160
 CGGGCTGAAC GGGGGGTTG TGCACACAGC CCAGCTTGGA GCGAACGACC TACACCGAAC 8220
 TGAGATACCT ACAGCGTGAG CATTGAGAAA GCGCCACGCT TCCCGAAGGG AGAAAGGCGG 8280
 25 ACAGGTATCC GGTAAGCGGC AGGGTCGGAA CAGGAGAGCG CACGAGGGAG CTTCCAGGGG 8340
 GAAACGCCTG GTATCTTTAT AGTCCTGTCG GGTTCGCCA CCTCTGACTT GAGCGTCGAT 8400
 30 TTTTGTGATG CTCGTCAGGG GGGCGGAGCC TATGGAAAA CGCCAGCAAC GCCGAGATGC 8460
 GCCGCCTCGA GTACACCTGC GTCATGCTGA GACCTCAAG CCTCACTAAA AGGGTCCCTG 8520
 CCTAGTTCTG TTTACTAATC TGCCTTATTC TGTTTTTGTT CCCATGTTAA AGATAGAGTA 8580
 35 AATGCAGTAT TCTCCACATA GAGATATAGA CTTCTGAAAT TCTAAGATTA GAATTATTTA 8640
 CAAGAAGAAG TGGGGAA 8657

40 (2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6359 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

55 TGAAGAATAA AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC 60
 CCCTCCCATC TGGA AACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT 120
 CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG 180
 60 TTGTTAAACT TCCCCTATTC CCTCCCCATT CCCCCTCCCA GTTTGTGGTT TTTTCCTTTA 240
 AAAGCTTGTG AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG 300
 65 AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTGAC CCCAGAGCTC 360

	TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TTAAATCTTA CCTTCTACAT TTTATGTATG	420
	GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT	480
5	TCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG	540
	CATTGGCCGG GAATTCGAAA ATCTTTCATT TGGTGCATTG GCCGGGAAAC AGCGCGACCA	600
10	CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTTGG TCTGATGTCT	660
	GTGTTCGAT GTCTGTGTTC TGTTTCTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG	720
	ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
15	GGTGTCCACC GTCCGTTTCG CCTGGGAGAC GTCCAGGAG GAACAGGGGA GGATCAGGGA	840
	CGCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCAGAT CGTGGGTTCG	900
	AGTCCCACCT CGTGCCAGT TGCAGATCG TGGGTTCGAG TCCCACCTCG TGTTTGTG	960
20	CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
	ACCTCGTGTT TTGTTGCGAG ATCGTGGGTG CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
25	CGTGGGTTCG AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
	CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
30	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTGCG	1260
	TTGGTAATTT TGTTTGTTTA CGTTTGTTTT TGTGAGTCGT CTATGTTGTC TGTACTATC	1320
	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTGAGA	1380
35	CTTGGACTGA TGA CTGACGA CTGTTTTTAA GTTATGCCTT CTAATAAAG CCTAAAAATC	1440
	CTGTGAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACTCGAGCTC	1500
40	AAGCTTCGAA TTCTGCAGTC GACGGTACCG CGGCCGCTAA CTAATAGCCC ATTCTCCAAG	1560
	GTACGTAGCG GGGATCAATT CCGCCCCCCC CCTAACGTTA CTGGCCGAAG CCGCTTGGAA	1620
	TAAGGCCGGT GTGCGTTTGT CTATATGTTA TTTTCCACCA TATTGCCGTC TTTTGGCAAT	1680
45	GTGAGGGCCC GGAAACCTGG CCCTGTCTTC TTGACGAGCA TTCCTAGGGG TCTTTCCCCT	1740
	CTCGCCAAAG GAATGCAAGG TCTGTTGAAT GTCGTGAAGG AAGCAGTTCC TCTGGAAGCT	1800
50	TCTTGAAGAC AAACAACGTC TG TAGCGACC CTTTGCAGGC AGCGGAACCC CCCACCTGGC	1860
	GACAGGTGCC TCTGCGGCCA AAAGCCACGT GTATAAGATA CACCTGCAAA GGCGGCACAA	1920
	CCCCAGTGCC ACGTTGTGAG TTGGATAGTT GTGGAAAGAG TCAAATGGCT CTCCTCAAGC	1980
55	GTATTCAACA AGGGGCTGAA GGATGCCCAG AAGGTACCCC ATTGTATGGG ATCTGATCTG	2040
	GGGCCTCGGT GCACATGCTT TACATGTGTT TAGTCGAGGT TAAAAAACG TCTAGGCCCC	2100
	CCGAACCACG GGGACGTGGT TTTCCTTTGA AAAACACGAT ACGGGATCCA CCGGTCGCCA	2160
60	CCATGGGTAA AGGAGAAGAA CTTTTCACAG GAGTTGTCCC AATTCTTGTT GAATTAGATG	2220
	GTGATGTTAA TGGGCACAAA TTTTCTGTCA GTGGAGAGGG TGAAGGTGAT GCAACATACG	2280
65	GAAACTTAC CCTTAAATTT ATTTGCACTA CTGGAAACT ACCTGTTCCA TGGCCAACAC	2340
	TTGTCACTAC TTTCACCTAT GGTGTTCAAT GCTTTTCAAG ATACCCAGAT CATATGAAAC	2400

	GGCATGACTT TTTCAAGAGT GCCATGCCCC AAGGTTATGT ACAGGAAAGA ACTATATTTT	2460
	TCAAAGATGA CGGGAAGTAC AAGACACGTG CTGAAGTCAA GTTTGAAGGT GATACCCTTG	2520
5	TTAATAGAAT CGAGTTAAAA GGTATTGATT TTAAAGAAGA TGGAAACATT CTTGGACACA	2580
	AATTGGAATA CAACTATAAC TCACACAATG TATACATCAT GGCAGACAAA CAAAAGAATG	2640
10	GAACCAAAGT TAACTTCAAA ATTAGACACA ACATTGAAGA TGAAGCGTT CAACTAGCAG	2700
	ACCATTATCA ACAAATACT CCAATTGGCG ATGGCCCTGT CCTTTTACCA GACAACCATT	2760
	ACCTGTCCAC ACAATCTGCC CTTTCGAAAG ATCCCAACGA AAAGAGAGAC CACATGGTCC	2820
15	TTCTTGAGTT TGTAACAGCT GCTGGGATTA CACATGGCAT GGATGAACTA TACAAGTCCG	2880
	GATCTAGATA ACTGTATCGA TGGATCCGAA GCGGGGACA GCAGTGCAGT GGTGGACAGA	2940
20	AAGCAAGTGA TCTAGGCCAG CAGCCTCCCT AAAGGGACTT CAGCCCACAA AGCCAAACTT	3000
	GTGGCTTTAA TACAAGCTCT GTAAATGGTA AAAAAAAAAA AGTCTACACG GACAGCAGGT	3060
	ATGCTCTTGC CACTGTACAG AGCAATATAC AGACAAAGAG AACTGTTGAC ATCTGCAGAG	3120
25	AAAGACCTAA GATGCTGTGG CTAAAAGAAA TCAGATGGCA AATCTAACCG CCCAGGCATC	3180
	CTAAAGAGCA ATGATCCTGA CAGTCTGAAG ACTATCAAGT TATAGACAAA TTAAGACTGG	3240
30	TAAAAAAAC CCTGTATAAA ATAGTAAAAA CTGAAAAAAG AAAACTAGTC CTCTCATGAG	3300
	AAGACAGACC TGACATCTAC TGAAAAATAG ACTTTACTGG AAAAAATATG TGTATGAATA	3360
	CCTTCTAGTT TTTGTGAACG TTCTCAAGAT GGATAAAGC TTTTCCTTGT AAAACGAGAC	3420
35	TGATCAGATA GTCATCAAGA AGATTGTAA AGAAAATTTT CCAAGGTTCTG GAGTGCCAAA	3480
	AGCAATAGTG TCAGATAATG GTCCTGCCTT TGTGCCCAG GTAAGTCAGG GTGTGGCCAA	3540
40	GTATTTAGAG GTCAAATGAA AATTCCATTG TGTGTACAGA CCTCAGAGCT CAGGAAAGAT	3600
	AAAAAGAAT AAATAAACT CTAAACAGAC CTTGACAAAA TTAATCCTAG AGACTGGCAC	3660
	AGACTTACTT GGTA CTCTT CCCCTTGCCC TATTTAGAAC TGAGAATACT CCCTCTTGAT	3720
45	TCGGTTTTAC TCTTTTAAAG ATCCTTTATG GGGCTCCTAT GCCATCACTG TCTTAAATGA	3780
	TGTGTTTAAA CCTATGTTGT TATAATAATG ATCTATATGT TAAGTTAAAA GGCTTGCAAG	3840
50	TGGTGCAGAA AGAAGTCTGG TCACAACTGG CTACAGTGAA CAAGCTGGGT ACCCCAAGGA	3900
	CATCTTACCA GTTCCAGCCA GAGATCTGAT CTACGATCCC CGGGTCGACC CGGGTCGACC	3960
	CTGTGGAATG TGTGTCAGTT AGGGTGTGGA AAGTCCCCAG GCTCCCCAGC AGGCAGAAGT	4020
55	ATGCAAAGCA TGCACTCTCA TTAGTCAGCA ACCAGGTGTG GAAAGTCCCC AGGCTCCCCA	4080
	GCAGGCAGAA GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCATAGT CCCGCCCTA	4140
	ACTCCGCCCA TCCCGCCCT AACTCCGCC AGTTCCGCC ATTCTCCGCC CCATGGCTGA	4200
60	CTAATTTTTT TTATTTATGC AGAGGCCGAG GCCGCTCGG CCTCTGAGCT ATTCCAGAAG	4260
	TAGTGAGGAG GCTTTTTTGG AGGCCTAGGC TTTTGCAAAA AGCTTCACGC TGCCGCAAGC	4320
65	ACTCAGGGCG CAAGGGCTGC TAAAGGAAGC GGAACACGTA GAAAGCCAGT CCGCAGAAAC	4380

	GGTGCTGACC CCGGATGAAT GTCAGCTACT GGGCTATCTG GACAAGGGAA AACGCAAGCG	4440
	CAAAGAGAAA GCAGGTAGCT TGCAGTGGGC TTACATGGCG ATAGCTAGAC TGGGCGGTTT	4500
5	TATGGACAGC AAGCGAACCG GAATTGCCAG CTGGGGCGCC CTCTGGTAAG GTTGGGAAGC	4560
	CCTGCAAAGT AACTGGATG GCTTTCTTGC CGCCAAGGAT CTGATGGCGC AGGGGATCAA	4620
10	GATCTGATCA AGAGACAGGA TGAGGATCGT TTCGCATGAT TGAACAAGAT GGATTGCACG	4680
	CAGGTTCTCC GGCCGCTTGG GTGGAGAGGC TATTCGGCTA TGAAGGGCA CAACAGACAA	4740
	TCGGCTGCTC TGATGCCGCC GTGTTCCGGC TGTCAGCGCA GGGGCGCCCG GTTCTTTTTG	4800
15	TCAAGACCGA CCTGTCCGGT GCCCTGAATG AACTGCAGGA CGAGGCAGCG CGGCTATCGT	4860
	GGCTGGCCAC GACGGGCGTT CCTTGCGCAG CTGTGCTCGA CGTTGTCACT GAAGCGGGAA	4920
	GGGACTGGCT GCTATTGGGC GAAGTGCCGG GGCAGGATCT CCTGTCATCT CACCTTGCTC	4980
20	CTGCCGAGAA AGTATCCATC ATGGCTGATG CAATGCGGCG GCTGCATACG CTTGATCCGG	5040
	CTACCTGCCC ATTCGACCAC CAAGCGAAAC ATCGCATCGA GCGAGCACGT ACTCGGATGG	5100
25	AAGCCGGTCT TGTCGATCAG GATGATCTGG ACGAAGAGCA TCAGGGGCTC GCGCCAGCCG	5160
	AACTGTTGCG CAGGCTCAAG GCGCGCATGC CCGACGGCGA GGATCTCGTC GTGACCCATG	5220
	GCGATGCCTG CTTGCCGAAT ATCATGGTGG AAAATGGCCG CTTTTCTGGA TTCATCGACT	5280
30	GTGGCCGGCT GGGTGTGGCG GACCGCTATC AGGACATAGC GTTGGCTACC CGTGATATTG	5340
	CTGAAGAGCT TGGCGGCGAA TGGGCTGACC GCTTCTCGT GCTTTACGGT ATCGCCGCTC	5400
35	CCGATTCGCA GCGCATCGCC TTCTATCGCC TTCTTGACGA GTTCTTCTGA GCGGGACTCT	5460
	GGGGTTTCGAA ATGACCGACC AAGCGACGCC CAACCTGCCA TCACGAGATT TCGATTCCAC	5520
	CGCCGCCTTC TATGAAAGGT TGGGCTTCGG AATCGTTTTT CGGGACGGAA TTCGTAATCT	5580
40	GCTGCTTGCA AACAAAAA CCACCGCTAC CAGCGGTGGT TTGTTTGCCG GATCAAGAGC	5640
	TACCAACTCT TTTTCCGAAG GTAAGTGGCT TCAGCAGAGC GCAGATACCA AATACTGTCC	5700
45	TTCTAGTGTA GCCGTAGTTA GGCCACCACT TCAAGAACTC TGTAGCACCG CCTACATACC	5760
	TCGCTCTGCT AATCCTGTTA CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG	5820
	GGTTGGACTC AAGACGATAG TTACCGGATA AGGCGCAGCG GTCGGGCTGA ACGGGGGGTT	5880
50	CGTGACACA GCCCAGCTTG GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG	5940
	AGCATTGAGA AAGCGCCACG CTTCCCGAAG GGAGAAAGGC GGACAGGTAT CCGGTAAGCG	6000
	GCAGGGTCGG AACAGGAGAG CGCACGAGGG AGCTTCCAGG GGGAAACGCC TGGTATCTTT	6060
55	ATAGTCCTGT CGGGTTTCGC CACCTCTGAC TTGAGCGTCG ATTTTGTGA TGCTCGTCAG	6120
	GGGGGCGGAG CCTATGGAAA AACGCCAGCA ACGCCGAGAT GCGCCGCCTC GAGTACACCT	6180
60	GCGTCATGCT GAGACCCTCA AGCCTCACTA AAAGGGTCCC TGCCTAGTTC TGTTTACTAA	6240
	TCTGCCTTAT TCTGTTTTTG TTCCCATGTT AAAGATAGAG TAAATGCAGT ATTCTCCACA	6300
65	TAGAGATATA GACTTCTGAA ATTCTAAGAT TAGAATTATT TACAAGAAGA AGTGGGGAA	6359

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 6891 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

15	TGAAGAATAA AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC	60
	CCCTCCCATC TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT	120
	CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG	180
20	TTGTTAAACT TCCCCTATTC CCTCCCCATT CCCCCTCCA GTTTGTGGTT TTTTCCTTTA	240
	AAAGCTTG TG AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG	300
25	AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTCGAC CCCAGAGCTC	360
	TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TTAAATCTTA CCTTCTACAT TTTATGTATG	420
	GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT	480
30	TCCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTGGTG	540
	CATTGGCCGG GAATTCGAAA ATCTTTCATT TGGTGCATTG GCCGGGAAAC AGCGCGACCA	600
35	CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTTGG TCTGATGTCT	660
	GTGTCTGAT GTCTGTGTTC TGTTTCTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG	720
	ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
40	GGTGTCACC GTCCGTTTCG CCTGGGAGAC GTCCAGGAG GAACAGGGGA GGATCAGGGA	840
	CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCGAGAT CGTGGGTTCG	900
45	AGTCCCACCT CGTGCCAGT TCGGAGATCG TGGGTTTCGAG TCCCACCTCG TGTTTTGTTG	960
	CGAGATCGTG GGTTGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
	ACCTCGTGTT TTGTTGCGAG ATCGTGGGT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
50	CGTGGGTTCG AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
	CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
55	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTGCG	1260
	TTGGTAATTT TGTTTGTTTA CGTTTGTTTT TGTGAGTCGT CTATGTTGTC TGTTACTATC	1320
	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTTACA	1380
60	CTTGACTGA TGA CTGACGA CTGTTTTTAA GTTATGCCTT CTAAAATAAG CCTAAAATC	1440
	CTGTCAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACTCGAGCTC	1500
65	AAGCTTCGAA TTCTGCAGTC GACGGTACCG CGGGGATCAA TTCCGCCCCC CCCCTAACGT	1560

	TACTGGCCGA AGCCGCTTGG AATAAGGCCG GTGTGCGTTT GTCTATATGT TATTTTCCAC	1620
	CATATTGCCG TCTTTTGGCA ATGTGAGGGC CCGGAAACCT GGCCCTGTCT TCTTGACGAG	1680
5	CATTCTAGG GGTCTTTCCC CTCTCGCCAA AGGAATGCAA GGTCTGTTGA ATGTCGTGAA	1740
	GGAAGCAGTT CCTCTGGAAG CTTCTTGAAG ACAAACAACG TCTGTAGCGA CCCTTTGCAG	1800
10	GCAGCGGAAC CCCCCACCTG GCGACAGGTG CCTCTGCGGC CAAAAGCCAC GTGTATAAGA	1860
	TACACCTGCA AAGGCGGCAC AACCCCAGTG CCACGTTGTG AGTTGGATAG TTGTGAAAG	1920
	AGTCAAATGG CTCTCCTCAA GCGTATTCAA CAAGGGGCTG AAGGATGCCC AGAAGGTACC	1980
15	CCATTGTATG GGATCTGATC TGGGGCCTCG GTGCACATGC TTTACATGTG TTTAGTCGAG	2040
	GTTAAAAAAC GTCTAGGCCC CCCGAACCAC GGGGACGTGG TTTTCCTTTG AAAAACACGA	2100
20	GCGGGATCAA TTCCGCCCCC CCCCTAACGT TACTGGCCGA AGCCGCTTGG AATAAGGCCG	2160
	GTGTGCGTTT GTCTATATGT TATTTTCCAC CATATTGCCG TCTTTTGGCA ATGTGAGGGC	2220
	CCGGAAACCT GGCCCTGTCT TCTTGACGAG CATTCTAGG GGTCTTTCCC CTCTCGCCAA	2280
25	AGGAATGCAA GGTCTGTTGA ATGTCGTGAA GGAAGCAGTT CCTCTGGAAG CTTCTTGAAG	2340
	ACAAACAACG TCTGTAGCGA CCCTTTGCAG GCAGCGGAAC CCCCCACCTG GCGACAGGTG	2400
30	CCTCTGCGGC CAAAAGCCAC GTGTATAAGA TACACCTGCA AAGGCGGCAC AACCCCAGTG	2460
	CCACGTTGTG AGTTGGATAG TTGTGAAAG AGTCAAATGG CTCTCCTCAA GCGTATTCAA	2520
	CAAGGGGCTG AAGGATGCCC AGAAGGTACC CCATTGTATG GGATCTGATC TGGGGCCTCG	2580
35	GTGCACATGC TTTACATGTG TTTAGTCGAG GTTAAAAAAA CGTCTAGGCC CCCCGAACCA	2640
	CGGGGACGTG GTTTTCCTTT GAAAAACACG ATACGGGATC CACCGGTCGC CACCATGGGT	2700
40	AAAGGAGAAG AACTTTTCAC AGGAGTTGTC CCAATTCTTG TTGAATTAGA TGGTGATGTT	2760
	AATGGGCACA AATTTTCTGT CAGTGGAGAG GGTGAAGGTG ATGCAACATA CGGAAAACCT	2820
	ACCCTTAAAT TTATTTGCAC TACTGGAAAA CTACCTGTTT CATGGCCAAC ACTTGTCACT	2880
45	ACTTTCACCT ATGGTGTTCA ATGCTTTTCA AGATACCCAG ATCATATGAA ACGGCATGAC	2940
	TTTTTCAAGA GTGCCATGCC CGAAGGTTAT GTACAGGAAA GAACTATATT TTTCAAAGAT	3000
	GACGGGAACCT ACAAGACACG TGCTGAAGTC AAGTTTGAAG GTGATACCCT TGTTAATAGA	3060
50	ATCGAGTTAA AAGGTATGTA TTTTAAAGAA GATGGAAACA TTCTTGACA CAAATTGGAA	3120
	TACAACTATA ACTCACACAA TGTATACATC ATGGCAGACA AACAAAAGAA TGGAACCAAA	3180
55	GTAACTTCA AAATTAGACA CAACATTGAA GATGGAAGCG TTCAACTAGC AGACCATTAT	3240
	CAACAAAATA CTCCAATTGG CGATGGCCCT GTCCTTTTAC CAGACAACCA TTACCTGTCC	3300
	ACACAATCTG CCCTTTCGAA AGATCCCAAC GAAAAGAGAG ACCACATGGT CCTTCTTGAG	3360
60	TTTGTAACAG CTGCTGGGAT TACACATGGC ATGGATGAAC TATACAAGTC CGGATCTAGA	3420
	TAAGTGTATC GATGGATCCG AAGGCGGGGA CAGCAGTGCA GTGGTGACA GAAAGCAAGT	3480
65	GATCTAGGCC AGCAGCCTCC CTAAAGGGAC TTCAGCCCAC AAAGCCAAAC TTGTGGCTTT	3540
	AATACAAGCT CTGTAAATGG TAAAAAATAA AAAGTCTACA CGGACAGCAG GTATGCTCTT	3600

	GCCACTGTAC AGAGCAATAT ACAGACAAAG AGAACTGTTG ACATCTGCAG AGAAAGACCT	3660
	AAGATGCTGT GGCTAAAAGA AATCAGATGG CAAATCTAAC CGCCCAGGCA TCCTAAAGAG	3720
5	CAATGATCCT GACAGTCTGA AGACTATCAA GTTATAGACA AATTAAGACT GGTAACAAAA	3780
	ACCCTGTATA AAATAGTAAA AACTGAAAAA AGAAAACTAG TCCTCTCATG AGAAGACAGA	3840
10	CCTGACATCT ACTGAAAAAT AGACTTTACT GGAAAAATA TGTGTATGAA TACCTTCTAG	3900
	TTTTTGTGAA CGTTCTCAAG ATGGATAAAA GCTTTTCCTT GTAAAACGAG ACTGATCAGA	3960
	TAGTCATCAA GAAGATTGTT AAAGAAAATT TTCCAAGGTT CGGAGTGCCA AAAGCAATAG	4020
15	TGTCAGATAA TGGTCCTGCC TTTGTTGCCC AGGTAAGTCA GGGTGTGGCC AAGTATTTAG	4080
	AGGTCAAATG AAAATTCCAT TGTGTGTACA GACCTCAGAG CTCAGGAAAG ATAAAAAAGA	4140
20	ATAAATAAAA CTCTAAACAG ACCTTGACAA AATTAATCCT AGAGACTGGC ACAGACTTAC	4200
	TTGGTACTCC TTCCCCTTGC CCTATTTAGA ACTGAGAATA CTCCTCTTG ATTTCGGTTTT	4260
	ACTCTTTTTA AGATCCTTTA TGGGGCTCCT ATGCCATCAC TGTCTTAAAT GATGTGTTTA	4320
25	AACCTATGTT GTTATAATAA TGATCTATAT GTTAAGTTAA AAGGCTTGCA GGTGGTGCAG	4380
	AAAGAAGTCT GGTCACAACCT GGCTACAGTG AACAAAGCTGG GTACCCCAAG GACATCTTAC	4440
30	CAGTTCCAGC CAGAGATCTG ATCTACGATC CCCGGGTCGA CCCGGGTCGA CCCTGTGGAA	4500
	TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG	4560
	CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG	4620
35	AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC TAACTCCGCC	4680
	CATCCCGCCC CTAACCTCCG CCAGTTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT	4740
40	TTTTATTTAT GCAGAGGCCG AGGCCGCTC GGCCTCTGAG CTATTCCAGA AGTAGTGAGG	4800
	AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTTCAC GCTGCCGCAA GCACTCAGGG	4860
	CGCAAGGGCT GCTAAAGGAA GCGGAACACG TAGAAAGCCA GTCCGCAGAA ACGGTGCTGA	4920
45	CCCCGATGA ATGTCAGCTA CTGGGCTATC TGGACAAGGG AAAACGCAAG CGCAAAGAGA	4980
	AAGCAGGTAG CTTGCAGTGG GCTTACATGG CGATAGCTAG ACTGGGCGGT TTTATGGACA	5040
	GCAAGCGAAC CGGAATTGCC AGCTGGGGCG CCCTCTGGTA AGGTTGGGAA GCCCTGCAAA	5100
50	GTAAACTGGA TGGCTTTCTT GCCGCCAAGG ATCTGATGGC GCAGGGGATC AAGATCTGAT	5160
	CAAGAGACAG GATGAGGATC GTTTCGCATG ATTGAACAAG ATGGATTGCA CGCAGGTTCT	5220
55	CCGGCCGCTT GGGTGGAGAG GCTATTCGGC TATGACTGGG CACAACAGAC AATCGGCTGC	5280
	TCTGATGCCG CCGTGTTCGG GCTGTCAGCG CAGGGGCGCC CGGTTCTTTT TGTCAAGACC	5340
	GACCTGTCCG GTGCCCTGAA TGAAGTGCAG GACGAGGCAG CGCGGCTATC GTGGCTGGCC	5400
60	ACGACGGGCG TTCCTTGCGC AGCTGTGCTC GACGTTGTCA CTGAAGCGGG AAGGGACTGG	5460
	CTGCTATTGG GCGAAGTGCC GGGGCAGGAT CTCCTGTCAT CTCACCTTGC TCCTGCCGAG	5520
65	AAAGTATCCA TCATGGCTGA TGCAATGCGG CGGCTGCATA CGCTTGATCC GGCTACCTGC	5580

	CCATTGCGACC ACCAAGCGAA ACATCGCATC GAGCGAGCAC GTACTCGGAT GGAAGCCGGT	5640
	CTTGTGCGATC AGGATGATCT GGACGAAGAG CATCAGGGGC TCGCGCCAGC CGAACTGTTC	5700
5	GCCAGGCTCA AGGCGCGCAT GCCCCACGGC GAGGATCTCG TCGTGACCCA TGGCGATGCC	5760
	TGCTTGCCGA ATATCATGGT GGAAAATGGC CGCTTTTCTG GATTCATCGA CTGTGGCCGG	5820
	CTGGGTGTGG CGGACCCTA TCAGGACATA GCGTTGGCTA CCCGTGATAT TGCTGAAGAG	5880
10	CTTGGCGGCG AATGGGCTGA CCGCTTCCTC GTGCTTTACG GTATCGCCGC TCCCGATTCC	5940
	CAGCGCATCG CCTTCTATCG CCTTCTTGAC GAGTTCTTCT GAGCGGGACT CTGGGGTTCC	6000
15	AAATGACCGA CCAAGCGACG CCCAACCTGC CATCACGAGA TTTCGATTCC ACCGCCGCCT	6060
	TCTATGAAAG GTTGGGCTTC GGAATCGTTT TCCGGGACGG AATTCGTAAT CTGCTGCTTG	6120
	CAAACAAAAA AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT	6180
20	CTTTTTCCGA AGGTAAGTGG CTTACAGAGA GCGCAGATAC CAAATACTGT CCTTCTAGTG	6240
	TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC CGCCTACATA CCTCGCTCTG	6300
25	CTAATCCTGT TACCAGTGGC TGCTGCCAGT GGCATAAGT CGTGTCTTAC CGGGTTGGAC	6360
	TCAAGACGAT AGTTACCGGA TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA	6420
	CAGCCCAGCT TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA	6480
30	GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG CGGCAGGGTC	6540
	GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG CCTGGTATCT TTATAGTCCT	6600
35	GTCGGGTTTC GCCACCTCTG ACTTGAGCGT CGATTTTGTG GATGCTCGTC AGGGGGGCGG	6660
	AGCCTATGGA AAAACGCCAG CAACGCCGAG ATGCGCCGCC TCGAGTACAC CTGCGTCATG	6720
	CTGAGACCCT CAAGCCTCAC TAAAAGGGTC CCTGCCTAGT TCTGTTTACT AATCTGCCTT	6780
40	ATTCTGTTTT TGTTCCCATG TTAAAGATAG AGTAAATGCA GTATTCTCCA CATAGAGATA	6840
	TAGACTTCTG AAATTCTAAG ATTAGAATTA TTTACAAGAA GAAGTGGGGA A	6891
45	(2) INFORMATION FOR SEQ ID NO:17:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 6321 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
50	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
55		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
	TGAAGAATAA AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC	60
60	CCCTCCCATC TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT	120
	CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG	180
65	TTGTTAAACT TCCCCTATTC CCTCCCCATT CCCCCTCCCA GTTTGTGGTT TTTTCCTTTA	240

	AAAGCTTG TG AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG	300
	AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTTGCTGCT TTATTTTCGAC CCCAGAGCTC	360
5	TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TTAAATCTTA CCTTCTACAT TTTATGTATG	420
	GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT	480
10	TCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG	540
	CATTGGCCGG GAATTCGAAA ATCTTTCATT TGGTGCATTG GCCGGGAAAC AGCGCGACCA	600
	CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTTGG TCTGATGTCT	660
15	GTGTTCTGAT GTCTGTGTTC TGTTTCTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG	720
	ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
20	GGTGTCCACC GTCCGTTTCG CCTGGGAGAC GTCCCAGGAG GAACAGGGGA GGATCAGGGA	840
	CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTGGG GTTGCAGAT CGTGGGTTCG	900
	AGTCCCACCT CGTGCCAGT TCGAGATCG TGGGTTCGAG TCCCACCTCG TGTTTTGTTG	960
25	CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
	ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
30	CGTGGGTTCG AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
	CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTC TCTGACTCTG GTTCTGTGCG	1260
35	TTGGTAATTT TGTTTGTTTA CGTTTGT TGTGAGTCGT CTATGTTGTC TGTACTATC	1320
	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTTCTG	1380
40	CTTGGACTGA TGA CTGTTTTTAA GTTATGCCTT CTAAAATAAG CCTAAAAATC	1440
	CTGTCAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACTCGAGCTC	1500
	AAGCTTCGAA TTCTGCAGTC GACGGTACCG CGGGGATCAA TTCCGCCCCC CCCCTAACGT	1560
45	TACTGGCCGA AGCCGCTTGG AATAAGGCCG GTGTGCGTTT GTCTATATGT TATTTTCCAC	1620
	CATATTGCCG TCTTTTGGCA ATGTGAGGGC CCGGAAACCT GGCCCTGTCT TCTTGACGAG	1680
50	CATTCTAGG GGTCTTTCCC CTCTCGCCAA AGGAATGCAA GGTCTGTTGA ATGTCGTGAA	1740
	GGAAGCAGTT CCTCTGGAAG CTTCTTGAAG ACAAACAACG TCTGTAGCGA CCCTTTGCAG	1800
	GCAGCGGAAC CCCCCACCTG GCGACAGGTG CCTCTGCGGC CAAAAGCCAC GTGTATAAGA	1860
55	TACACCTGCA AAGGCGGCAC ACCCCAGTG CCACGTTGTG AGTTGGATAG TTGTGGAAG	1920
	AGTCAAATGG CTCTCCTCAA GCGTATTCAA CAAGGGGCTG AAGGATGCCC AGAAGGTACC	1980
60	CCATTGTATG GGATCTGATC TGGGGCCTCG GTGCACATGC TTTACATGTG TTTAGTCGAG	2040
	GTTAAAAAAA CGTCTAGGCC CCCC GAACCA CGGGGACGTG GTTTTCCTTT GAAAAACACG	2100
	ATACGGGATC CACCGGTCGC CACCATGGGT AAAGGAGAAG AACTTTTCAC AGGAGTTGTC	2160
65	CCAATTCTTG TTGAATTAGA TGGTGATGTT AATGGGCACA AATTTTCTGT CAGTGGAGAG	2220

	GGTGAAGGTG ATGCAACATA CGGAAACTT ACCCTTAAAT TTATTGTCAC TACTGGAAAA	2280
	CTACCTGTTT CATGGCCAAC ACTTGTCAC TTTTCTACTT ATGGTGTTC ATGCTTTTCA	2340
5	AGATACCCAG ATCATATGAA ACGGCATGAC TTTTCAAGA GTGCCATGCC CGAAGGTTAT	2400
	GTACAGGAAA GAACTATATT TTCAAAGAT GACGGGAAC ACAAGACACG TGCTGAAGTC	2460
10	AAGTTGAAG GTGATACCCT TGTTAATAGA ATCGAGTTAA AAGGTATTGA TTTTAAAGAA	2520
	GATGGAAACA TTCTTGACA CAAATTGGAA TACAATACT ACTCACACAA TGTATACATC	2580
	ATGGCAGACA AACAAAAGAA TGGAAACAAA GTTAACCTCA AAATTAGACA CAACATTGAA	2640
15	GATGGAAGCG TTCAACTAGC AGACCATTAT CAACAAAATA CTCCAATTGG CGATGGCCCT	2700
	GTCCTTTTAC CAGACAACCA TTACCTGTCC ACACAATCTG CCCTTTCGAA AGATCCCAAC	2760
20	GAAAAGAGAG ACCACATGGT CCTTCTTGAG TTTGTAACAG CTGCTGGGAT TACACATGGC	2820
	ATGGATGAAC TATACAAGTC CGGATCTAGA TAACTGTATC GATGGATCCG AAGGCGGGGA	2880
	CAGCAGTGCA GTGGTGGACA GAAAGCAAGT GATCTAGGCC AGCAGCCTCC CTAAAGGGAC	2940
25	TTCAGCCCAC AAAGCCAAAC TTGTGGCTTT AATACAAGCT CTGTAAATGG TAAAAAATA	3000
	AAAGTCTACA CGGACAGCAG GTATGCTCTT GCCACTGTAC AGAGCAATAT ACAGACAAAG	3060
30	AGAACTGTTG ACATCTGCAG AGAAAGACCT AAGATGCTGT GGCTAAAAGA AATCAGATGG	3120
	CAAATCTAAC CGCCAGGCA TCCTAAAGAG CAATGATCCT GACAGTCTGA AGACTATCAA	3180
	GTTATAGACA AATTAAGACT GGTAAAAAA ACCCTGTATA AAATAGTAAA AACTGAAAA	3240
35	AGAAACTAG TCCTCTCATG AGAAGACAGA CCTGACATCT ACTGAAAAAT AGACTTTACT	3300
	GGAAAAATA TGTGTATGAA TACCTTCTAG TTTTGTGAA CGTTCTCAAG ATGGATAAAA	3360
40	GCTTTTCCTT GTAAACGAG ACTGATCAGA TAGTCATCAA GAAGATTGTT AAAGAAAATT	3420
	TTCCAAGGTT CGGAGTGCCA AAAGCAATAG TGTAGATAA TGGTCCTGCC TTTGTTGCCC	3480
45	AGGTAAGTCA GGGTGTGGCC AAGTATTTAG AGGTCAAATG AAAATTCCAT TGTGTGTACA	3540
	GACCTCAGAG CTCAGGAAAG ATAAAAAAGA ATAAATAAAA CTCTAAACAG ACCTTGACAA	3600
	AATTAATCCT AGAGACTGGC ACAGACTTAC TTGGTACTCC TTCCCCTTGC CCTATTTAGA	3660
	ACTGAGAATA CTCCCTCTTG ATTGCGTTTT ACTCTTTTAA AGATCCTTTA TGGGGCTCCT	3720
50	ATGCCATCAC TGTCTTAAAT GATGTGTTA AACCTATGTT GTTATAATAA TGATCTATAT	3780
	GTTAAGTTAA AAGGCTTGCA GGTGGTGCAG AAAGAAGTCT GGTCACAACT GGCTACAGTG	3840
55	AACAAGCTGG GTACCCCAAG GACATCTTAC CAGTTCAGC CAGAGATCTG ATCTACGATC	3900
	CCCGGGTCGA CCCGGGTCGA CCCTGTGGAA TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC	3960
	AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG	4020
60	TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC	4080
	AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAATCCGC CCAGTCCGC	4140
65	CCATTCTCCG CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC	4200
	GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA	4260

	AAAGCTTCAC	GCTGCCGCAA	GCACTCAGGG	CGCAAGGGCT	GCTAAAGGAA	GCGGAACACG	4320
	TAGAAAGCCA	GTCCGCAGAA	ACGGTGCTGA	CCCCGGATGA	ATGTCAGCTA	CTGGGCTATC	4380
5	TGGACAAGGG	AAAACGCAAG	CGCAAAGAGA	AAGCAGGTAG	CTTGCAGTGG	GCTTACATGG	4440
	CGATAGCTAG	ACTGGGCGGT	TTTATGGACA	GCAAGCGAAC	CGGAATTGCC	AGCTGGGGCG	4500
10	CCCTCTGGTA	AGGTTGGGAA	GCCCTGCAAA	GTAAACTGGA	TGGCTTTCTT	GCCGCCAAGG	4560
	ATCTGATGGC	GCAGGGGATC	AAGATCTGAT	CAAGAGACAG	GATGAGGATC	GTTTCGCATG	4620
	ATTGAACAAG	ATGGATTGCA	CGCAGGTTCT	CCGGCCGCTT	GGGTGGAGAG	GCTATTCCGG	4680
15	TATGACTGGG	CACAACAGAC	AATCGGCTGC	TCTGATGCCG	CCGTGTTCCG	GCTGTCAGCG	4740
	CAGGGGCGCC	CGGTTCTTTT	TGTCAAGACC	GACCTGTCCG	GTGCCCTGAA	TGAACTGCAG	4800
20	GACGAGGCAG	CGCGGCTATC	GTGGCTGGCC	ACGACGGGCG	TTCCTTGCGC	AGCTGTGCTC	4860
	GACGTTGTCA	CTGAAGCGGG	AAGGGACTGG	CTGCTATTGG	GCGAAGTGCC	GGGGCAGGAT	4920
	CTCCTGTCAT	CTCACCTTGC	TCCTGCCGAG	AAAGTATCCA	TCATGGCTGA	TGCAATGCGG	4980
25	CGGCTGCATA	CGCTTGATCC	GGCTACCTGC	CCATTGACCC	ACCAAGCGAA	ACATCGCATC	5040
	GAGCGAGCAC	GTA CTGCGAT	GGAAGCCGGT	CTTGTCGATC	AGGATGATCT	GGACGAAGAG	5100
30	CATCAGGGGC	TCGCGCCAGC	CGAACTGTTT	GCCAGGCTCA	AGGCGCGCAT	GCCCCAGCGC	5160
	GAGGATCTCG	TCGTGACCCA	TGGCGATGCC	TGCTTGCCGA	ATATCATGGT	GGAAAATGGC	5220
	CGCTTTTCTG	GATTCATCGA	CTGTGGCCGG	CTGGGTGTGG	CGGACCGCTA	TCAGGACATA	5280
35	GCGTTGGCTA	CCCGTGATAT	TGCTGAAGAG	CTTGGCGGCG	AATGGGCTGA	CCGCTTCCTC	5340
	GTGCTTTACG	GTATCGCCGC	TCCCGATTCT	CAGCGCATCG	CCTTCTATCG	CCTTCTTGAC	5400
40	GAGTTCTTCT	GAGCGGGACT	CTGGGGTTCT	AAATGACCGA	CCAAGCGACG	CCCAACTTGC	5460
	CATCAGCAGA	TTTCGATTCC	ACCGCCGCCT	TCTATGAAAG	GTTGGGCTTC	GGAATCGTTT	5520
	TCCGGGACGG	AATTCGTAAT	CTGCTGCTTG	CAAACAAAAA	AACCACCGCT	ACCAGCGGTG	5580
45	GTTTGTGTTG	CGGATCAAGA	GCTACCAACT	CTTTTCCGA	AGGTAAGTGG	CTTCAGCAGA	5640
	GCGCAGATAC	CAAATACTGT	CCTTCTAGTG	TAGCCGTAGT	TAGGCCACCA	CTTCAAGAAC	5700
	TCTGTAGCAC	CGCCTACATA	CCTCGCTCTG	CTAATCCTGT	TACCAGTGGC	TGCTGCCAGT	5760
50	GGCGATAAGT	CGTGTCTTAC	CGGGTTGGAC	TCAAGACGAT	AGTTACCGGA	TAAGGCGCAG	5820
	CGGTGCGGGT	GAACGGGGGG	TTCTGTGACA	CAGCCCAGCT	TGGAGCGAAC	GACCTACACC	5880
55	GAACTGAGAT	ACCTACAGCG	TGAGCATTGA	GAAAGCGCCA	CGCTTCCCGA	AGGGAGAAAAG	5940
	GCGGACAGGT	ATCCGGTAAG	CGGCAGGGTC	GGAACAGGAG	AGCGCACGAG	GGAGCTTCCA	6000
	GGGGGAAACG	CCTGGTATCT	TTATAGTCCT	GTCGGGTTTC	GCCACCTCTG	ACTTGAGCGT	6060
60	CGATTTTTGT	GATGCTCGTC	AGGGGGGCGG	AGCCTATGGA	AAAACGCCAG	CAACGCCGAG	6120
	ATGCGCCGCC	TCGAGTACAC	CTGCGTCATG	CTGAGACCCT	CAAGCCTCAC	TAAAAGGGTC	6180
65	CCTGCCTAGT	TCTGTTTACT	AATCTGCCTT	ATTCTGTTTT	TGTTCCCATG	TTAAAGATAG	6240

AGTAAATGCA GTATTCTCCA CATAGAGATA TAGACTTCTG AAATTCTAAG ATTAGAATTA 6300
TTTACAAGAA GAAGTGGGGA A 6321

5 (2) INFORMATION FOR SEQ ID NO:18:

10 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 5754 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:18:	
20	TGAAGAATAA	AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC	60
	CCCTCCCATC	TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT	120
	CAACAGGCCA	GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG	180
25	TTGTTAAACT	TCCCCTATT C C TCCCCATT CCCCCTCCA GTTTGTGGTT TTTTCCTTTA	240
	AAAGCTTGTG	AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG	300
30	AGTTTCGACC	CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTCGAC CCCAGAGCTC	360
	TGGTCTGTGT	GCTTTCATGT CGCTGCTTTA TTAATCTTA CCTTCTACAT TTTATGTATG	420
	GTCTCAGTGT	CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT	480
35	TCCCTCGAGG	GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG	540
	CATTGGCCGG	GAATTCGAAA ATCTTTCATT TGGTGCAATG GCCGGGAAAC AGCGCGACCA	600
40	CCCAGAGGTC	CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTTGG TCTGATGTCT	660
	GTGTTCTGAT	GTCTGTGTTT TGTCTTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG	720
	ACGCTCAGTG	AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
45	GGTGTCCACC	GTCCGTTTCG C C TGGGAGAC GTCCAGGAG GAACAGGGGA GGATCAGGGA	840
	CGCCTGGTGG	ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCGAGAT CGTGGGTTTCG	900
50	AGTCCCACCT	CGTGCCAGT TGCAGATCG TGGGTTCGAG TCCCACCTCG TGTTTTGTTG	960
	CGAGATCGTG	GGTTCGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
	ACCTCGTGTT	TTGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
55	CGTGGGTTTCG	AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
	CTGATTCTTC	TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
60	TTTTTCTAAA	AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTGCG	1260
	TTGGTAATTT	TGTTTGTTTA CGTTTGTTTT TGTGAGTCGT CTATGTTGTC TGTTACTATC	1320
	TTGTTTTTGT	TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTTCTG	1380
65	CTTGGACTGA	TGACTGACGA CTGTTTTTAA GTTATGCCTT CTAATAAAG CCTAAAAATC	1440

	CTGTCAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACTCGAGCTC	1500
	AAGCTTCGAA TTCTGCAGTC GACGGTACCG CGGGCCCCGG ATCCACCGGT CGCCACCATG	1560
5	GGTAAAGGAG AAGAACTTTT CACAGGAGTT GTCCCAATTC TTGTTGAATT AGATGGTGAT	1620
	GTTAATGGGC ACAAATTTTC TGTCAGTGGA GAGGGTGAAG GTGATGCAAC ATACGGAAAA	1680
10	CTTACCCTTA AATTTATTTG CACTACTGGA AAACCTACCTG TTCCATGGCC AACACTTGTC	1740
	ACTACTTTCA CTTATGGTGT TCAATGCTTT TCAAGATACC CAGATCATAT GAAACGGCAT	1800
	GACTTTTTCA AGAGTGCCAT GCCCGAAGGT TATGTACAGG AAAGAACTAT ATTTTTCAAA	1860
15	GATGACGGGA ACTACAAGAC ACGTGCTGAA GTCAAGTTTG AAGGTGATAC CCTTGTTAAT	1920
	AGAATCGAGT TAAAAGGTAT TGATTTTAA GAAGATGGAA ACATTCTTGG ACACAAATTG	1980
20	GAATACAAC ATAACTCACA CAATGTATAC ATCATGGCAG ACAAACAAAA GAATGGAACC	2040
	AAAGTTAACT TCAAAATTAG ACACAACATT GAAGATGGAA GCGTTCAACT AGCAGACCAT	2100
	TATCAACAAA ATACTCCAAT TGGCGATGGC CCTGTCTTT TACCAGACAA CCATTACCTG	2160
25	TCCACACAAT CTGCCCTTTC GAAAGATCCC AACGAAAAGA GAGACCACAT GGTCTTCTT	2220
	GAGTTTGTA CAGCTGCTGG GATTACACAT GGCATGGATG AACTATACAA GTCCGGATCT	2280
30	AGATAACTGT ATCGATGGAT CCGAAGGCGG GGACAGCAGT GCAGTGGTGG ACAGAAAGCA	2340
	AGTGATCTAG GCCAGCAGCC TCCCTAAAGG GACTTCAGCC CACAAAGCCA AACTTGTTGGC	2400
	TTTAATACAA GCTCTGTAAA TGGTAAAAA AAAAAAGTCT ACACGGACAG CAGGTATGCT	2460
35	CTTGCCACTG TACAGAGCAA TATACAGACA AAGAGAACTG TTGACATCTG CAGAGAAAGA	2520
	CCTAAGATGC TGTGGCTAAA AGAAATCAGA TGGCAAATCT AACCGCCCAG GCATCCTAAA	2580
40	GAGCAATGAT CCTGACAGTC TGAAGACTAT CAAGTTATAG ACAAATTAAG ACTGGTAAAA	2640
	AAAACCCTGT ATAAAATAGT AAAAAGTGA AAAAGAAAAC TAGTCCTCTC ATGAGAAGAC	2700
	AGACCTGACA TCTACTGAAA AATAGACTTT ACTGGAAAAA ATATGTGTAT GAATACCTTC	2760
45	TAGTTTTTGT GAACGTTCTC AAGATGGATA AAAGCTTTTC CTTGTAAAC GAGACTGATC	2820
	AGATAGTCAT CAAGAAGATT GTTAAAGAAA ATTTTCCAAG GTTCGGAGTG CCAAAGCAA	2880
	TAGTGTCAGA TAATGGTCCT GCCTTTGTTG CCCAGGTAAG TCAGGGTGTG GCCAAGTATT	2940
50	TAGAGGTCAA ATGAAAATTC CATTGTGTGT ACAGACCTCA GAGCTCAGGA AAGATAAAAA	3000
	AGAATAAATA AAACCTCTAA CAGACCTTGA CAAAATTAAT CCTAGAGACT GGCACAGACT	3060
55	TACTTGGTAC TCCTTCCCCT TGCCCTATTT AGAACTGAGA ATACTCCCTC TTGATTCCGT	3120
	TTTACTCTTT TTAAGATCCT TTATGGGGCT CCTATGCCAT CACTGTCTTA AATGATGTGT	3180
	TTAAACCTAT GTTGTATATA TAATGATCTA TATGTTAAGT TAAAAGGCTT GCAGGTGGTG	3240
60	CAGAAAGAAG TCTGGTCACA ACTGGCTACA GTGAACAAGC TGGGTACCCC AAGGACATCT	3300
	TACCAGTTCC AGCCAGAGAT CTGATCTACG ATCCCCGGGT CGACCCGGGT CGACCCTGTG	3360
65	GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA GAAGTATGCA	3420

	AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG TCCCCAGGCT CCCCAGCAGG	3480
	CAGAAGTATG CAAAGCATGC ATCTCAATTA GTCAGCAACC ATAGTCCCGC CCCTAACTCC	3540
5	GCCCATCCCG CCCCTAACTC CGCCCAGTTC CGCCCATTCT CCGCCCCATG GCTGACTAAT	3600
	TTTTTTTATT TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG	3660
10	AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAAGCTT CACGCTGCCG CAAGCACTCA	3720
	GGGCGCAAGG GCTGCTAAAG GAAGCGGAAC ACGTAGAAAG CCAGTCCGCA GAAACGGTGC	3780
	TGACCCCGGA TGAATGTCAG CTACTGGGCT ATCTGGACAA GGGAAAACGC AAGCGCAAAG	3840
15	AGAAAGCAGG TAGCTTGCAAG TGGGCTTACA TGGCGATAGC TAGACTGGGC GGTTTTATGG	3900
	ACAGCAAGCG AACCGGAATT GCCAGCTGGG GCGCCCTCTG GTAAGGTTGG GAAGCCCTGC	3960
20	AAAGTAAACT GGATGGCTTT CTTGCCGCCA AGGATCTGAT GGCGCAGGGG ATCAAGATCT	4020
	GATCAAGAGA CAGGATGAGG ATCGTTTCGC ATGATTGAAC AAGATGGATT GCACGCAGGT	4080
	TCTCCGGCCG CTTGGGTGGA GAGGCTATTC GGCTATGACT GGGCACAACA GACAATCGGC	4140
25	TGCTCTGATG CCGCCGTGTT CCGGCTGTCA GCGCAGGGGC GCCCGGTTCT TTTTGTCAAG	4200
	ACCGACCTGT CCGGTGCCCT GAATGAACTG CAGGACGAGG CAGCGCGGCT ATCGTGGCTG	4260
30	GCCACGACGG GCGTTCCTTG CGCAGCTGTG CTCGACGTTG TCACTGAAGC GGGGAAGGGAC	4320
	TGGCTGCTAT TGGGCGAAGT GCCGGGGCAG GATCTCCTGT CATCTCACCT TGCTCCTGCC	4380
	GAGAAAGTAT CCATCATGGC TGATGCAATG CGGCGGCTGC ATACGCTTGA TCCGGCTACC	4440
35	TGCCCATTCTG ACCACCAAGC GAAACATCGC ATCGAGCGAG CACGTACTCG GATGGAAGCC	4500
	GGTCTTGTCG ATCAGGATGA TCTGGACGAA GAGCATCAGG GGCTCGCGCC AGCCGAACTG	4560
40	TTCGCCAGGC TCAAGGCGCG CATGCCCGAC GGCGAGGATC TCGTCGTGAC CCATGGCGAT	4620
	GCCTGCTTGC CGAATATCAT GGTGGAAAAT GGCCGCTTTT CTGGATTTCAT CGACTGTGGC	4680
	CGGCTGGGTG TGGCGGACCG CTATCAGGAC ATAGCGTTGG CTACCCGTGA TATTGCTGAA	4740
45	GAGCTTGCGC GCGAATGGGC TGACCGCTTC CTCGTGCTTT ACGGTATCGC CGCTCCCGAT	4800
	TCGCAGCGCA TCGCCTTCTA TCGCCTTCTT GACGAGTTCT TCTGAGCGGG ACTCTGGGGT	4860
	TCGAAATGAC CGACCAAGCG ACGCCCAACC TGCCATCAGC AGATTTCGAT TCCACCGCCG	4920
50	CCTTCTATGA AAGGTTGGGC TTCGGAATCG TTTTCCGGGA CGGAATTTCGT AATCTGCTGC	4980
	TTGCAAACAA AAAAACCACC GCTACCAGCG GTGGTTTGTT TGCCGGATCA AGAGCTACCA	5040
55	ACTCTTTTTC CGAAGGTAAC TGGCTTCAGC AGAGCGCAGA TACCAAATAC TGTCTTCTA	5100
	GTGTAGCCGT AGTTAGGCCA CCACTTCAAG AACTCTGTAG CACCGCCTAC ATACCTCGCT	5160
	CTGCTAATCC TGTTACCAGT GGCTGCTGCC AGTGGCGATA AGTCGTGTCT TACCGGGTTG	5220
60	GA CTCAAGAC GATAGTTACC GGATAAGGCG CAGCGGTCGG GCTGAACGGG GGGTTCTGTC	5280
	ACACAGCCCA GCTTGAGCG AACGACCTAC ACCGAACTGA GATACCTACA GCGTGAGCAT	5340
65	TGAGAAAGCG CCACGCTTCC CGAAGGGAGA AAGGCGGACA GGTATCCGGT AAGCGGCAGG	5400
	GTCGGAACAG GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA TCTTTATAGT	5460

CCTGTCGGGT TTCGCCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC GTCAGGGGGG 5520
CGGAGCCTAT GGAAAAACGC CAGCAACGCC GAGATGCGCC GCCTCGAGTA CACCTGCGTC 5580
5 ATGCTGAGAC CCTCAAGCCT CACTAAAAGG GTCCCTGCCT AGTTCTGTTT ACTAATCTGC 5640
CTTATTCTGT TTTTGTTCCT ATGTTAAAGA TAGAGTAAAT GCAGTATTCT CCACATAGAG 5700
10 ATATAGACTT CTGAAATTCT AAGATTAGAA TTATTTACAA GAAGAAGTGG GGAA 5754

(2) INFORMATION FOR SEQ ID NO:19:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 5754 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: DNA (genomic)

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

TGAAGAATAA AAAATTACTG GCCTCTTG TG AGAACATGAA CTTTCACCTC GGAGCCCACC 60
CCCTCCCATC TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT 120
30 CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG 180
TTGTTAAACT TCCCCTATTC CCTCCCCATT CCCCCTCCCA GTTTGTGGTT TTTTCCTTTA 240
35 AAAGCTTG TG AAAAATTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG 300
AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTGAC CCCAGAGCTC 360
TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TAAATCTTA CCTTCTACAT TTTATGTATG 420
40 GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT 480
TCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG 540
45 CATTGGCCGG GAATTCGAAA ATCTTTCATT TGGTGCAATT GCCGGGAAAC AGCGCGACCA 600
CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTGG TCTGATGTCT 660
GTGTTCTGAT GTCTGTGTTC TGTTTCTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG 720
50 ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA 780
GGTGTCCACC GTCCGTTTCG CCTGGGAGAC GTCCAGGAG GAACAGGGGA GGATCAGGGA 840
55 CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCAGAT CGTGGGTTCG 900
AGTCCCACCT CGTGCCAGT TGCAGATCG TGGGTTGAG TCCCACCTCG TGTTTTGTTG 960
CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC 1020
60 ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT 1080
CGTGGGTTCG AGTCCCACCT CGTGCAGAGG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT 1140
65 CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG 1200

	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTGCGC	1260
	TTGGTAATTT TGTTTGTTTA CGTTTGTTTT TGTGAGTCGT CTATGTTGTC TGTTACTATC	1320
5	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTTCTG	1380
	CTTGGACTGA TGAAGTACGA CTGTTTTTAA GTTATGCCTT CTAAAATAAG CCTAAAAATC	1440
10	CTGTGAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTT ACTCGAGCTC	1500
	AAGCTTCGAA TTCTGCAGTC GACGGTACCG CGGGCCCGGG ATCCACCGGT CGCCACCATG	1560
	GGTAAAGGAG AAGAACTTTT CACTGGAGTT GTCCCAATTC TTGTTGAATT AGATGGTGAT	1620
15	GTTAATGGGC ACAAATTTTC TGTCAGTGGA GAGGGTGAAG GTGATGCAAC ATACGGAAAA	1680
	CTTACCCTTA AATTTATTTG CACTACTGGA AAACCTACCTG TTCCATGGCC AACACTTGTC	1740
20	ACTACTTTCT CTTATGGTGT TCAATGCTTT TCAAGATACC CAGATCATAT GAAACGGCAT	1800
	GACTTTTTCA AGAGTGCCAT GCCCGAAGGT TATGTACAGG AAAGAAGTAT ATTTTTCAAA	1860
	GATGACGGGA ACTACAAGAC ACGTGTGAA GTCAAGTTG AAGGTGATAC CCTTGTTAAT	1920
25	AGAATCGAGT TAAAAGGTAT TGATTTTAAA GAAGATGGAA ACATTCTTGG ACACAAATTG	1980
	GAATACAACCT ATAACCTACA CAATGTATAC ATCATGGCAG ACAAACAAAA GAATGGAACC	2040
30	AAAGTTAACT TCAAAATTAG ACACAACATT GAAGATGGAA GCGTTCAACT AGCAGACCAT	2100
	TATCAACAAA ATACTCCAAT TGGCGATGGC CCTGTCTTTT TACCAGACAA CCATTACCTG	2160
	TCCACACAAT CTGCCCTTTC GAAAGATCCC AACGAAAAGA GAGACCACAT GGTCTTCTT	2220
35	GAGTTTGTAA CAGCTGCTGG GATTACACAT GGCATGGATG AACTATACAA GTCCGGATCT	2280
	AGATAACTGT ATCGATGGAT CCGAAGGCGG GGACAGCAGT GCAGTGGTGG ACAGAAAGCA	2340
	AGTGATCTAG GCCAGCAGCC TCCCTAAAGG GACTTCAGCC CACAAAGCCA AACTTGTGGC	2400
40	TTTAATACAA GCTCTGTAAA TGGTAAAAAA AAAAAAGTCT ACACGGACAG CAGGTATGCT	2460
	CTTGCCACTG TACAGAGCAA TATACAGACA AAGAGAACTG TTGACATCTG CAGAGAAAGA	2520
45	CCTAAGATGC TGTGGCTAAA AGAAATCAGA TGGCAAATCT AACCGCCCAG GCATCCTAAA	2580
	GAGCAATGAT CCTGACAGTC TGAAGACTAT CAAGTTATAG ACAAATTAAG ACTGGTAAAA	2640
	AAAACCTGT ATAAAATAGT AAAAAGTAA AAAAGAAAAC TAGTCCTCTC ATGAGAAGAC	2700
50	AGACCTGACA TCTACTGAAA AATAGACTTT ACTGGAAAAA ATATGTGTAT GAATACCTTC	2760
	TAGTTTTTGT GAACGTTCTC AAGATGGATA AAAGCTTTTC CTTGTAAAAC GAGACTGATC	2820
55	AGATAGTCAT CAAGAAGATT GTTAAAGAAA ATTTTCCAAG GTTCGGAGTG CCAAAGCAA	2880
	TAGTGTGAGA TAATGGTCTT GCCTTTGTTG CCCAGGTAAG TCAGGGTGTG GCCAAGTATT	2940
	TAGAGGTCAA ATGAAAATTC CATTGTGTGT ACAGACCTCA GAGCTCAGGA AAGATAAAAA	3000
60	AGAATAAATA AACTCTAAA CAGACCTTGA CAAAATTAAT CCTAGAGACT GGCACAGACT	3060
	TACTTGGTAC TCCTTCCCTT TGCCCTATTT AGAACTGAGA ATACTCCCTC TTGATTCGGT	3120
65	TTTACTCTTT TTAAGATCCT TTATGGGGCT CCTATGCCAT CACTGTCTTA AATGATGTGT	3180
	TTAAACCTAT GTTGTATATA TAATGATCTA TATGTTAAGT TAAAAGGCTT GCAGGTGGTG	3240

	CAGAAAGAAG	TCTGGTCACA	ACTGGCTACA	GTGAACAAGC	TGGGTACCCC	AAGGACATCT	3300
	TACCAGTTCC	AGCCAGAGAT	CTGATCTACG	ATCCCCGGGT	CGACCCGGGT	CGACCCTGTG	3360
5	GAATGTGTGT	CAGTTAGGGT	GTGGAAAGTC	CCCAGGCTCC	CCAGCAGGCA	GAAGTATGCA	3420
	AAGCATGCAT	CTCAATTAGT	CAGCAACCAG	GTGTGGAAAG	TCCCCAGGCT	CCCCAGCAGG	3480
10	CAGAAGTATG	CAAAGCATGC	ATCTCAATTA	GTCAGCAACC	ATAGTCCCGC	CCCTAACTCC	3540
	GCCCATCCCG	CCCCTAACTC	CGCCCAGTTC	CGCCCATTCT	CCGCCCCATG	GCTGACTAAT	3600
	TTTTTTTATT	TATGCAGAGG	CCGAGCCCGC	CTCGGCCTCT	GAGCTATTCC	AGAAGTAGTG	3660
15	AGGAGGCTTT	TTTGGAGGCC	TAGGCTTTTG	CAAAAAGCTT	CACGCTGCCG	CAAGCACTCA	3720
	GGGCGCAAGG	GCTGCTAAAG	GAAGCGGAAC	ACGTAGAAAG	CCAGTCCGCA	GAAACGGTGC	3780
20	TGACCCCGGA	TGAATGTCAG	CTACTGGGCT	ATCTGGACAA	GGGAAAACGC	AAGCGCAAAG	3840
	AGAAAGCAGG	TAGCTTGACG	TGGGCTTACA	TGGCGATAGC	TAGACTGGGC	GGTTTTATGG	3900
	ACAGCAAGCG	AACCGGAATT	GCCAGCTGGG	GCGCCCTCTG	GTAAGGTTGG	GAAGCCCTGC	3960
25	AAAGTAAACT	GGATGGCTTT	CTTGCCGCCA	AGGATCTGAT	GGCGCAGGGG	ATCAAGATCT	4020
	GATCAAGAGA	CAGGATGAGG	ATCGTTTCGC	ATGATTGAAC	AAGATGGATT	GCACGCAGGT	4080
30	TCTCCGGCCG	CTTGGGTGGA	GAGGCTATTC	GGCTATGACT	GGGCACAACA	GACAATCGGC	4140
	TGCTCTGATG	CCGCCGTGTT	CCGGCTGTCA	GCGCAGGGGC	GCCCGGTTCT	TTTTGTCAAG	4200
	ACCGACCTGT	CCGGTGCCCT	GAATGAACTG	CAGGACGAGG	CAGCGCGGCT	ATCGTGGCTG	4260
35	GCCACGACGG	GCGTTCCTTG	CGCAGCTGTG	CTCGACGTTG	TCACTGAAGC	GGGAAGGGAC	4320
	TGGCTGCTAT	TGGGCGAAGT	GCCGGGGCAG	GATCTCCTGT	CATCTCACCT	TGCTCCTGCC	4380
	GAGAAAGTAT	CCATCATGGC	TGATGCAATG	CGGCGGCTGC	ATACGCTTGA	TCCGGCTACC	4440
40	TGCCCCATTG	ACCACCAAGC	GAAACATCGC	ATCGAGCGAG	CACGTACTCG	GATGGAAGCC	4500
	GGTCTTGTCG	ATCAGGATGA	TCTGGACGAA	GAGCATCAGG	GGCTCGCGCC	AGCCGAAGTG	4560
45	TTCGCCAGGC	TCAAGGCGCG	CATGCCCCGAC	GGCGAGGATC	TCGTCTGTAC	CCATGGCGAT	4620
	GCCTGCTTGC	CGAATATCAT	GGTGGAAAAT	GGCCGCTTTT	CTGGATTTCAT	CGACTGTGGC	4680
	CGGCTGGGTG	TGGCGGACCG	CTATCAGGAC	ATAGCGTTGG	CTACCCGTGA	TATTGCTGAA	4740
50	GAGCTTGCGG	GCGAATGGGC	TGACCGCTTC	CTCGTGCTTT	ACGGTATCGC	CGCTCCCGAT	4800
	TCGCAGCGCA	TCGCCTTCTA	TCGCCTTCTT	GACGAGTTCT	TCTGAGCGGG	ACTCTGGGGT	4860
55	TCGAAATGAC	CGACCAAGCG	ACGCCCAACC	TGCCATCACG	AGATTTTCGAT	TCCACCGCCG	4920
	CCTTCTATGA	AAGGTTGGGC	TTCGGAATCG	TTTTCCGGGA	CGGAATTCGT	AATCTGCTGC	4980
60	TTGCAAACAA	AAAAACCACC	GCTACCAGCG	GTGGTTTGTT	TGCCGGATCA	AGAGCTACCA	5040
	ACTCTTTTTT	CGAAGGTAAC	TGGCTTCAGC	AGAGCGCAGA	TACCAAATAC	TGTCCTTCTA	5100
	GTGTAGCCGT	AGTTAGGCCA	CACTTCAAG	AACTCTGTAG	CACCGCCTAC	ATACCTCGCT	5160
65	CTGCTAATCC	TGTTACCAGT	GGCTGCTGCC	AGTGGCGATA	AGTCGTGTCT	TACCGGGTTG	5220

5 GACTCAAGAC GATAGTTACC GGATAAGGCG CAGCGGTCGG GCTGAACGGG GGGTTCGTGC 5280
ACACAGCCCA GCTTGGAGCG AACGACCTAC ACCGAACTGA GATACCTACA GCGTGAGCAT 5340
10 TGAGAAAGCG CCACGCTTCC CGAAGGGAGA AAGGCGGACA GGTATCCGGT AAGCGGCAGG 5400
GTCGGAACAG GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA TCTTTATAGT 5460
CCTGTCGGGT TTCGCCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC GTCAGGGGGG 5520
15 CGGAGCCTAT GGAAAAACGC CAGCAACGCC GAGATGCGCC GCCTCGAGTA CACCTGCGTC 5580
ATGCTGAGAC CCTCAAGCCT CACTAAAAGG GTCCTGCCT AGTTCTGTTT ACTAATCTGC 5640
CTTATTCTGT TTTTGTTCCT ATGTTAAAGA TAGAGTAAAT GCAGTATTCT CCACATAGAG 5700
ATATAGACTT CTGAAATTCT AAGATTAGAA TTATTACAA GAAGAAGTGG GGAA 5754

20 (2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4958 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
25 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

35 AGGCGGGGAC AGCAGTGCAG TGGTGGACAG AAAGCAAGTG ATCTAGGCCA GCAGCCTCCC 60
TAAAGGGACT TCAGCCCACA AAGCCAACT TGTGGCTTTA ATACAAGCTC TGTAATGGT 120
AAAAAAAAA AAGTCTACAC GGACAGCAGG TATGCTCTTG CCACTGTACA GAGCAATATA 180
40 CAGACAAAGA GAACTGTTGA CATCTGCAGA GAAAGACCTA AGATGCTGTG GCTAAAAGAA 240
ATCAGATGGC AAATCTAACC GCCCAGGCAT CCTAAAGAGC AATGATCCTG ACAGTCTGAA 300
GACTATCAAG TTATAGACAA ATTAAGACTG GTAAAAAAA CCCTGTATAA AATAGTAAAA 360
45 ACTGAAAAA GAAACTAGT CCTCTCATGA GAAGACAGAC CTGACATCTA CTGAAAAATA 420
GACTTTACTG GAAAAATAT GTGTATGAAT ACCTTCTAGT TTTTGTGAAC GTTCTCAAGA 480
50 TGGATAAAG CTTTCCTTG TAAAACGAGA CTGATCAGAT AGTCATCAAG AAGATTGTTA 540
AAGAAAATT TCCAAGGTTT GGAGTGCCAA AAGCAATAGT GTCAGATAAT GGTCTGCCT 600
TTGTTGCCCA GGTAAGTCAG GGTGTGGCCA AGTATTTAGA GGTCAAATGA AAATTCCATT 660
55 GTGTGTACAG ACCTCAGAGC TCAGGAAAGA TAAAAAGAA TAAATAAAC TCTAACAGA 720
CCTTGACAAA ATTAATCCTA GAGACTGGCA CAGACTTACT TGGTACTCCT TCCCCTTGCC 780
60 CTATTTAGAA CTGAGAATAC TCCCTCTTGA TTCGGTTTTA CTCTTTTAA GATCCTTTAT 840
GGGGCTCCTA TGCCATCACT GTCTTAAATG ATGTGTTTAA ACCTATGTTG TTATAATAAT 900
GATCTATATG TTAAGTTAAA AGGCTTGCAG GTGGTGCAGA AAGAAGTCTG GTCACAACTG 960
65 GCTACAGTGA ACAAGCTGGG TACCCCAAGG ACATCTTACC AGTTCCAGCC AGAGATCTGA 1020

	TCTACGATCC	CCGGGTCGAC	CCGGGTCGAC	CCTGTGGAAT	GTGTGTCAGT	TAGGGTGTGG	1080
	AAAGTCCCCA	GGCTCCCCAG	CAGGCAGAAG	TATGCAAAGC	ATGCATCTCA	ATTAGTCAGC	1140
5	AACCAGGTGT	GGAAAGTCCC	CAGGCTCCCC	AGCAGGCAGA	AGTATGCAAA	GCATGCATCT	1200
	CAATTAGTCA	GCAACCATAG	TCCCCGCCCT	AACTCCGCCC	ATCCCGCCCC	TAACTCCGCC	1260
10	CAGTTCCGCC	CATTCTCCGC	CCCATGGCTG	ACTAATTTTT	TTTATTTATG	CAGAGGCCGA	1320
	GGCCGCCTCG	GCCTCTGAGC	TATTCCAGAA	GTAGTGAGGA	GGCTTTTTTG	GAGGCCTAGG	1380
	CTTTTGCAA	AAGCTTCACG	CTGCCCAAG	CACTCAGGGC	GCAAGGGCTG	CTAAAGGAAG	1440
15	CGGAACACGT	AGAAAGCCAG	TCCGCAGAAA	CGGTGCTGAC	CCCGGATGAA	TGTCAGCTAC	1500
	TGGGCTATCT	GGACAAGGGA	AAACGCAAGC	GCAAAGAGAA	AGCAGGTAGC	TTGCAGTGGG	1560
20	CTTACATGGC	GATAGCTAGA	CTGGGCGGTT	TTATGGACAG	CAAGCGAACC	GGAATTGCCA	1620
	GCTGGGGCGC	CCTCTGGTAA	GGTTGGGAAG	CCCTGCAAAG	TAAACTGGAT	GGCTTTCTTG	1680
	CCGCCAAGGA	TCTGATGGCG	CAGGGGATCA	AGATCTGATC	AAGAGACAGG	ATGAGGATCG	1740
25	TTTCGCATGA	TTGAACAAGA	TGGATTGCAC	GCAGGTTCTC	CGGCCGCTTG	GGTGGAGAGG	1800
	CTATTCGGCT	ATGACTGGGC	ACAACAGACA	ATCGGCTGCT	CTGATGCCGC	CGTGTTCCGG	1860
30	CTGTCAGCGC	AGGGGCGCCC	GGTTCTTTTT	GTCAAGACCG	ACCTGTCCGG	TGCCCTGAAT	1920
	GAACTGCAGG	ACGAGGCAGC	GCGGCTATCG	TGGCTGGCCA	CGACGGGCGT	TCCTTGCGCA	1980
	GCTGTGCTCG	ACGTTGTCAC	TGAAGCGGGA	AGGGACTGGC	TGCTATTGGG	CGAAGTGCCG	2040
35	GGGCAGGATC	TCCTGTCATC	TCACCTTGCT	CCTGCCGAGA	AAGTATCCAT	CATGGCTGAT	2100
	GCAATGCGGC	GGCTGCATAC	GCTTGATCCG	GCTACCTGCC	CATTTCGACCA	CCAAGCGAAA	2160
	CATCGCATCG	AGCGAGCACG	TACTCGGATG	GAAGCCGGTC	TTGTTCGATCA	GGATGATCTG	2220
40	GACGAAGAGC	ATCAGGGGCT	CGCGCCAGCC	GAACTGTTTCG	CCAGGCTCAA	GGCGCGCATG	2280
	CCCGACGGCG	AGGATCTCGT	CGTGACCCAT	GGCGATGCCT	GCTTGCCGAA	TATCATGGTG	2340
45	GAAAATGGCC	GCTTTTCTGG	ATTCATCGAC	TGTGGCCGGC	TGGGTGTGGC	GGACCCTAT	2400
	CAGGACATAG	CGTTGGCTAC	CCGTGATATT	GCTGAAGAGC	TTGGCGGCGA	ATGGGCTGAC	2460
	CGCTTCCTCG	TGCTTTACGG	TATCGCCGCT	CCCGATTGCG	AGCGCATCGC	CTTCTATCGC	2520
50	CTTCTTGACG	AGTTCTTCTG	AGCGGGACTC	TGGGGTTCGA	AATGACCGAC	CAAGCGACGC	2580
	CCAACCTGCC	ATCACGAGAT	TTCGATTCCA	CCGCCGCCCT	CTATGAAAGG	TTGGGCTTCG	2640
55	GAATCGTTTT	CCGGGACGGA	ATTCGTAATC	TGCTGCTTGC	AAACAAAAAA	ACCACCGCTA	2700
	CCAGCGGTGG	TTTGTTTGCC	GGATCAAGAG	CTACCAACTC	TTTTTCCGAA	GGTAACTGGC	2760
	TTCAGCAGAG	CGCAGATACC	AAATACTGTC	CTTCTAGTGT	AGCCGTAGTT	AGGCCACCAC	2820
60	TTCAAGAACT	CTGTAGCACC	GCCTACATAC	CTCGCTCTGC	TAATCCTGTT	ACCAGTGGCT	2880
	GCTGCCAGTG	GCGATAAGTC	GTGTCTTACC	GGGTTGGACT	CAAGACGATA	GTTACCGGAT	2940
65	AAGGCGCAGC	GGTCGGGCTG	AACGGGGGGT	TCGTGCACAC	AGCCCAGCTT	GGAGCGAACG	3000

	ACCTACACCG AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA	3060
	GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA GCGCACGAGG	3120
5	GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG TCGGGTTTCG CCACCTCTGA	3180
	CTTGAGCGTC GATTTTTGTG ATGCTCGTCA GGGGGGCGGA GCCTATGGAA AAACGCCAGC	3240
10	AACGCCGAGA TGCGCCGCCT CGAGTACACC TGCCTCATGC TGAGACCCTC AAGCCTCACT	3300
	AAAAGGGTCC CTGCCTAGTT CTGTTTACTA ATCTGCCTTA TTCTGTTTTT GTTCCCATGT	3360
	TAAAGATAGA GTAAATGCAG TATTCTCCAC ATAGAGATAT AGACTTCTGA AATTCTAAGA	3420
15	TTAGAATTAT TTACAAGAAG AAGTGGGGAA TGAAGAATAA AAAATTACTG GCCTCTTGTG	3480
	AGAACATGAA CTTTCACCTC GGAGCCCACC CCCTCCCATC TGGAAAACAT ACTTGAGAAA	3540
20	AACATTTTCT GGAACAACCA CAGAATGTTT CAACAGGCCA GATGTATTGC CAAACACAGG	3600
	ATATGACTCT TTGGTTGAGT AAATTTGTGG TTGTTAAACT TCCCCTATTC CCTCCCCATT	3660
	CCCCCTCCCA GTTTGTGGTT TTTTCCTTTA AAAGCTTGTG AAAAAATTGA GTCGTCGTCG	3720
25	AGACTCCTCT ACCCTGTGCA AAGGTGTATG AGTTTCGACC CCAGAGCTCT GTGTGCTTTC	3780
	TGTTGCTGCT TTATTTGAC CCCAGAGCTC TGGTCTGTGT GCTTTCATGT CGTGCTTTA	3840
30	TTAAATCTTA CTTCTACAT TTTATGTATG GTCTCAGTGT CTTCTTGGGT ACGCGGCTGT	3900
	CCCGGGACTT GAGTGTCTGA GTGAGGGTCT TCCCTCGAGG GTCTTTCATT TGGTACATGG	3960
35	GCCGGGAATT CGAGAATCTT TCATTTGGTG CATTGGCCGG GAATTCGAAA ATCTTTCATT	4020
	TGGTGCAATTG GCCGGGAAAC AGCGCGACCA CCCAGAGGTC CTAGACCCAC TTAGAGGTAA	4080
	GATTCTTTGT TCTGTTTTGG TCTGATGTCT GTGTTCTGAT GTCTGTGTTC TGTTTCTAAG	4140
	TCTGGTGCGA TCGCAGTTTC AGTTTTCGCG ACGCTCAGTG AGACCGCGCT CCGAGAGGGA	4200
40	GTGCGGGGTG GATAAGGATA GACGTGTCCA GGTGTCCACC GTCCGTTGCG CCTGGGAGAC	4260
	GTCCCAGGAG GAACAGGGGA GGATCAGGGA CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG	4320
45	ACCATTTGGG GTTGCGAGAT CGTGGGTTTC AGTCCCACCT CGTGCCAGT TGCAGATCG	4380
	TGGGTTCGAG TCCCACCTCG TGTTTTGTTG CGAGATCGTG GGTTCGAGTC CCACCTCGCG	4440
	TCTGGTCACG GGATCGTGGG TTCGAGTCCC ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT	4500
50	CGAGTCCCAC CTCGCTCTG GTCACGGGAT CGTGGGTTTC AGTCCCACCT CGTGCAGAGG	4560
	GTCTCAATTG GCCGGCCTTA GAGAGGCCAT CTGATTCTTC TGTTTTCTCT TTTTGTCTTA	4620
55	GTCTCGTGTC CGCTCTGTG GTGACTACTG TTTTCTAAA AATGGGACAA TCTGTGTCCA	4680
	CTCCCCTTTC TCTGACTCTG GTTCTGTGCG TTGGTAATTT TGTTTGTTTA CGTTTGTTTT	4740
	TGTGAGTCGT CTATGTTGTC TGTTACTATC TTGTTTTTGT TTGTGGTTTA CGTTTCTGT	4800
60	GTGTGTCTTG TGTGTCTCTT TGTGTTTCTA CTGGACTGA TGAAGTACGA CTGTTTTTAA	4860
	GTTATGCCCT CTAAAATAAG CCTAAAAATC CTGTCAGATC CCTATGCTGA CCACTTCCTT	4920
65	TCAGATCAAC AGCTGCCCTT ACGTATCGAT GGATCCGA	4958

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7080 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

15	GAATACAAGC TTGCATGCCT GCAGGTCGAC TCTAGAGGAT CTTGAAGAAT AAAAAATTAC	60
	TGGCCTCTTG TGAGAACATG AACTTTCACC TCGGAGCCCA CCCCCTCCCA TCTGGAAAAC	120
20	ATACTTGAGA AAAACATTTT CTGGAACAAC CACAGAATGT TTCAACAGGC CAGATGTATT	180
	GCCAAACACA GGATATGACT CTTTGTTTGA GTAAATTTGT GGTTGTTAAA CTTCCCCTAT	240
	TCCCTCCCCA TTCCCCCTCC CAGTTTGTGG TTTTTCCTT TAAAAGCTTG TGAAAAATTT	300
25	GAGTCGTCGT CGAGACTCCT CTACCCTGTG CAAAGGTGTA TGAGTTTCGA CCCAGAGCT	360
	CTGTGTGCTT TCTGTTGCTG CTTTATTTTCG ACCCCAGAGC TCTGGTCTGT GTGCTTTCAT	420
30	GTCGCTGCTT TATTAAATCT TACCTTCTAC ATTTTATGTA TGGTCTCAGT GTCTTCTTGG	480
	GTACGCGGCT GTCCCGGGAC TTGAGTGTCT GAGTGAGGCT CTTCCCTCGA GGGTCTTTCA	540
	TTTGGTACAT GGGCCGGGAA TTCGAGAATC TTTCATTTGG TGCATTGGCC GGAATTCGA	600
35	AAATCTTTCA TTTGGTGCAT TGGCCGGGAA ACAGCGCGAC CACCCAGAGG TCCTAGACCC	660
	ACTTAGAGGT AAGATTCTTT GTTCTGTTTT GGTCTGATGT CTGTGTTCTG ATGTCTGTGT	720
40	TCTGTTTCTA AGTCTGGTGC GATCGCAGTT TCAGTTTTCG GGACGCTCAG TGAGACCGCG	780
	CTCCGAGAGG GAGTGCGGGG TGGATAAGGA TAGACGTGTC CAGGTGTCCA CCGTCCGTTC	840
	GCCCTGGGAG ACGTCCCAGG AGGAACAGGG GAGGATCAGG GACGCCTGGT GGACCCCTTT	900
45	GAAGGCCAAG AGACCATTG GGGTTGCGAG ATCGTGGGTT CGAGTCCCAC CATCGATGGT	960
	GCAGAGGGTC TCAATTGGCC GGCCTTAGAA TTACGGATCT AGCATGATTG AACAAGATGG	1020
50	ATTGCACGCA GGTCTCCGG CCGCTTGGGT GGAGAGGCTA TTCGGCTATG ACTGGGCACA	1080
	ACAGACAATC GGCTGCTCTG ATGCCGCCGT GTTCCGGCTG TCAGCGCAGG GGCGCCCGGT	1140
	TCTTTTGTG AAGACCGACC TGTCCGGTGC CCTGAATGAA CTGCAGGACG AGGCAGCGCG	1200
55	GCTATCGTGG CTGGCCACGA CGGGCGTTCC TTGCGCAGCT GTGCTCGACG TTGTCACTGA	1260
	AGCGGGAAGG GACTGGCTGC TATTGGGCGA AGTGCCGGGG CAGGATCTCC TGTCACTCA	1320
60	CCTTGCTCCT GCCGAGAAAG TATCCATCAT GGCTGATGCA ATGCGGCGGC TGCATACGCT	1380
	TGATCCGGCT ACCTGCCCAT TCGACCACCA AGCGAAACAT CGCATCGAGC GAGCACGTAC	1440
	TCGGATGGAA GCCGGTCTTG TCGATCAGGA TGATCTGGAC GAAGAGCATC AGGGGCTCGC	1500
65	GCCAGCCGAA CTGTTGCGCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG ATCTCGTCGT	1560

	GACCCATGGC GATGCCTGCT TGCCGAATAT CATGGTGGAA AATGGCCGCT TTTCTGGATT	1620
	CATCGACTGT GGCCGGCTGG GTGTGGCGGA CCGCTATCAG GACATAGCGT TGGCTACCCG	1680
5	TGATATTGCT GAAGAGCTTG GCGGCGAATG GGCTGACCGC TTCCTCGTGC TTTACGGTAT	1740
	CGCCGCTCCC GATTGCGAGC GCATCGCCTT CTATCGCCTT CTTGACGAGT TCTTCTGAGC	1800
10	GGGACTCTGG GGTTCGTAAT GACCGACCAA GCGACGCCA ACCTGCCATC ACGAGATTTC	1860
	GATTCCACCG CCGCCTTCTA TGAAAGGTTG GGCTTCGGAG TTAGCTTGTT TCTTTACTGT	1920
	TTGTCAATTC TATTATTTCA ATACAGAACA ATAGCTTCTA TAACTGAAAT ATATTTGCTA	1980
15	TTGTATATTA TGATTGTCCC TCGAACCATG AACACTCCTC CAGCTGAATT TCACAATTCC	2040
	TCTGTCATCT GCCAGGCCAT TAAGTTATTC ATGGAAGATC TTTGAGGAAC ACTGCAAGTT	2100
20	CATATCATAA ACACATTTGA AATTGAGTAT TGTTTTGCAT TGTATGGAGC TATGTTTTGC	2160
	TGTATCCTCA GAAAAAAGT TTGTTATAAA GCATTACAC CCATAAAAAG ATAGATTTAA	2220
	ATATTCCAGC TATAGGAAAG AAAGTGCCTC TGCTCTTAC TCTAGTCTCA GTTGGCTCCT	2280
25	TCACATGCAT GCTTCTTTAT TTCTCCTATT TTGTCAAGAA AATAATAGGT CACGTCTTGT	2340
	TCTCACTTAT GTCCTGCCTA GCATGGCTCA GATGCACGTT GTAGATACAA GAAGGATCAA	2400
30	ATGAAACAGA CTTCTGGTCT GTTACTACAA CCATAGTAAT AAGCACACTA ACTAATAATT	2460
	GCTAATTATG TTTTCCATCT CTAAGGTTCC CACATTTTTC TGTTTTCTTA AAGATCCCAT	2520
	TATCTGGTTG TAACTGAAGC TCAATGGAAC ATGAGCAATA TTTCCAGTC TTCTCTCCCA	2580
35	TCCAACAGTC CTGATGGATT AGCAGAACAG GCAGAAAACA CATTGTTACC CAGAATTAAA	2640
	AACTAATATT TGCTCTCCAT TCAATCCAAA ATGGACCTAT TGAAACTAAA ATCTAACCCA	2700
	ATCCCATTA AATGATTTCTA TGGCGTCAAA GGTCAAACCT CTGAAGGGAA CCTGTGGGTG	2760
40	GGTCACAATT CAGGCTATAT ATTCCCCAGG GCTCAGCCAG TGTCTGTACA TACACAACGG	2820
	ATCCTGTGGA CAGCTCACCT AGCTGCAATG GCTACAGGCT CCCGGACGTC CCTGCTCCTG	2880
45	GCTTTTGGCC TGCTCTGCCT GCCCTGGCTT CAAGAGGGCA GTGCCTTCCC AACCATTCCC	2940
	TTATCCAGGC TTTTGGACAA CGCTATGCTC CGCGCCCATC GTCTGCACCA GCTGGCCTTT	3000
	GACACCTACC AGGAGTTTGA AGAAGCCTAT ATCCCAAAGG AACAGAAGTA TTCATTCTTG	3060
50	CAGAACCCCC AGACCTCCCT CTGTTTCTCA GAGTCTATTC CGACACCCTC CAACAGGGAG	3120
	GAAACACAAC AGAAATCCAA CCTAGAGCTG CTCCGCATCT CCCTGCTGCT CATCCAGTCG	3180
55	TGGCTGGAGC CCGTGCAGTT CCTCAGGAGT GTCTTCGCCA ACAGCTTGGT GTACGGCGCC	3240
	TCTGACAGCA ACGTCTATGA CCTCCTAAAG GACCTAGAGG AAGGCATCCA AACGCTGATG	3300
	GGGAGGCTGG AAGATGGCAG CCCCCGACT GGGCAGATCT TCAAGCAGAC CTACAGCAAG	3360
60	TTCGACACAA ACTCACACAA CGATGACGCA CTA CTCAAGA ACTACGGGCT GCTCTACTGC	3420
	TTCAGGAAGG ACATGGACAA GGTCGAGACA TTCCTGCGCA TCGTGCAGTG CCGCTCTGTG	3480
65	GAGGGCAGCT GTGGCTTCTA GCTGCCCGGG TGGCATCCTG TGACCCCTCC CCAGTGCCTC	3540
	TCCTGGCCCT GGAAGTTGCC ACTCCAGTGC CCACCAGCCT TGTCTAATA AAATTAAGTT	3600

	GCATCAAAAA AAAAAAAAAAG CTAGCGGCCG CTAGACTTCT GAAATTCTAA GATTAGAATT	3660
	ATTTACAAGA AGAAGTGGGG AATGAAGAAT AAAAAATTAC TGGCCTCTTG TGAGAACATG	3720
5	AACTTTCACC TCGGAGCCCA CCCCTCCCA TCTGGAAAAC ATACTTGAGA AAAACATTTT	3780
	CTGGAACAAC CACAGAATGT TTCAACAGGC CAGATGTATT GCCAAACACA GGATATGACT	3840
10	CTTTGGTTGA GTAAATTTGT GGTTGTTAAA CTTCCCTAT TCCCTCCCA TTCCCCCTCC	3900
	CAGTTTGTGG TTTTTTCCTT TAAAAGCTTG TGAAAAATTT GAGTCGTCGT CGAGACTCCT	3960
	CTACCCTGTG CAAAGGTGTA TGAGTTTCGA CCCAGAGCT CTGTGTGCTT TCTGTTGCTG	4020
15	CTTTATTTTCG ACCCCAGAGC TCTGGTCTGT GTGCTTTCAT GTCGCTGCTT TATTAAATCT	4080
	TACCTTCTAC ATTTTATGTA TGGTCTCAGT GTCTTCTTGG GTACGCGGCT GTCCCGGGAC	4140
20	TTGAGTGTCT GAGTGAGGGT CTTCCCTCGA GGGTCTTTCA TTTGGTACAT GGGCCGGGAA	4200
	TTCGAGAATC TTTCATTTGG TGCATTGGCC GGGAATTCGA AAATCTTTCA GATCCCCGGG	4260
	TACCGAGCTC GAATTCGGT CTCCCTATAG TGAGTCGTAT TAATTTTCGAT AAGCCAGCTG	4320
25	CATTAATGAA TCGGCCAACG CGCGGGGAGA GCGGTTTTGC GTATTGGGCG CTCTTCCGCT	4380
	TCCTCGCTCA CTGACTCGCT GCGCTCGGTC GTTCGGCTGC GGCAGCGGT ATCAGCTCAC	4440
30	TCAAAGGCGG TAATACGGTT ATCCACAGAA TCAGGGGATA ACGCAGGAAA GAACATGTGA	4500
	GCAAAAGGCC AGCAAAAGGC CAGGAACCGT AAAAAGGCCG CGTTGCTGGC GTTTTTCCAT	4560
	AGGCTCCGCC CCCCTGACGA GCATCACAAA AATCGACGCT CAAGTCAGAG GTGGCGAAAC	4620
35	CCGACAGGAC TATAAAGATA CCAGGCGTTT CCCCTTGAA GCTCCCTCGT GCGCTCTCCT	4680
	GTTCCGACCC TGCCGCTTAC CGGATACCTG TCCGCCTTTC TCCCTTCGGG AAGCGTGGCG	4740
	CTTTCTCATA GCTCACGCTG TAGGTATCTC AGTTCGGTGT AGGTGCTTCG CTCCAAGCTG	4800
40	GGCTGTGTGC ACGAACCCCC CGTTCAGCCC GACCGCTGCG CTTATCCGG TAACTATCGT	4860
	CTTGAGTCCA ACCCGGTAAG ACACGACTTA TCGCCACTGG CAGCAGCCAC TGGTAACAGG	4920
45	ATTAGCAGAG CGAGGTATGT AGGCGGTGCT ACAGAGTTCT TGAAGTGGTG GCCTAACTAC	4980
	GGCTACACTA GAAGGACAGT ATTTGGTATC TCGCTCTGC TGAAGCCAGT TACCTTCGGA	5040
	AAAAGAGTTG GTAGCTCTTG ATCCGGCAAA CAAACCACCG CTGGTAGCGG TGGTTTTTTT	5100
50	GTTTGCAAGC AGCAGATTAC GCGCAGAAAA AAAGGATCTC AAGAAGATCC TTTGATCTTT	5160
	TCTACGGGGT CTGACGCTCA GTGGAACGAA AACTCACGTT AAGGGATTTT GGTCATGAGA	5220
55	TTATCAAAAA GGATCTTCAC CTAGATCCTT TTAAATTAAA AATGAAGTTT TAAATCAATC	5280
	TAAAGTATAT ATGAGTAAAC TTGGTCTGAC AGTTACCAAT GCTTAATCAG TGAGGCACCT	5340
	ATCTCAGCGA TCTGTCTATT TCGTTCATCC ATAGTTGCCT GACTCCCCGT CGTGTAGATA	5400
60	ACTACGATAC GGGAGGGCTT ACCATCTGGC CCCAGTGCTG CAATGATACC GCGAGACCCA	5460
	CGCTCACCGG CTCCAGATTT ATCAGCAATA AACCAGCCAG CCGGAAGGGC CGAGCGCAGA	5520
65	AGTGGTCTCG CAACTTTATC CGCCTCCATC CAGTCTATTA ATTGTTGCCG GGAAGCTAGA	5580

GTAAGTAGTT CGCCAGTTAA TAGTTTGCGC AACGTTGTTG CCATTGCTAC AGGCATCGTG 5640
GTGTCACGCT CGTCGTTTGG TATGGCTTCA TTCAGCTCCG GTTCCCAACG ATCAAGGCGA 5700
5 GTTACATGAT CCCCCATGTT GTGCAAAAAA GCGGTTAGCT CCTTCGGTCC TCCGATCGTT 5760
GTCAGAAGTA AGTTGGCCGC AGTGTATCA CTCATGGTTA TGGCAGCACT GCATAATTCT 5820
CTTACTGTCA TGCCATCCGT AAGATGCTTT TCTGTGACTG GTGAGTACTC AACCAAGTCA 5880
10 TTCTGAGAAT AGTGTATGCG GCGACCGAGT TGCTCTTGCC CGGCGTCAAT ACGGGATAAT 5940
ACCGCGCCAC ATAGCAGAAC TTTAAAAGTG CTCATCATTG GAAAACGTTT TTCGGGGCGA 6000
15 AAACTCTCAA GGATCTTACC GCTGTTGAGA TCCAGTTCGA TGTAACCCAC TCGTGCACCC 6060
AACTGATCTT CAGCATCTTT TACTTTCACC AGCGTTTCTG GGTGAGCAAA AACAGGAAGG 6120
CAAAATGCCG CAAAAAAGGG AATAAGGGCG ACACGGAAAT GTTGAATACT CATACTCTTC 6180
20 CTTTTTCAAT ATTATTGAAG CATTTATCAG GGTATTGTG TCATGAGCGG ATACATATTT 6240
GAATGTATTT AGAAAAATAA ACAAATAGGG GTTCCGCGCA CATTTCCTCCG AAAAGTGCCA 6300
25 CCTGACGCTT AAGAAACCAT TATTATCATG ACATTAACCT ATAAAAATAG GCGTATCACG 6360
AGGCCCTTTC GTCTCGCGCG TTTTCGGTGAT GACGGTGAAA ACCTCTGACA CATGCAGCTC 6420
CCGGAGACGG TCACAGCTTG TCTGTAAGCG GATGCCGGGA GCAGACAAGC CCGTCAGGGC 6480
30 GCGTCAGCGG GTGTTGGCGG GTGTCGGGGC TGGCTTAACT ATGCGGCATC AGAGCAGATT 6540
GTACTGAGAG TGCACCATAT CGACGCTCTC CCTTATGCGA CTCCTGCATT AGGAAGCAGC 6600
CCAGTAGTAG GTTGAGGCCG TTGAGCACCG CCGCCGAAG GAATGGTGCA AGGAGATGGC 6660
35 GCCCAACAGT CCCCCGGCCA CGGGGCCTGC CACCATACCC ACGCCGAAAC AAGCGCTCAT 6720
GAGCCCGAAG TGGCGAGCCC GATCTTCCCC ATCGGTGATG TCGGCGATAT AGGCGCCAGC 6780
40 AACC GCACCT GTGGCGCCGG TGATGCCGGC CACGATGCGT CCGGCGTAGA GGATCTGGCT 6840
AGCGATGACC CTGCTGATTG GTTCGCTGAC CATTTCCGGG GTGCGGAACG GCGTTACCAG 6900
AAACTCAGAA GGTTCGTCCA ACCAAACCGA CTCTGACGGC AGTTTACGAG AGAGATGATA 6960
45 GGGTCTGCTT CAGTAAGCCA GATGCTACAC AATTAGGCTT GTACATATTG TCGTTAGAAC 7020
GCGGCTACAA TTAATACATA ACCTTATGTA TCATACACAT ACGATTTAGG TGACACTATA 7080

50

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

55

(A) LENGTH: 6795 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

60

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

65

AATGAAAGAC CCCACCTGTA GGTTCGGCAA GCTAGCTTAA GTAACGCCAT TTTGCAAGGC

60

	ATGGAAAAAT ACATAACTGA GAATAGAGAA GTTCAGATCA AGGTCAGGAA CAGATGGAAC	120
	AGCTGAATAT GGGCCAAACA GGATATCTGT GGTAAGCAGT TCCTGCCCCG GCTCAGGGCC	180
5	AAGAACAGAT GGAACAGCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG	240
	CCCCGGCTCA GGGCCAAGAA CAGATGGTCC CCAGATGCGG TCCAGCCCTC AGCAGTTTCT	300
10	AGAGAACCAT CAGATGTTTC CAGGGTGCCC CAAGGACCTG AAATGACCCT GTGCCATTAT	360
	TGAACTAACC AATCAGTTCG CTTCTCGCTT CTGTTCGCGC GCTTCTGCTC CCCGAGCTCA	420
	ATAAAGAGC CCACAACCCC TCACTCGGGG CGCCAGTCCCT CCGATTGACT GAGTCGCCCCG	480
15	GGTACCCGTG TATCCAATAA ACCCTCTTGC AGTTGCATCC GACTTGTGGT CTCGCTGTTC	540
	CTTGGGAGGG TCTCCTCTGA GTGATTGACT ACCCGTCAGC GGGGGTCTTT CATTTGGGGG	600
20	CTCGTCCGGG ATCGGGAGAC CCCTGCCCAG GGACCACCGA CCCACCACCG GGAGGTAAGC	660
	TGGCCAGCAA CTTATCTGTG TCTGTCCGAT TGTCTAGTGT CTATGACTGA TTTTATGCGC	720
	CTGCGTCGGT ACTAGTTAGC TAACTAGCTC TGTATCTGGC GGACCCGTGG TGGAAGTGAC	780
25	GAGTTCGGAA CACCCGGCCG CAACCCTGGG AGACGTCCCA GGAGGAACAG GGGAGGATCA	840
	GGGACGCCTG GTGGACCCCT TTGAAGGCCA AGAGACCATT TGGGGTTGCG AGATCGTGGG	900
	TTGAGTCCC ACCTCGTGCC CAGTTGCAG AGTCGTGGGT CGAGTCCCAC CTCGTGTTTT	960
30	GTTGCGAGAT CGTGGGTTCG AGTCCCACCT CGCGTCTGGT CACGGGATCG TGGGTTGAG	1020
	TCCCACCTCG TGTTTTGTG CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG	1080
35	GGATCGTGGG TTCGAGTCCC ACCTCGTGCA GAGGGTCTCA ATTGGCCGGC CTTAGAGAGG	1140
	CCATCTGATT CTTCTGGTTT CTCTTTTGT CTTAGTCTCG TGTCGGCTCT GTTGTGACT	1200
	ACTGTTTTTC TAAAAATGGG ACAATCTGTG TCCACTCCCC TTTCTCTGAC TCTGGTTCTG	1260
40	TCGCTTGGA ATTTTGTGTTG TTTACGTTT TTTTGTGAG TCGTCTATGT TGTCTGTTAC	1320
	TATCTTGTTT TTGTTGTGG TTTACGTTT CTGTGTGTGT CTTGTGTGTC TCTTTGTGTT	1380
45	CAGACTTGGA CTGATGACTG ACGACTGTTT TTAAGTTATG CTTCTAAAA TAAGCCTAAA	1440
	AATCCTGTCA GATCCCTATG CTGACCACTT CCTTTCAGAT CAACAGCTGC CCTTACTCGA	1500
	GCTCAAGCTT CGAATTCTGC AGTCGACGGT ACCGCGGCCG CTAACATAA GCCCATTCTC	1560
50	CAAGGTACGT AGCGGGGATC AATTCCGCCC CCCCCTAAC GTTACTGGCC GAAGCCGCTT	1620
	GGAATAAGGC CGGTGTGCGT TTGTCTATAT GTTATTTTCC ACCATATTGC CGTCTTTTGG	1680
55	CAATGTGAGG GCGCGGAAAC CTGGCCCTGT CTTCTTGACG AGCATTCCTA GGGGTCTTTC	1740
	CCCTCTCGCC AAAGGAATGC AAGGTCTGTT GAATGTCGTG AAGGAAGCAG TTCCTCTGGA	1800
	AGCTTCTTGA AGACAAACAA CGTCTGTAGC GACCTTTGTC AGGCAGCGGA ACCCCCCACC	1860
60	TGGCGACAGG TGCTCTGCG GCCAAAAGCC ACGTGATAA GATACACCTG CAAAGGCGGC	1920
	ACAACCCAG TGCCACGTTG TGAGTTGGAT AGTTGTGGAA AGAGTCAAAT GGCTCTCCTC	1980
65	AAGCGTATTC AACAAGGGGC TGAAGGATGC CCAGAAGGTA CCCCATGTGA TGGGATCTGA	2040

	TCTGGGGCCT	CGGTGCACAT	GCTTTACATG	TGTTTAGTCG	AGGTTAAAAA	AACGTCTAGG	2100
	CCCCCGAAC	CACGGGGACG	TGGTTTTCCCT	TTGAAAAACA	CGATACGGGA	TCCACCGGTC	2160
5	GCCACCATGG	GTAAAGGAGA	AGAACTTTTC	ACAGGAGTTG	TCCCAATTCT	TGTTGAATTA	2220
	GATGGTGATG	TTAATGGGCA	CAAATTTTCT	GTCAGTGGAG	AGGGTGAAGG	TGATGCAACA	2280
	TACGGAAAAAC	TTACCCTTAA	ATTTATTTGC	ACTACTGGAA	AACTACCTGT	TCCATGGCCA	2340
10	ACACTTGTCA	CTACTTTCAC	TTATGGTGTT	CAATGCTTTT	CAAGATACCC	AGATCATATG	2400
	AAACGGCATG	ACTTTTTCAA	GAGTGCCATG	CCCGAAGGTT	ATGTACAGGA	AAGAACTATA	2460
15	TTTTTCAAAG	ATGACGGGAA	CTACAAGACA	CGTGCTGAAG	TCAAGTTTGA	AGGTGATACC	2520
	CTTGTTAATA	GAATCGAGTT	AAAAGGTATT	GATTTTAAAG	AAGATGGAAA	CATTCTTGGA	2580
	CACAAATTGG	AATACAATA	TAACACACAC	AATGTATACA	TCATGGCAGA	CAAACAAAAG	2640
20	AATGGAACCA	AAGTTAACTT	CAAAATTAGA	CACAACATTG	AAGATGGAAG	CGTTCAACTA	2700
	GCAGACCATT	ATCAACAAAA	TACTCCAATT	GGCGATGGCC	CTGTCCTTTT	ACCAGACAAC	2760
25	CATTACCTGT	CCACACAATC	TGCCCTTTCG	AAAGATCCCA	ACGAAAAGAG	AGACCACATG	2820
	GTCCTTCTTG	AGTTTGTAA	AGCTGCTGGG	ATTACACATG	GCATGGATGA	ACTATACAAG	2880
	TCCGGATCTA	GATAACTGTA	TCGATGGATC	CGAAGGCGGG	GACAGCAGTG	CAGTGGTGGG	2940
30	CAGAAAGCAA	GTGATCTAGG	CCAGCAGCCT	CCCTAAAGGG	ACTTCAGCCC	ACAAAGCCAA	3000
	ACTTGTGGCT	TTAATACAAG	CTCTGTAAAT	GGTAAAAAAA	AAAAAGTCTA	CACGGACAGC	3060
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35	AGAGAAAGAC	CTAAGATGCT	GTGGCTAAAA	GAAATCAGAT	GGCAAATCTA	ACCGCCCAGG	3180
	CATCCTAAAG	AGCAATGATC	CTGACAGTCT	GAAGACTATC	AAGTTATAGA	CAAATTAAGA	3240
40	CTGGTAAAAA	AAACCCTGTA	TAAAATAGTA	AAAACGTAAA	AAAGAAAAC	AGTCCTCTCA	3300
	TGAGAAGACA	GACCTGACAT	CTACTGAAAA	ATAGACTTTA	CTGGAAAAAA	TATGTGTATG	3360
	AATACCTTCT	AGTTTTTGTG	AACGTTCTCA	AGATGGATAA	AAGCTTTTCC	TTGTAAAACG	3420
45	AGACTGATCA	GATAGTCATC	AAGAAGATTG	TTAAAGAAAA	TTTCCAAGG	TTCGGAGTGC	3480
	CAAAAGCAAT	AGTGTGAGAT	AATGGTCCTG	CCTTTGTGTC	CCAGGTAAGT	CAGGGTGTGG	3540
50	CCAAGTATTT	AGAGGTCAAA	TGAAAATTCC	ATTGTGTGTA	CAGACCTCAG	AGCTCAGGAA	3600
	AGATAAAAAA	GAATAAATAA	AACTCTAAAC	AGACCTTGAC	AAAATTAATC	CTAGAGACTG	3660
	GCACAGACTT	ACTTGGTACT	CCTTCCCCTT	GCCCTATTTA	GAAGTGAAG	TACTCCCCTT	3720
55	TGATTCGGTT	TTACTCTTTT	TAAGATCCTT	TATGGGGCTC	CTATGCCATC	ACTGTCTTAA	3780
	ATGATGTGTT	TAAACCTATG	TTGTTATAAT	AATGATCTAT	ATGTTAAGTT	AAAAGGCTTG	3840
60	CAGGTGGTGC	AGAAAGAAGT	CTGGTCACAA	CTGGCTACAG	TGAACAAGCT	GGGTACCCCA	3900
	AGGACATCTT	ACCAGTTCCA	GCCAGAGATC	TGATCTACGA	TCCCCGGGTC	GACCCGGGTC	3960
	GACCCTGTGG	AATGTGTGTC	AGTTAGGGTG	TGGAAAGTCC	CCAGGCTCCC	CAGCAGGCAG	4020
65	AAGTATGCAA	AGCATGCATC	TCAATTAGTC	AGCAACCAGG	TGTGGAAAGT	CCCCAGGCTC	4080

	CCCAGCAGGC	AGAAGTATGC	AAAGCATGCA	TCTCAATTAG	TCAGCAACCA	TAGTCCCGCC	4140
5	CCTAACTCCG	CCCATCCCGC	CCCTAACTCC	GCCCAGTTCC	GCCCATTCTC	CGCCCCATGG	4200
	CTGACTAATT	TTTTTTATTT	ATGCAGAGGC	CGAGGCCGCC	TCGGCCTCTG	AGCTATTCCA	4260
	GAAGTAGTGA	GGAGGCTTTT	TTGGAGGCCT	AGGCTTTTGC	AAAAAGCTTC	ACGCTGCCGC	4320
10	AAGCACTCAG	GGCGCAAGGG	CTGCTAAAGG	AAGCGGAACA	CGTAGAAAGC	CAGTCCGCAG	4380
	AAACGGTGCT	GACCCCGGAT	GAATGTCAGC	TACTGGGCTA	TCTGGACAAG	GGAAAACGCA	4440
15	AGCGCAAAGA	GAAAGCAGGT	AGCTTGCACT	GGGCTTACAT	GGCGATAGCT	AGACTGGGCG	4500
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	AAGCCCTGCA	AAGTAACTG	GATGGCTTTC	TTGCCGCCAA	GGATCTGATG	GCGCAGGGGA	4620
20	TCAAGATCTG	ATCAAGAGAC	AGGATGAGGA	TCGTTTCGCA	TGATTGAACA	AGATGGATTG	4680
	CACGCAGGTT	CTCCGGCCGC	TTGGGTGGAG	AGGCTATTCT	GCTATGACTG	GGCACAACAG	4740
25	ACAATCGGCT	GCTCTGATGC	CGCCGTGTTC	CGGCTGTCAG	CGCAGGGGCG	CCCGGTTCTT	4800
	TTTGTCAAGA	CCGACCTGTC	CGGTGCCCTG	AATGAACTGC	AGGACGAGGC	AGCGCGGCTA	4860
	TCGTGGCTGG	CCACGACGGG	CGTTCCTTGC	GCAGCTGTGC	TCGACGTTGT	CACTGAAGCG	4920
30	GGAAGGGACT	GGCTGCTATT	GGGCGAAGTG	CCGGGGCAGG	ATCTCCTGTC	ATCTCACCTT	4980
	GCTCCTGCCG	AGAAAGTATC	CATCATGGCT	GATGCAATGC	GGCGGCTGCA	TACGCTTGAT	5040
	CCGGCTACCT	GCCCATTCTG	CCACCAAGCG	AAACATCGCA	TCGAGCGAGC	ACGTACTCGG	5100
35	ATGGAAGCCG	GTCTTGTCGA	TCAGGATGAT	CTGGACGAAG	AGCATCAGGG	GCTCGCGCCA	5160
	GCCGAACCTG	TCGCCAGGCT	CAAGGCGCGC	ATGCCCGACG	GCGAGGATCT	CGTCGTGACC	5220
40	CATGGCGATG	CCTGCTTGCC	GAATATCATG	GTGGAAAATG	GCCGCTTTTC	TGGATTTCATC	5280
	GACTGTGGCC	GGCTGGGTGT	GGCGGACCGC	TATCAGGACA	TAGCGTTGGC	TACCCGTGAT	5340
	ATTGCTGAAG	AGCTTGCGCG	CGAATGGGCT	GACCGCTTCC	TCGTGCTTTA	CGGTATCGCC	5400
45	GCTCCCGATT	CGCAGCGCAT	CGCCTTCTAT	CGCCTTCTTG	ACGAGTTCTT	CTGAGCGGGA	5460
	CTCTGGGGTT	CGAAATGACC	GACCAAGCGA	CGCCCAACCT	GCCATCACGA	GATTTGATT	5520
50	CCACCGCCGC	CTTCTATGAA	AGGTTGGGCT	TCGGAATCGT	TTCCGGGAC	GGAATTCGTA	5580
	ATCTGCTGCT	TGCAAAACAAA	AAAACCACCG	CTACCAGCGG	TGGTTTGTTT	GCCGGATCAA	5640
	GAGCTACCAA	CTCTTTTTTC	GAAGGTAAC	GGCTTCAGCA	GAGCGCAGAT	ACCAAATACT	5700
55	GTCCTTCTAG	TGTAGCCGTA	GTTAGGCCAC	CACTTCAAGA	ACTCTGTAGC	ACCGCCTACA	5760
	TACCTCGCTC	TGCTAATCCT	GTTACCAGTG	GCTGCTGCCA	GTGGCGATAA	GTCGTGTCTT	5820
60	ACCGGGTTGG	ACTCAAGACG	ATAGTTACCG	GATAAGGCGC	AGCGGTCGGG	CTGAACGGGG	5880
	GGTTCGTGCA	CACAGCCCAG	CTTGGAGCGA	ACGACCTACA	CCGAACTGAG	ATACCTACAG	5940
	CGTGAGCATT	GAGAAAGCGC	CACGCTTCCC	GAAGGGAGAA	AGGCGGACAG	GTATCCGGTA	6000
65	AGCGGCAGGG	TCGGAACAGG	AGAGCGCACG	AGGGAGCTTC	CAGGGGGAAA	CGCCTGGTAT	6060

	CTTTATAGTC CTGTCGGGTT TCGCCACCTC TGACTTGAGC GTCGATTTTT GTGATGCTCG	6120
	TCAGGGGGGC GGAGCCTATG GAAAAACGCC AGCAACGCCG AGATGCGCCG CCTCGAGAAC	6180
5	CCTGGCCCTA TTATTGGGTG GACTAACCAT GGGGGGAATT GCCGCTGGAA TAGGAACAGG	6240
	GACTACTGCT CTAATGGCCA CTCAGCAATT CCAGCAGCTC CAAGCCGCAG TACAGGATGA	6300
10	TCTCAGGGAG GTTGAAAAAT CAATCTCTAA CCTAGAAAAG TCTCTCACTT CCCTGTCTGA	6360
	AGTTGTCCTA CAGAATCGAA GGGGCCTAGA CTTGTTATTT CTAAAAGAAG GAGGGCTGTG	6420
	TGCTGCTCTA AAAGAAGAAT GTTGCTTCTA TGCGGACCAC ACAGGACTAG TGAGAGACAG	6480
15	CATGGCCAAA TTGAGAGAGA GGCTTAATCA GAGACAGAAA CTGTTTGAGT CAACTCAAGG	6540
	ATGGTTTGAG GGACTGTTTA ACAGATCCCC TTGGTTTACC ACCTTGATAT CTACCATTAT	6600
20	GGGACCCCTC ATTGTACTCC TAATGATTTT GCTCTTCGGA CCCTGCATTG TTAATCGATT	6660
	AGTCCAATTT GTTAAAGACA GGATATCAGT GGTCCAGGCT CTAGTTTGA CTCAACAATA	6720
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25	CCAGAAAAAG GGGGG	6795
	(2) INFORMATION FOR SEQ ID NO:23:	
	(i) SEQUENCE CHARACTERISTICS:	
30	(A) LENGTH: 9093 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
35	(ii) MOLECULE TYPE: DNA (genomic)	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
40	AATGAAAGAC CCCACCTGTA GGTTCGGCAA GCTAGCTTAA GTAACGCCAT TTTGCAAGGC	60
	ATGGAAAAAT ACATAACTGA GAATAGAGAA GTTCAGATCA AGGTCAGGAA CAGATGGAAC	120
45	AGCTGAATAT GGGCCAAACA GGATATCTGT GGTAAGCAGT TCCTGCCCCG GCTCAGGGCC	180
	AAGAACAGAT GGAACAGCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG	240
50	CCCCGGCTCA GGGCCAAGAA CAGATGGTCC CCAGATGCGG TCCAGCCCTC AGCAGTTTCT	300
	AGAGAACCAT CAGATGTTTC CAGGGTGCCC CAAGGACCTG AAATGACCCT GTGCCTTATT	360
	TGAATAACC AATCAGTTCG CTTCTCGCTT CTGTTGCGGC GCTTCTGCTC CCCGAGCTCA	420
55	ATAAAAGAGC CCACAACCCC TCACTCGGGG CGCCAGTCCT CCGATTGACT GAGTCGCCCC	480
	GGTACCCGTG TATCCAATAA ACCCTCTTGC AGTTGCATCC GACTTGTGGT CTCGCTGTTC	540
60	CTTGGGAGGG TCTCCTCTGA GTGATTGACT ACCCGTCAGC GGGGGTCTTT CATTTGGGGG	600
	CTCGTCCGGG ATCGGGAGAC CCCTGCCCAG GGACCACCGA CCCACCACCG GGAGGTAAGC	660
	TGGCCAGCAA CTTATCTGTG TCTGTCCGAT TGTCTAGTGT CTATGACTGA TTTTATGCGC	720
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	GGGACGCCTG GTGGACCCCT TTGAAGGCCA AGAGACCATT TGGGGTTGCG AGATCGTGGG	900
5	TTCGAGTCCC ACCTCGTGCC CAGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGTGTTTT	960
	GTTGCGAGAT CGTGGGTTG AGTCCCACCT CGCGTCTGGT CACGGGATCG TGGGTTTCGAG	1020
10	TCCCACCTCG TGTTTTGTTG CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG	1080
	GGATCGTGGG TTCGAGTCCC ACCTCGTGCA GAGGGTCTCA ATTGGCCGGC CTTAGAGAGG	1140
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15	ACTGTTTTTC TAAAAATGGG ACAATCTGTG TCCACTCCCC TTTCTCTGAC TCTGGTTCTG	1260
	TCGCTTGTA ATTTTGTGTTG TTTACGTTT TTTTGTGAG TCGTCTATGT TGTCTGTTAC	1320
20	TATCTTGTGTTT TTGTTTGTGG TTTACGTTT CTGTGTGTGT CTTGTGTGTC TCTTTGTGTT	1380
	CAGACTTGGA CTGATGACTG ACGACTGTTT TTAAGTTATG CCTTCTAAAA TAAGCCTAAA	1440
	AATCCTGTCA GATCCCTATG CTGACCACTT CCTTTCAGAT CAACAGCTGC CCTTACGTAT	1500
25	CGATGGATCC CTCGACTAAC TAATAGCCCA TTCTCCAAGG TCGAGCGGGA TCAATTCCGC	1560
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35	GCGACCCCTT GCAGGCAGCG GAACCCCCCA CCTGGCGACA GGTGCCTCTG CGGCCAAAAG	1860
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	GCGAATGGCG CTTTGCCTGG TTTCCGGCAC CAGAAGCGGT GCCGAAAGC TGGCTGGAGT	2340
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	GGCGCTGGGT CGGTTACGGC CAGGACAGTC GTTTGCCGTC TGAATTTGAC CTGAGCGCAT	2640
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5	TAACAGTTTC TTTATGGCAG GGTGAAACGC AGGTGCGCAG CGGCACCGCG CCTTTCGGCG	2940
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10	AAAACCCGAA ACTGTGGAGC GCCGAAATCC CGAATCTCTA TCGTGGGTG GTTGAAGTGC	3060
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60	ACCAGGCCGA AGCAGCGTTG TTGCAGTGCA CGGCAGATAC ACTTGCTGAT GCGGTGCTGA	4620
	TTACGACCGC TCACGCGTGG CAGCATCAGG GGAAAACCTT ATTTATCAGC CGGAAAACCT	4680
	ACCGGATTGA TGGTAGTGGT CAAATGGCGA TTACCGTTGA TGTTGAAGTG GCGAGCGATA	4740
65	CACCGCATCC GCGCGGATT GGCCTGAACT GCCAGCTGGC GCAGGTAGCA GAGCGGGTAA	4800

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5	GTCTGCGCTG	CGGGACGCGC	GAATTGAATT	ATGGCCCACA	CCAGTGGCGC	GGCGACTTCC	4980
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10	TGCACGCGGA	AGAAGGCACA	TGGCTGAATA	TCGACGGTTT	CCATATGGGG	ATTGGTGGCG	5100
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	GACCTCAGAG	CTCAGGAAAG	ATAAAAAAGA	ATAAATAAAA	CTCTAAACAG	ACCTTGACAA	5940
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40	ACTGAGAATA	CTCCCTCTTG	ATTCGGTTTT	ACTCTTTTTA	AGATCCTTTA	TGGGGCTCCT	6060
	ATGCCATCAC	TGTCTTAAAT	GATGTGTTTA	AACCTATGTT	GTTATAATAA	TGATCTATAT	6120
45	GTTAAGTTAA	AAGGCTTGCA	GGTGGTGCAG	AAAGAAGTCT	GGTCACAAC	GGCTACAGTG	6180
	AACAAGCTGG	GTACCCCAAG	GACATCTTAC	CAGTTCAGC	CAGAGATCTG	ATCTACGATC	6240
	CCCGGGTCGA	CCCGGGTCGA	CCCTGTGGAA	TGTGTGTCAG	TTAGGGTGTG	GAAAGTCCCC	6300
50	AGGCTCCCCA	GCAGGCAGAA	GTATGCAAAG	CATGCATCTC	AATTAGTCAG	CAACCAGGTG	6360
	TGGAAAGTCC	CCAGGCTCCC	CAGCAGGCAG	AAGTATGCAA	AGCATGCATC	TCAATTAGTC	6420
55	AGCAACCATA	GTCCCGCCCC	TAATCCGCC	CATCCGCCCC	CTAACTCCGC	CCAGTTCCGC	6480
	CCATTCTCCG	CCCCATGGCT	GACTAATTTT	TTTTATTAT	GCAGAGGCCG	AGGCCGCCTC	6540
	GGCTCTGAG	CTATTCCAGA	AGTAGTGAGG	AGGCTTTTTT	GGAGGCTTAG	GCTTTTGCAA	6600
60	AAAGCTTCAC	GCTGCCGCAA	GCACTCAGGG	CGCAAGGGCT	GCTAAAGGAA	GCGGAACACG	6660
	TAGAAAGCCA	GTCCGCAGAA	ACGGTGCTGA	CCCCGGATGA	ATGTCAGCTA	CTGGGCTATC	6720
65	TGGACAAGGG	AAAACGCAAG	CGCAAAGAGA	AAGCAGGTAG	CTTGCAAGTG	GCTTACATGG	6780

	CGATAGCTAG	ACTGGGCGGT	TTTATGGACA	GCAAGCGAAC	CGGAATTGCC	AGCTGGGGCG	6840
	CCCTCTGGTA	AGGTTGGGAA	GCCCTGCAAA	GTAACTGGA	TGGCTTTCTT	GCCGCCAAGG	6900
5	ATCTGATGGC	GCAGGGGATC	AAGATCTGAT	CAAGAGACAG	GATGAGGATC	GTTTCGCATG	6960
	ATTGAACAAG	ATGGATTGCA	CGCAGGTTCT	CCGGCCGCTT	GGGTGGAGAG	GCTATTCCGC	7020
10	TATGACTGGG	CACAACAGAC	AATCGGCTGC	TCTGATGCCG	CCGTGTCCG	GCTGTCAGCG	7080
	CAGGGGCGCC	CGTTCTTTT	TGTCAAGACC	GACCTGTCCG	GTGCCCTGAA	TGAACTGCAG	7140
	GACGAGGCAG	CGCGGCTATC	GTGGCTGGCC	ACGACGGGCG	TTCCTTGCGC	AGCTGTGCTC	7200
15	GACGTTGTCA	CTGAAGCGGG	AAGGGACTGG	CTGCTATTGG	GCGAAGTGCC	GGGGCAGGAT	7260
	CTCCTGTCAT	CTCACCTTGC	TCCTGCCGAG	AAAGTATCCA	TCATGGCTGA	TGCAATGCGG	7320
20	CGGCTGCATA	CGCTTGATCC	GGCTACCTGC	CCATTCGACC	ACCAAGCGAA	ACATCGCATC	7380
	GAGCGAGCAC	GTAATCGGAT	GGAAGCCGGT	CTTGTCGATC	AGGATGATCT	GGACGAAGAG	7440
	CATCAGGGGC	TCGCGCCAGC	CGAACTGTTC	GCCAGGCTCA	AGGCGCGCAT	GCCCGACGGC	7500
25	GAGGATCTCG	TCGTGACCCA	TGGCGATGCC	TGCTTGCCGA	ATATCATGGT	GGAAAATGGC	7560
	CGCTTTTCTG	GATTCATCGA	CTGTGGCCGG	CTGGGTGTGG	CGGACCGCTA	TCAGGACATA	7620
	GCGTTGGCTA	CCCGTGATAT	TGCTGAAGAG	CTTGGCGGCG	AATGGGCTGA	CCGCTTCCTC	7680
30	GTGCTTTACG	GTATCGCCGC	TCCCATTTCG	CAGCGCATCG	CCTTCTATCG	CCTTCTTGAC	7740
	GAGTTCTTCT	GAGCGGGACT	CTGGGGTTTC	AAATGACCGA	CCAAGCGACG	CCCAACCTGC	7800
35	CATCACGAGA	TTTCGATTCC	ACCGCCGCCT	TCTATGAAAG	GTTGGGCTTC	GGAATCGTTT	7860
	TCCGGGACGG	AATTCGTAAT	CTGCTGCTTG	CAAACAAAA	AACCACCGCT	ACCAGCGGTG	7920
	GTTTGTTTGC	CGGATCAAGA	GCTACCAACT	CTTTTCCGA	AGGTAAGTGG	CTTCAGCAGA	7980
40	GCGCAGATAC	CAAATACTGT	CCTTCTAGTG	TAGCCGTAGT	TAGGCCACCA	CTTCAAGAAC	8040
	TCTGTAGCAC	CGCCTACATA	CCTCGCTCTG	CTAATCCTGT	TACCAGTGGC	TGCTGCCAGT	8100
45	GGCGATAAGT	CGTGTCTTAC	CGGGTTGGAC	TCAAGACGAT	AGTTACCGGA	TAAGGCGCAG	8160
	CGGTCGGGCT	GAACGGGGGG	TTCGTGCACA	CAGCCCAGCT	TGGAGCGAAC	GACCTACACC	8220
	GAACTGAGAT	ACCTACAGCG	TGAGCATTGA	GAAAGCGCCA	CGCTTCCCGA	AGGGAGAAAAG	8280
50	GCGGACAGGT	ATCCGGTAAG	CGGCAGGGTC	GGAACAGGAG	AGCGCACGAG	GGAGCTTCCA	8340
	GGGGGAAACG	CCTGGTATCT	TTATAGTCCT	GTCGGGTTTC	GCCACCTCTG	ACTTGAGCGT	8400
55	CGATTTTTGT	GATGCTCGTC	AGGGGGGCGG	AGCCTATGGA	AAAACGCCAG	CAACGCCGAG	8460
	ATGCGCCGCC	TCGAGAACCC	TGGCCCTATT	ATTGGGTGGA	CTAACCATGG	GGGGAATTGC	8520
	CGCTGGAATA	GGAACAGGGA	CTACTGCTCT	AATGGCCACT	CAGCAATTCC	AGCAGCTCCA	8580
60	AGCCGCAGTA	CAGGATGATC	TCAGGGAGGT	TGAAAAATCA	ATCTCTAACC	TAGAAAAGTC	8640
	TCTCACTTCC	CTGTCTGAAG	TTGTCTTACA	GAATCGAAGG	GGCCTAGACT	TGTTATTTCT	8700
65	AAAAGAAGGA	GGGCTGTGTG	CTGCTCTAAA	AGAAGAATGT	TGCTTCTATG	CGGACCACAC	8760
	AGGACTAGTG	AGAGACAGCA	TGGCCAAATT	GAGAGAGAGG	CTTAATCAGA	GACAGAAACT	8820

5 GTTTGAGTCA ACTCAAGGAT GGTTCGAGGG ACTGTTTAAC AGATCCCCTT GGTTTACCAC 8880
CTTGATATCT ACCATTATGG GACCCCTCAT TGTACTCCTA ATGATTTTGC TCTTCGGACC 8940
CTGCATTCTT AATCGATTAG TCCAATTTGT TAAAGACAGG ATATCAGTGG TCCAGGCTCT 9000
AGTTTTGACT CAACAATATC ACCAGCTGAA GCCTATAGAG TACGAGCCAT AGATAAAATA 9060
10 AAAGATTTTA TTTAGTCTCC AGAAAAAGGG GGG 9093

(2) INFORMATION FOR SEQ ID NO:24:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 46 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
20 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

25 GACTAACCTT GATTCCCTGG AGGCGGGGGT CTTTCATTTG GGGGCT 46

(2) INFORMATION FOR SEQ ID NO:25:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4834 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
35 (ii) MOLECULE TYPE: DNA (genomic)

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

TGAAGAATAA AAAATTACTG GCCTCTTGTC AGAACATGAA CTTTCACCTC GGAGCCCACC 60
45 CCCTCCCATC TGGAAAACAT ACTTGAGAAA AACATTTTCT GGAACAACCA CAGAATGTTT 120
CAACAGGCCA GATGTATTGC CAAACACAGG ATATGACTCT TTGGTTGAGT AAATTTGTGG 180
TTGTTAAACT TCCCCTATTC CCTCCCCATT CCCCCTCCCA GTTTGTGGTT TTTTCCTTTA 240
50 AAAGCTTGTC AAAAATTTGA GTCGTCGTCG AGACTCCTCT ACCCTGTGCA AAGGTGTATG 300
AGTTTCGACC CCAGAGCTCT GTGTGCTTTC TGTGCTGCT TTATTTCGAC CCCAGAGCTC 360
55 TGGTCTGTGT GCTTTCATGT CGCTGCTTTA TTAAATCTTA CCTTCTACAT TTTATGTATG 420
GTCTCAGTGT CTTCTGGGT ACGCGGCTGT CCCGGGACTT GAGTGTCTGA GTGAGGGTCT 480
TCCCTCGAGG GTCTTTCATT TGGTACATGG GCCGGGAATT CGAGAATCTT TCATTTGGTG 540
60 CATTGGCCCG GAATTCGAAA ATCTTTCATT TGGTGCATTG GCCGGGAAAC AGCGCGACCA 600
CCCAGAGGTC CTAGACCCAC TTAGAGGTAA GATTCTTTGT TCTGTTTGG TCTGATGTCT 660
65 GTGTTCTGAT GTCTGTGTTT TGTCTTAAG TCTGGTGCGA TCGCAGTTTC AGTTTTGCGG 720

	ACGCTCAGTG AGACCGCGCT CCGAGAGGGA GTGCGGGGTG GATAAGGATA GACGTGTCCA	780
	GGTGTCCACC GTCCGTTCGC CCTGGGAGAC GTCCCAGGAG GAACAGGGGA GGATCAGGGA	840
5	CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG ACCATTTGGG GTTGCGAGAT CGTGGGTTCTG	900
	AGTCCCACCT CGTGCCCACT TGCAGATCG TGGGTTTCGAG TCCCACCTCG TGTTTTGTG	960
	CGAGATCGTG GGTTCGAGTC CCACCTCGCG TCTGGTCACG GGATCGTGGG TTCGAGTCCC	1020
10	ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT CGAGTCCCAC CTCGCGTCTG GTCACGGGAT	1080
	CGTGGGTTCTG AGTCCCACCT CGTGCAAGAG GTCTCAATTG GCCGGCCTTA GAGAGGCCAT	1140
15	CTGATTCTTC TGGTTTCTCT TTTTGTCTTA GTCTCGTGTC CGCTCTTGTT GTGACTACTG	1200
	TTTTTCTAAA AATGGGACAA TCTGTGTCCA CTCCCCTTTC TCTGACTCTG GTTCTGTCTG	1260
	TTGGTAATTT TGTGTTGTTA CGTTTGTGTT TGTGAGTCGT CTATGTTGTC TGTTACTATC	1320
20	TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT GTGTGTCTTG TGTGTCTCTT TGTGTTTCTG	1380
	CTTGGACTGA TGAAGTACGA CTGTTTTTAA GTTATGCCTT CTAAAATAAG CCTAAAAATC	1440
	CTGTGAGATC CCTATGCTGA CCACTTCCTT TCAGATCAAC AGCTGCCCTG CCTCCCACTC	1500
25	CAACTCCAGA GAGCAGCCAG CGGGTCACAG TGGTCCCGCC CATGAACCTG GAGCCTAGGG	1560
	AAAAATGAGC TCGGAAATCC GGAGCAAATG AGGAGTGGTC CCTGAGAAGT CAGTGGCCTA	1620
30	AATGTTGTGG CTGCTGAAGC AAAAGAAGAG GAGGCTGTTT GAGTAGCCGG CCAAGAGCGC	1680
	CGCGGGTTCC CAGGCAGCTT CTCATTCCCC TGTCCTCTCC ATCCCGTCTC TTGTAAACAG	1740
	AAAAACTGCT TTCACTTTGA GATATGAGTG GCCCGATACA GCCAGCTGTG AGAGCTGTAC	1800
35	TCCCTTCCCT GCCCCACGTG TTTTCTCTTC TCAGGCGACC CCTCCCTGAG CTGCTGGCAG	1860
	TGAGTCTGTT CTAAGCTCCA GTGAGGGAGG CATCCGCCCA CTTGGGGCTT CTGTCCAAGG	1920
40	TAAGGAGCAC CTGTGAGTCT AACTGCCAGG CTCTGATGGG GGTCTCGTCT CTGTGGGACT	1980
	AGAAAGTGTC CCAACAATCT GACCAAGGTA ACAGGAAGTT AAGACAAAGA CAGAGACCAA	2040
	AGTCAGAATC AGAGCTGTGC TGTGAGACAA AAAGATAAAA AAAATAAAAT GCTGGCCACA	2100
45	AAAGTCAGGA AAAGTAGAAA ACTTAGATAG TACCTGGCAA CAAAAGAAAG CTTTGTGGCTA	2160
	AAGATCAACG TGTATACTGT AAAGAAAATG AGCACTGGGT GAGAGACTGC CCCAACAAAA	2220
50	AGAAGAGGAG CCCCCCTCAT GACCAAACCC TTCACCTGTT CGTGGCTAAA AGTAAAGAGA	2280
	TAACAAAAGG GTTGCTAACA CAGAAGCTGA GTCCTTAAAA GAGTCCGGTG GCCTACCTGT	2340
	TGAAGCAGCT AAAAAAGAGA CTGTGTTTCA TACTCTCCA CTGACCAGTG CAAAACAAGC	2400
55	TAAAAAGTTC CTGGGCACTG CGGGCTTTTG CAGATTGTGG ATTCCAGGTT TTGCTGAGTT	2460
	AAAGAGATAA ACAGCCCTTC GTATAGAAAA ATAAAAACA ACCTTGATG TCCTTGATG	2520
60	CTATTGAGAC TGCCCTAATG TTGTCCCCAG CTATGGGACT CCTAGATGTG ACTGAGAACA	2580
	AAGGTATTGC CAAAGAAGTT CTTACTCAGA GATTGGGACC CTGAAAAGA CCTGTGGCAT	2640
	ACTTGTAAGA AATTAGACCT GGTGGCTGTA AGATGGCCTG CTTGTCTGCA CATAGTGGCT	2700
65	TCTGGTCAAG GACGCAGATA AATTGACTCT GAGACAAAAC TTGGCACATG TCCTAGAAAG	2760

	TGTGGTTCAG	CCCCATGAC	CGATGGCTGA	CTAACGCTCT	TGAAAACATT	ATCCAACGT	2820
	TCCCCTGACC	GATGGACACA	TTGTCAGAGC	TTTTTTTGAC	TGAACGAGTG	ACCTTCGCTC	2880
5	CCCCTGCTAT	CCTCGATCTC	ACTACTGCCT	GAGACTTCAC	CTACTCATCA	TTGTGCTGAC	2940
	ATTCTGGCAG	AAGAACTCA	TACTCGAAAT	GATCTGAAGG	ATCAGATCAG	CCTTGGCCTG	3000
10	AGAGTTTGAG	CTGGTACACG	GATGGCAGTA	GCCTGGAGGT	TAAGGGTAAG	CGGAAGGCGG	3060
	GGACAGCAGT	GCAGTGGTGG	ACAGAAAGCA	AGTGATCTAG	GCCAGCAGCC	TCCCTAAAGG	3120
	GACTTCAGCC	CACAAAGCCA	AACTTGTGGC	TTTAATACAA	GCTCTGTAAA	TGGTAAAAAA	3180
15	AAAAAAGTCT	ACACGGACAG	CAGGTATGCT	CTTGCCACTG	TACAGAGCAA	TATACAGACA	3240
	AAGAGAACTG	TTGACATCTG	CAGAGAAAGA	CCTAAGATGC	TGTGGCTAAA	AGAAATCAGA	3300
20	TGGCAAATCT	AACCGCCCAG	GCATCCTAAA	GAGCAATGAT	CCTGACAGTC	TGAAGACTAT	3360
	CAAGTTATAG	ACAAATTAAG	ACTGGTAAAA	AAAACCCTGT	ATAAAATAGT	AAAAACTGAA	3420
	AAAAGAAAAC	TAGTCCTCTC	ATGAGAAGAC	AGACCTGACA	TCTACTGAAA	AATAGACTTT	3480
25	ACTGGAAAAA	ATATGTGTAT	GAATACCTTC	TAGTTTTTGT	GAACGTCTC	AAGATGGATA	3540
	AAAGCTTTTC	CTTGTAATAAC	GAGACTGATC	AGATAGTCAT	CAAGAAGATT	GTTAAAGAAA	3600
	ATTTTCCAAG	GTTGCGAGTG	CCAAAAGCAA	TAGTGTGAGA	TAATGGTCCT	GCCTTTGTTG	3660
30	CCCAGGTAAG	TCAGGGTGTG	GCCAAGTATT	TAGAGGTCAA	ATGAAAATTC	CATTGTGTGT	3720
	ACAGACCTCA	GAGCTCAGGA	AAGATAAAAA	AGAATAAATA	AAACTCTAAA	CAGACCTTGA	3780
35	CAAAATTAAT	CCTAGAGACT	GGCACAGACT	TACTTGGTAC	TCCTTCCCCT	TGCCCTATTT	3840
	AGAACTGAGA	ATACTCCCTC	TTGATTCGGT	TTTACTCTTT	TTAAGATCCT	TTATGGGGCT	3900
	CCTATGCCAT	CACTGTCTTA	AATGATGTGT	TTAAACCTAT	GTTGTTATAA	TAATGATCTA	3960
40	TATGTTAAGT	TAAAAGGCTT	GCAGGTGGTG	CAGAAAGAAG	TCTGGTCACA	ACTGGCTACA	4020
	GTGAACAAGC	TGGGTACCCC	AAGGACATCT	TACCAGTTCC	AGCCAGAGAT	CTGATCTACG	4080
45	TACACCTGCG	TCATGCTGAG	ACCCTCAAGC	CTCACTAAAA	GGGTCCCTGC	CTAGTTCTGT	4140
	TTACTAATCT	GCCTTATTCT	GTTTTTGTTT	CCATGTTAAA	GATAGAGTAA	ATGCAGTATT	4200
	CTCCACATAG	AGATATAGAC	TTCTGAAATT	CTAAGATTAG	AATTATTTAC	AAGAAGAAGT	4260
50	GGGGAATGAA	GAATAAAAAA	TTACTGGCCT	CTTGTGAGAA	CATGAACTTT	CACCTCGGAG	4320
	CCCACCCCTT	CCCATCTGGA	AAACATACTT	GAGAAAAACA	TTTTCTGGAA	CAACCACAGA	4380
55	ATGTTTCAAC	AGGCCAGATG	TATTGCCAAA	CACAGGATAT	GACTCTTTGG	TTGAGTAAAT	4440
	TTGTGGTTGT	TAAACTTCCC	CTATTCCCTC	CCCATTCCCC	CTCCCAGTTT	GTGGTTTTTT	4500
	CCTTTAAAAG	CTTGTGAAAA	ATTTGAGTCG	TCGTCGAGAC	TCCTCTACCC	TGTGCAAAGG	4560
60	TGTATGAGTT	TCGACCCAG	AGCTCTGTGT	GCTTTCTGTT	GCTGCTTTAT	TTCGACCCCA	4620
	GAGCTCTGGT	CTGTGTGCTT	TCATGTCGCT	GCTTTATTAA	ATCTTACCTT	CTACATTTTA	4680
65	TGTATGGTCT	CAGTGTCTTC	TTGGGTACGC	GGCTGTCCCG	GGACTTGAGT	GTCTGAGTGA	4740

GGGTCTTCCC TCGAGGGTCT TTCATTGGT ACATGGGCCG GGAATTCGAG AATCTTTCAT 4800
TTGGTGCATT GGCCGGGAAT TCGAAAATCT TTCA 4834

5 (2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
10 (A) LENGTH: 4518 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

20 CACCTGACGC GCCCTGTAGC GGCGCATTAA GCGCGGCGGG TGTGGTGGTT ACGCGCAGCG 60
TGACCGCTAC ACTTGCCAGC GCCCTAGCGC CCGCTCCTTT CGCTTCTTTC CCTTCCTTTC 120
TCGCCACGTT CGCCGGCTTT CCCCCTCAAG CTCTAAATCG GGGGCTCCCT TTAGGGTTCC 180
25 GATTTAGTGC TTTACGGCAC CTCGACCCCA AAAAATTGA TTAGGGTGAT GGTTCACGTA 240
GTGGGCCATC GCCCTGATAG ACGGTTTTTC GCCCTTTGAC GTTGGAGTCC ACGTTCTTTA 300
30 ATAGTGGACT CTTGTTCCAA ACTGGAACAA CACTCAACCC TATCTCGGTC TATTCTTTTG 360
ATTTATAAGG GATTTTGCCG ATTTGCGCCT ATTGGTTAAA AAATGAGCTG ATTTAACAAA 420
AATTTAACGC GAATTTTAAC AAAATATTAA CGCTTACAAT TTACGCGTTA AGATACATTG 480
35 ATGAGTTTGG ACAAACCACA ACTAGAATGC AGTGAAAAAA ATGCTTTATT TGTGAAATTT 540
GTGATGCTAT TGCTTTATTT GTAACCATTA TAAGCTGCAA TAAACAAGTT AACAACAACA 600
40 ATTGCATTCA TTTTATGTTT CAGGTTCAGG GGGAGGTGTG GGAGGTTTTT TAAAGCAAGT 660
AAAACCTCTA CAAATGTGGT ATGGCTGATT ATGATCATGA ACAGACTGTG AGGACTGAGG 720
GGCCTGAAAT GAGCCTTGGG ACTGTGAATC TAAATACAC AAACAATTAG AATCAGTAGT 780
45 TTAACACATT ATACACTTAA AAATTGGATC TCCATTGCGC ATTCAGGCTG CGCAACTGTT 840
GGGAAGGGCG ATCGGTGCGG GCCTCTTCGC TATTACGCCA GCTGGCGAAA GGGGGATGTG 900
50 CTGCAAGGCG ATTAAGTTGG GTAACGCCAG GGTTTTCCCA GTCACGACGT TGTAACACGA 960
CGGCCAGTGA ATTGTAATAC GACTCACTAT AGGGCGAATT GGGTACACTT ACCTGGTACC 1020
CCACCCGGGT GGAAAATCGA TGGGCCCCGCG GCCGCTCTAG AAGTACTCTC GAGAAGCTTT 1080
55 TTGAATTCTT TGGATCCACT AGTGTGACCG TGCAGGCGCG CGAGCTCCAG CTTTGTGTTCC 1140
CTTTAGTGAG GGTTAATTTT GAGCTTGGCG TAATCAAGGT CATAGCTGTT TCCTGTGTGA 1200
60 AATTGTTATC CGCTCACAAT TCCACACAAT ATACGAGCCG GAAGTATAAA GTGTAAAGCC 1260
TGGGGTGCCT AATGAGTGAG CTAATCACA GTAATTGCGG CTAGCGGATC TGACGGTTCA 1320
CTAAACCAGC TCTGCTTATA TAGACCTCCC ACCGTACACG CCTACCGCCC ATTTGCGTCA 1380
65 ATGGGGCGGA GTTGTTACGA CATTTTGGAA AGTCCCGTTG ATTTTGGTGC CAAAACAAAC 1440

	TCCCATTGAC	GTCAATGGGG	TGGAGACTTG	GAAATCCCCG	TGAGTCAAAC	CGCTATCCAC	1500
	GCCCATTGAT	GTACTGCCAA	AACCGCATCA	CCATGGTAAT	AGCGATGACT	AATACGTAGA	1560
5	TGTACTGCCA	AGTAGGAAAG	TCCCATAAGG	TCATGTACTG	GGCATAATGC	CAGGCGGGCC	1620
	ATTTACCGTC	ATTGACGTCA	ATAGGGGGCG	TACTTGGCAT	ATGATACACT	TGATGTACTG	1680
10	CCAAGTGGGC	AGTTTACCGT	AAATACTCCA	CCCATTGACG	TCAATGGAAA	GTCCCTATTG	1740
	GCGTTACTAT	GGGAACATAC	GTCATTATTG	ACGTCAATGG	GCGGGGTCG	TTGGGCGGTC	1800
	AGCCAGGCGG	GCCATTTACC	GTAAGTTATG	TAACGCGGAA	CTCCATATAT	GGGCTATGAA	1860
15	CTAATGACCC	CGTAATTGAT	TACTATTAAT	AACTAATGCA	TGGCGGTAAT	ACGGTTATCC	1920
	ACAGAATCAG	GGGATAACGC	AGGAAAGAAC	ATGTGAGCAA	AAGGCCAGCA	AAAGGCCAGG	1980
	AACCGTAAAA	AGGCCGCGTT	GCTGGCGTTT	TTCATAGGC	TCCGCCCCC	TGACGAGCAT	2040
20	CACAAAAATC	GACGCTCAAG	TCAGAGGTGG	CGAAACCCGA	CAGGACTATA	AAGATACCAG	2100
	GCGTTTCCCC	CTGGAAGCTC	CCTCGTGCGC	TCTCCTGTTC	CGACCCTGCC	GCTTACCGGA	2160
25	TACCTGTCCG	CCTTTCTCCC	TTCGGGAAGC	GTGGCGCTTT	CTCATAGCTC	ACGCTGTAGG	2220
	TATCTCAGTT	CGGTGTAGGT	CGTTCGCTCC	AAGCTGGGCT	GTGTGCACGA	ACCCCCGTT	2280
	CAGCCCGACC	GCTGCGCCTT	ATCCGGTAAC	TATCGTCTTG	AGTCCAACCC	GGTAAGACAC	2340
30	GACTTATCGC	CACTGGCAGC	AGCCACTGGT	AACAGGATTA	GCAGAGCGAG	GTATGTAGGC	2400
	GGTGCTACAG	AGTTCTTGAA	GTGGTGGCCT	AACTACGGCT	ACACTAGAAG	GACAGTATTT	2460
35	GGTATCTGCG	CTCTGCTGAA	GCCAGTTACC	TTCGGAAAAA	GAGTTGGTAG	CTCTTGATCC	2520
	GGCAAACAAA	CCACCGCTGG	TAGCGGTGGT	TTTTTTGTTT	GCAAGCAGCA	GATTACGCGC	2580
	AGAAAAAAG	GATCTCAAGA	AGATCCTTTG	ATCTTTTCTA	CGGGGTCTGA	CGCTCAGTGG	2640
40	AACGAAACT	CACGTTAAGG	GATTTTGGTC	ATGAGATTAT	CAAAAAGGAT	CTTCACCTAG	2700
	ATCCTTTTAA	ATTAAAAATG	AAGTTTTAAA	TCAATCTAAA	GTATATATGA	GTAACCTGAG	2760
45	GCTATGGCAG	GGCCTGCCGC	CCCGACGTTG	GCTGCGAGCC	CTGGGCCTTC	ACCCGAACCT	2820
	GGGGGTGGG	GTGGGGAAAA	GGAAGAAACG	CGGGCGTATT	GGCCCCAATG	GGGTCTCGGT	2880
	GGGGTATCGA	CAGAGTGCCA	GCCCTGGGAC	CGAACCCCGC	GTTTATGAAC	AAACGACCCA	2940
50	ACACCGTGCG	TTTTATTCTG	TCTTTTATT	GCCGTCATAG	CGCGGGTTCC	TTCCGGTATT	3000
	GTCTCCTTCC	GTGTTTCAGT	TAGCCTCCCC	CTAGGGTGGG	CGAAGAACTC	CAGCATGAGA	3060
55	TCCCCGCGCT	GGAGGATCAT	CCAGCCGGCG	TCCCGGAAAA	CGATTCCGAA	GCCCAACCTT	3120
	TCATAGAAGG	CGGCGGTGGA	ATCGAAATCT	CGTGATGGCA	GGTTGGGCGT	CGCTTGGTCG	3180
	GTCATTTCTGA	ACCCAGAGT	CCCGCTCAGA	AGAACTCGTC	AAGAAGGCGA	TAGAAGGCGA	3240
60	TGCGCTGCGA	ATCGGGAGCG	GCGATACCGT	AAAGCACGAG	GAAGCGGTCA	GCCCATTCGC	3300
	CGCCAAGCTC	TTCAGCAATA	TCACGGGTAG	CCAACGCTAT	GTCCTGATAG	CGGTCCGCCA	3360
65	CACCCAGCCG	GCCACAGTCG	ATGAATCCAG	AAAAGCGGCC	ATTTTCCACC	ATGATATTCTG	3420

GCAAGCAGGC ATCGCCATGG GTCACGACGA GATCCTCGCC GTCGGGCATG CTCGCCTTGA 3480
GCCTGGCGAA CAGTTCGGCT GGCGCGAGCC CCTGATGCTC TTCGTCCAGA TCATCCTGAT 3540
5 CGACAAGACC GGCTTCCATC CGAGTACGTG CTCGCTCGAT GCGATGTTTC GCTTGGTGGT 3600
CGAATGGGCA GGTAGCCGGA TCAAGCGTAT GCAGCCGCCG CATTGCATCA GCCATGATGG 3660
ATACTTTCTC GGCAGGAGCA AGGTGAGATG ACAGGAGATC CTGCCCCGGC ACTTCGCCCA 3720
10 ATAGCAGCCA GTCCCTTCCC GCTTCAGTGA CAACGTCGAG CACAGCTGCG CAAGGAACGC 3780
CCGTCGTGGC CAGCCACGAT AGCCGCGCTG CCTCGTCTTG CAGTTCATTC AGGGCACC GG 3840
15 ACAGGTCGGT CTTGACAAAA AGAACCGGGC GCCCCTGCGC TGACAGCCGG AACACGGCGG 3900
CATCAGAGCA GCCGATTGTC TGTGTGCCC AGTCATAGCC GAATAGCCTC TCCACCCAAG 3960
CGGCCGGAGA ACCTGCGTGC AATCCATCTT GTTCAATCAT GCGAAACGAT CCTCATCCTG 4020
20 TCTCTTGATC GATCTTTGCA AAAGCCTAGG CCTCCAAAAA AGCCTCCTCA CTACTTCTGG 4080
AATAGCTCAG AGGCCGAGGC GGCCTCGGCC TCTGCATAAA TAAAAAAAT TAGTCAGCCA 4140
TGGGGCGGAG AATGGGCGGA ACTGGGCGGA GTTAGGGGCG GGATGGGCGG AGTTAGGGGC 4200
25 GGGACTATGG TTGCTGACTA ATTGAGATGC ATGCTTTGCA TACTTCTGCC TGCTGGGGAG 4260
CCTGGGGACT TTCCACACCT GGTGCTGAC TAATTGAGAT GCATGCTTTG CATACTTCTG 4320
30 CCTGCTGGGG AGCCTGGGGA CTTTCCACAC CCTAACTGAC ACACATTCCA CAGCTGGTTC 4380
TTTCCGCCTC AGGACTCTTC CTTTTTCAAT ATTATTGAAG CATTATCAG GGTATTGTC 4440
TCATGAGCGG ATACATATTT GAATGTATTT AGAAAAATAA ACAAATAGGG GTTCCGCGCA 4500
35 CATTTCCCCG AAAAGTGC 4518

(2) INFORMATION FOR SEQ ID NO:27:

- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
45 (ii) MOLECULE TYPE: DNA (genomic)

- 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

CTCCACATAG AGATATAGAC TTCTG

25

- 55 (2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
60 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

CGATCTTATT AATTAAGTGG AGTTTTGAGC CCRMCCCCTC CCATC

45

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(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 5594 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

10

(ii) MOLECULE TYPE: DNA (genomic)

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

20	TGCATTAGTT ATTAATAGTA ATCAATTACG GGGTCATTAG TTCATAGCCC ATATATGGAG	60
	TTCCGCGTTA CATAACTTAC GGTAATGGC CCGCCTGGCT GACCGCCCAA CGACCCCGC	120
	CCATTGACGT CAATAATGAC GTATGTTCCC ATAGTAACGC CAATAGGGAC TTTCCATTGA	180
25	CGTCAATGGG TGGAGTATTT ACGGTAACT GCCCACTTGG CAGTACATCA AGTGTATCAT	240
	ATGCCAAGTA CGCCCCCTAT TGACGTCAAT GACGGTAAAT GGCCCGCCTG GCATTATGCC	300
30	CAGTACATGA CCTTATGGGA CTTTCTACT TGGCAGTACA TCTACGTATT AGTCATCGCT	360
	ATTACCATGG TGATGCGGTT TTGGCAGTAC ATCAATGGGC GTGGATAGCG GTTTGACTCA	420
	CGGGGATTTC CAAGTCTCCA CCCCATGAC GTCAATGGGA GTTTGTTTGT GCACCAAAAT	480
35	CAACGGGACT TTCCAAAATG TCGTAACAAC TCCGCCCAT TGACGCAAAT GGGCGGTAGG	540
	CGTGTACGGT GGGAGGTCTA TATAAGCAGA GCTGGTTTAG TGAACCGTCA GATCCGCGCC	600
40	AGTCTCCGA TTGACTGAGT CGCCCGGTA CCCGTGTATC CAATAAACCC TCTTGCACTT	660
	GCATCCGACT TGTGGTCTCG CTGTTCTTGG GGAGGGTCTC CTCTGAGTGA TTGACTACCC	720
	GTCAGCGGGG GTCTTTCATT TGGGGGCTCG TCCGGGATCG GGAGACCCCT GCCCAGGGAC	780
45	CACCGACCCA CCACCGGGAG GTAAGCTGGC CAGCAACTTA TCTGTGTCTG TCCGATTGTC	840
	TAGTGTCTAT GACTGATTTT ATGCGCCTGC GTCGGTACTA GTTAGCTAAC TAGCTCTGTA	900
50	TCTGGCGGAC CCGTGGTGA ACTGACGAGT TCGGAACACC CGGCCGCAAC CCTGGGAGAC	960
	GTCCCAGGAG GAACAGGGGA GGATCAGGGA CGCCTGGTGG ACCCCTTTGA AGGCCAAGAG	1020
	ACCATTTGGG GTTGCAGAT CGTGGGTTCT AGTCCCACCT CGTGCCAGT TGCAGATCG	1080
55	TGGGTTTCGAG TCCCACCTCG TGTTTTGTTG CGAGATCGTG GGTTTCGAGT CCACCTCGCG	1140
	TCTGGTCACG GGATCGTGGG TTCGAGTCCC ACCTCGTGTT TTGTTGCGAG ATCGTGGGTT	1200
60	CGAGTCCCAC CTCGCGTCTG GTCACGGGAT CGTGGGTTCT AGTCCCACCT CGTGCAGAGG	1260
	GTCTCAATTG GCCGGCCTTA GAGAGGCCAT CTGATTCTTC TGGTTTCTCT TTTTGTCTTA	1320
	GTCTCGTGTC CGCTCTTGTT GTGACTACTG TTTTCTAAA AATGGGACAA TCTGTGTCCA	1380
65	CTCCCCTTTC TCTGACTCTG GTTCTGTCTG TTGGTAATTT TGTTTGTTTA CGTTTGTTTT	1440

	TGTGAGTCGT CTATGTTGTC TGTTACTATC TTGTTTTTGT TTGTGGTTTA CGGTTTCTGT	1500
	GTGTGTCCTTG TGTGTCTCTT TGTGTTTCTG CTTGGACTGA TGAAGTACGA CTGTTTTTAA	1560
5	GTTATGCCTT CTAAATAAG CCTAAAAATC CTGTCAGATC CCTATGCTGA CCACTTCCTT	1620
	TCAGATCAAC AGCTGCCCTT ACGTATCGAT GGATCCCTCG ACTAACTAAT AGCCCATCTT	1680
10	CCAAGGTCGA GCGGGATCAA TTCCGCCCCC CCCCTAACGT TACTGGCCGA AGCCGCTTGG	1740
	AATAAGGCCG GTGTGCGTTT GTCTATATGT TATTTTCCAC CATATTGCCG TCTTTTGGCA	1800
	ATGTGAGGGC CCGGAAACCT GGCCCTGTCT TCTTGACGAG CATTCTAGG GGTCTTTCCC	1860
15	CTCTCGCCAA AGGAATGCAA GGTCTGTTGA ATGTCGTGAA GGAAGCAGTT CCTCTGGAAG	1920
	CTTCTTGAAG ACAAACAACG TCTGTAGCGA CCCTTTGCAG GCAGCGGAAC CCCCCACCTG	1980
20	GCGACAGGTG CCTCTGCGGC CAAAAGCCAC GTGTATAAGA TACACCTGCA AAGGCGGCAC	2040
	AACCCAGTG CCACGTTGTG AGTTGGATAG TTGTGAAAG AGTCAAATGG CTCTCCTCAA	2100
	GCGTATTCAA CAAGGGGCTG AAGGATGCCC AGAAGGTACC CCATTGTATG GGATCTGATC	2160
25	TGGGGCCTCG GTGCACATGC TTTACATGTG TTTAGTCGAG GTTAAAAAAA CGTCTAGGCC	2220
	CCCCGAACCA CGGGGACGTG GTTTTCCTTT GAAAAACACG ATAATAATCA TGGCTACAGG	2280
30	CTCCCGGACG TCCCTGCTCC TGGCTTTTGG CCTGCTCTGC CTGCCCTGGC TTCAAGAGGG	2340
	CAGTGCCTTC CCAACCATTG CCTTATCCAG GCTTTTTGAC AACGCTATGC TCCGCGCCCA	2400
	TCGTCTGCAC CAGCTGGCCT TTGACACCTA CCAGGAGTTT GAAGAAGCCT ATATCCCAAA	2460
35	GGAACAGAAG TATTCATTCC TGCAGAACCC CCAGACCTCC CTCTGTTTCT CAGAGTCTAT	2520
	TCCGACACCC TCCAACAGGG AGGAAACACA ACAGAAATCC AACCTAGAGC TGCTCCGCAT	2580
40	CTCCCTGCTG CTATCCAGT CGTGGCTGGA GCGCGTGAG TTCCTCAGGA GTGTCTTCGC	2640
	CAACAGCCTG GTGTACGGCG CCTCTGACAG CAACGTCTAT GACCTCCTAA AGGACCTAGA	2700
	GGAAGGCATC CAAACGCTGA TGGGGAGGCT GGAAGATGGC AGCCCCCGGA CTGGGCAGAT	2760
45	CTTCAAGCAG ACCTACAGCA AGTCGACAC AAACCTACAC AACGATGACG CACTACTCAA	2820
	GAACTACGGG CTGCTCTACT GCTTCAGGAA GGACATGGAC AAGGTCGAGA CATTCTGCG	2880
50	CATCGTGCAG TGCCGCTCTG TGGAGGGCAG CTGTGGCTTC TAGCTGCCCG GGTGGCATCC	2940
	TGTGACCCCT CCCCAGTGCC TCTCCTGGCC CTGGAAGTTG CCACTCCAGT GCCCACCAGC	3000
	CTTGTCTTAA TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA	3060
55	GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC	3120
	CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC	3180
60	TAACCTCGCC CATCCCGCCC CTAACCTCCG CCAGTTCCGC CCATTCTCCG CCCCATGGCT	3240
	GAATAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCCT GGCCTCTGAG CTATTCCAGA	3300
	AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTTCAC GCTGCCGCAA	3360
65	GCACTCAGGG CGCAAGGGCT GCTAAAGGAA GCGGAACACG TAGAAAGCCA GTCCGCAGAA	3420

	ACGGTGCTGA	CCCCGGATGA	ATGTCAGCTA	CTGGGCTATC	TGGACAAGGG	AAAACGCAAG	3480
	CGCAAAGAGA	AAGCAGGTAG	CTTGCACTGG	GCTTACATGG	CGATAGCTAG	ACTGGGCGGT	3540
5	TTTATGGACA	GCAAGCGAAC	CGGAATTGCC	AGCTGGGGCG	CCCTCTGGTA	AGGTTGGGAA	3600
	GCCCTGCAAA	GTAAACTGGA	TGGCTTTCTT	GCCGCCAAGG	ATCTGATGGC	GCAGGGGATC	3660
10	AAGATCTGAT	CAAGAGACAG	GATGAGGATC	GTTTCGCATG	ATTGAACAAG	ATGGATTGCA	3720
	CGCAGGTTCT	CCGGCCGCTT	GGGTGGAGAG	GCTATTTCGGC	TATGACTGGG	CACAACAGAC	3780
	AATCGGCTGC	TCTGATGCCG	CCGTGTTCCG	GCTGTCAGCG	CAGGGGCGCC	CGGTTCTTTT	3840
15	TGTCAAGACC	GACCTGTCCG	GTGCCCTGAA	TGAACTGCAG	GACGAGGCAG	CGCGGCTATC	3900
	GTGGCTGGCC	ACGACGGGCG	TTCCTTGCGC	AGCTGTGCTC	GACGTTGTCA	CTGAAGCGGG	3960
20	AAGGGACTGG	CTGCTATTGG	GCGAAGTGCC	GGGGCAGGAT	CTCCTGTCAT	CTCACCTTGC	4020
	TCCTGCCGAG	AAAGTATCCA	TCATGGCTGA	TGCAATGCGG	CGGCTGCATA	CGCTTGATCC	4080
	GGCTACCTGC	CCATTGACCC	ACCAAGCGAA	ACATCGCATC	GAGCGAGCAC	GTAATCGGAT	4140
25	GGAAGCCGGT	CTTGTGATC	AGGATGATCT	GGACGAAGAG	CATCAGGGGC	TCGCGCCAGC	4200
	CGAACTGTTT	GCCAGGCTCA	AGGCGCGCAT	GCCCGACGGC	GAGGATCTCG	TCGTGACCCA	4260
	TGGCGATGCC	TGCTTGCCGA	ATATCATGGT	GGAAAATGGC	CGCTTTTCTG	GATTCATCGA	4320
30	CTGTGGCCGG	CTGGGTGTGG	CGGACCGCTA	TCAGGACATA	GCGTTGGCTA	CCCGTGATAT	4380
	TGCTGAAGAG	CTTGGCGGCG	AATGGGCTGA	CCGCTTCCTC	GTGCTTTACG	GTATCGCCGC	4440
35	TCCCATTTCG	CAGCGCATCG	CCTTCTATCG	CCTTCTTGAC	GAGTTCTTCT	GAGCGGGACT	4500
	CTGGGGTTCG	AAATGACCGA	CCAAGCGACG	CCCAACCTCC	AGAAAAAGGG	GGGAATGAAA	4560
40	GACCCACCT	GTAGGTTTGG	CAAGCTAGCT	TAAGTAACGC	CATTTTGCAA	GGCATGGAAA	4620
	AATACATAAC	TGAGAATAGA	GAAGTTCAGA	TCAAGGTCAG	GAACAGATGG	AACAGCTGAA	4680
	TATGGGCCAA	ACAGGATATC	TGTGGTAAGC	AGTTCCTGCC	CCGGCTCAGG	GCCAAGAACA	4740
45	GATGGAACAG	CTGAATATGG	GCCAAACAGG	ATATCTGTGG	TAAGCAGTTC	CTGCCCCGGC	4800
	TCAGGGCCAA	GAACAGATGG	TCCCCAGATG	CGGTCCAGCC	CTCAGCAGTT	TCTAGAGAAC	4860
	CATCAGATGT	TTCCAGGGTG	CCCCAAGGAC	CTGAAATGAC	CCTGTGCCTT	ATTGAACTA	4920
50	ACCAATCAGT	TCGCTTCTCG	CTTCTGTTCG	CGCGCTTCTG	CTCCCCGAGC	TCAATAAAAG	4980
	AGCCCACAAC	CCCTCACTCG	GGGCGCCAGT	AATCTGCTGC	TTGCAAACAA	AAAAACCACC	5040
55	GCTACCAGCG	GTGGTTTGTT	TGCCGGATCA	AGAGCTACCA	ACTCTTTTTC	CGAAGGTAAC	5100
	TGGCTTCAGC	AGAGCGCAGA	TACCAAATAC	TGTCCTTCTA	GTGTAGCCGT	AGTTAGGCCA	5160
60	CCACTTCAAG	AACTCTGTAG	CACCGCCTAC	ATACCTCGCT	CTGCTAATCC	TGTTACCAGT	5220
	GGCTGCTGCC	AGTGGCGATA	AGTCGTGTCT	TACCGGGTTG	GACTCAAGAC	GATAGTTACC	5280
	GGATAAGGCG	CAGCGGTCCG	GCTGAACGGG	GGGTTTCGTG	ACACAGCCCA	GCTTGGAGCG	5340
65	AACGACCTAC	ACCGAACTGA	GATACCTACA	GCGTGAGCAT	TGAGAAAGCG	CCACGCTTCC	5400

CGAAGGGAGA AAGGCGGACA GGTATCCGGT AAGCGGCAGG GTCGGAACAG GAGAGCGCAC 5460
GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA TCTTTATAGT CCTGTCGGGT TTCGCCACCT 5520
5 CTGACTTGAG CGTCGATTTT TGTGATGCTC GTCAGGGGGG CGGAGCCTAT GGAAAAACGC 5580
CAGCAACGCC GAGA 5594

(2) INFORMATION FOR SEQ ID NO:30:

10 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6561 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
15 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

25 GATCCCCGGG TCGACCCGGG TCGACCCTGT GGAATGTGTG TCAGTTAGGG TGTGGAAAGT 60
CCCCAGGCTC CCCAGCAGGC AGAAGTATGC AAAGCATGCA TCTCAATTAG TCAGCAACCA 120
GGTGTGAAAA GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT GCAAAGCATG CATCTCAATT 180
30 AGTCAGCAAC CATAGTCCCG CCCCTAACTC CGCCCATCCC GCCCCTAACT CCGCCCAGTT 240
CCGCCCATT CCGCCCCAT GGCTGACTAA TTTTTTTTAT TTATGCAGAG GCCGAGGCCG 300
CCTCGGCCTC TGAGCTATT CAGAAGTAGT GAGGAGGCTT TTTTGAGGC CTAGGCTTTT 360
35 GCAAAAAGCT TCACGCTGCC GCAAGCACTC AGGGCGCAAG GGCTGCTAAA GGAAGCGGAA 420
CACGTAGAAA GCCAGTCCGC AGAAACGGTG CTGACCCCGG ATGAATGTCA GCTACTGGGC 480
40 TATCTGGACA AGGGAAAACG CAAGCGCAAA GAGAAAGCAG GTAGCTTGCA GTGGGCTTAC 540
ATGGCGATAG CTAGACTGGG CGGTTTTATG GACAGCAAGC GAACCGGAAT TGCCAGCTGG 600
GGCGCCCTCT GGTAAGGTTG GGAAGCCCTG CAAAGTAAAC TGGATGGCTT TCTTGCCGCC 660
45 AAGGATCTGA TGGCGCAGGG GATCAAGATC TGATCAAGAG ACAGGATGAG GATCGTTTCG 720
CATGATTGAA CAAGATGGAT TGCACGCAGG TTCTCCGGCC GCTTGGGTGG AGAGGCTATT 780
50 CGGCTATGAC TGGGCACAAC AGACAATCGG CTGCTCTGAT GCCGCCGTGT TCCGGCTGTC 840
AGCGCAGGGG CGCCCGGTTT TTTTGTCAA GACCGACCTG TCCGGTGCCC TGAATGAACT 900
GCAGGACGAG GCAGCGCGGC TATCGTGGCT GGCCACGACG GCGTTCCTT GCGCAGCTGT 960
55 GCTCGACGTT GTCATGAAG CGGGAAGGGA CTGGCTGCTA TTGGGCGAAG TGCCGGGGCA 1020
GGATCTCCTG TCATCTCACC TTGCTCCTGC CGAGAAAGTA TCCATCATGG CTGATGCAAT 1080
60 GCGGCGGCTG CATACTGCTG ATCCGGCTAC CTGCCCATTC GACCACCAAG CGAAACATCG 1140
CATCGAGCGA GCACGTACTC GGATGGAAGC CGGTCTTGTC GATCAGGATG ATCTGGACGA 1200
AGAGCATCAG GGGCTCGCGC CAGCCGAACT GTTCGCCAGG CTCAAGGCGC GCATGCCCGA 1260
65 CGGCGAGGAT CTCGTCGTGA CCCATGGCGA TGCTTGCTTG CCGAATATCA TGGTGAAAA 1320

	TGGCCGCTTT	TCTGGATTCA	TCGACTGTGG	CCGGCTGGGT	GTGGCGGACC	GCTATCAGGA	1380
	CATAGCGTTG	GCTACCCGTG	ATATTGCTGA	AGAGCTTGGC	GGCGAATGGG	CTGACCGCTT	1440
5	CCTCGTGCTT	TACGGTATCG	CCGCTCCCGA	TTCGCAGCGC	ATCGCCTTCT	ATCGCCTTCT	1500
	TGACGAGTTC	TTCTGAGCGG	GACTCTGGGG	TTCGAAATGA	CCGACCAAGC	GACGCCCAAC	1560
10	CTGCCATCAC	GAGATTTTCGA	TTCCACCGCC	GCCTTCTATG	AAAGGTTGGG	CTTCGGAATC	1620
	GTTTTCCGGG	ACGCCGGCTG	GATGATCCTC	CAGCGCGGGG	ATCTCATGCT	GGAGTTCTTC	1680
	GCCCAACCCG	GAATTCGTAA	TCTGCTGCTT	GCAAACAAAA	AAACCACCGC	TACCAGCGGT	1740
15	GGTTTGTTTG	CCGGATCAAG	AGCTACCAAC	TCTTTTTCCG	AAGGTAAGTG	GCTTCAGCAG	1800
	AGCGCAGATA	CCAAATACTG	TCCTTCTAGT	GTAGCCGTAG	TTAGGCCACC	ACTTCAAGAA	1860
20	CTCTGTAGCA	CCGCCTACAT	ACCTCGCTCT	GCTAATCCTG	TTACCAGTGG	CTGCTGCCAG	1920
	TGGCGATAAG	TCGTGTCTTA	CCGGGTTGGA	CTCAAGACGA	TAGTTACCGG	ATAAGGCGCA	1980
	GCGGTCGGGC	TGAACGGGGG	GTTCTGTCAC	ACAGCCCAGC	TTGGAGCGAA	CGACCTACAC	2040
25	CGAACTGAGA	TACCTACAGC	GTGAGCATTG	AGAAAGCGCC	ACGCTTCCCG	AAGGGAGAAA	2100
	GGCGGACAGG	TATCCGGTAA	GCGGCAGGGT	CGGAACAGGA	GAGCGCACGA	GGGAGCTTCC	2160
30	AGGGGGAAAC	GCCTGGTATC	TTTATAGTCC	TGTCGGGTTT	CGCCACCTCT	GACTTGAGCG	2220
	TCGATTTTTG	TGATGCTCGT	CAGGGGGGCG	GAGCCTATGG	AAAAACGCCA	GCAACGCCGA	2280
	GATGCGCCGC	CTCGAGTACA	CCTGCGTCAT	GCTGAGACCC	TCAAGCCTCA	CTAAAAGGGT	2340
35	CCCTGCTAG	TTCTGTTTAC	TAATCTGCCT	TATTCTGTTT	TTGTTCCCAT	GTAAAGATA	2400
	GAGTAAATGC	AGTATTCTCC	ACATAGAGAT	ATAGACTTCT	GAAATTCTAA	GATTAGAATT	2460
40	ATTTACAAGA	AGAAGTGGGG	AATGAAGAAT	AAAAAATTAC	TGGCCTCTTG	TGAGAACATG	2520
	AACTTTCACC	TCGGAGCCCA	CCCCCTCCCA	TCTGGAAAAC	ATACTTGAGA	AAAACATTTT	2580
	CTGGAACAAC	CACAGAATGT	TTCAACAGGC	CAGATGTATT	GCCAAACACA	GGATATGACT	2640
45	CTTTGGTTGA	GTAAATTTGT	GGTTGTTAAA	CTTCCCCTAT	TCCCTCCCA	TTCCCCTCC	2700
	CAGTTTGTGG	TTTTTTCCTT	TAAAAGCTTG	TGAAAAATTT	GAGTCGTCGT	CGAGACTCCT	2760
50	CTACCTGTG	CAAAGGTGTA	TGAGTTTCGA	CCCCAGAGCT	CTGTGTGCTT	TCTGTTGCTG	2820
	CTTTATTTTCG	ACCCAGAGC	TCTGGTCTGT	GTGCTTTCAT	GTCGCTGCTT	TATTAAATCT	2880
	TACCTTCTAC	ATTTTATGTA	TGGTCTCAGT	GTCTTCTTGG	GTACGCGGCT	GTCCCGGGAC	2940
55	TTGAGTGTCT	GAGTGAGGGT	CTTCCCTCGA	GGGTCTTTCA	TTTGGTACAT	GGGCCGGGAA	3000
	TTGAGAATC	TTTCATTTGG	TGCATTGGCC	GGGAATTCGA	AAATCTTTCA	TTTGGTGCAT	3060
60	TGGCCGGGAA	ACAGCGCGAC	CACCCAGAGG	TCCTAGACCC	ACTTAGAGGT	AAGATTCTTT	3120
	GTTCTGTTTT	GGTCTGATGT	CTGTGTTCTG	ATGTCTGTGT	TCTGTTTCTA	AGTCTGGTGC	3180
	GATCGCAGTT	TCAGTTTTGC	GGACGCTCAG	TGAGACCGCG	CTCCGAGAGG	GAGTGCGGGG	3240
65	TGGATAAGGA	TAGACGTGTC	CAGGTGTCCA	CCGTCCGTTC	GCCCTGGGAG	ACGTCCCAGG	3300

	AGGAACAGGG	GAGGATCAGG	GACGCCTGGT	GGACCCCTTT	GAAGGCCAAG	AGACCATTTC	3360
	GGGTTGCGAG	ATCGTGGGTT	CGAGTCCCAC	CTCGTGCCCA	GTTGCGAGAT	CGTGGGTTTC	3420
5	AGTCCCACCT	CGTGTTTTGT	TGCGAGATCG	TGGGTTTCGAG	TCCCACCTCG	CGTCTGGTCA	3480
	CGGGATCGTG	GGTTCGAGTC	CCACCTCGTG	TTTGTTCGCG	AGATCGTGGG	TTCGAGTCCC	3540
10	ACCTCGCGTC	TGGTCACGGG	ATCGTGGGTT	CGAGTCCCAC	CTCGTGCAGA	GGGTCTCAAT	3600
	TGGCCGGCCT	TAGAGAGGCC	ATCTGATTCT	TCTGGTTTCT	CTTTTGTCT	TAGTCTCGTG	3660
	TCCGCTCTTG	TTGTGACTAC	TGTTTTTCTA	AAAATGGGAC	AATCTGTGTC	CACTCCCCTT	3720
15	TCTCTGACTC	TGGTTCTGTC	GCTTGGAAT	TTTGTTCGTT	TACGTTTGT	TTTGTGAGTC	3780
	GTCTATGTTG	TCTGTACTA	TCTTGTTCCT	GTTGTGGTT	TACGGTTTCT	GTGTGTGTCT	3840
	TGTGTGTCTC	TTTGTGTTCA	GACTTGGACT	GATGACTGAC	GACTGTTTTT	AAGTTATGCC	3900
20	TTCTAAAATA	AGCCTAAAAA	TCCTGTCAGA	TCCCTATGCT	GACCACTTCC	TTTCAGATCA	3960
	ACAGTGCCCC	TGCCTCCCAC	TCCAACCTCA	GAGAGCAGCC	AGCGGGTCAC	AGTGGTCCCC	4020
25	CCCATGAACC	TGGAGCCTAG	GGAAAAATGA	GCTCGGAAAT	CCGGAGCAAA	TGAGGAGTGG	4080
	TCCCTGAGAA	GTCAGTGGCC	TAAATGTTGT	GGCTGCTGAA	GCAAAAGAAG	AGGAGGCTGT	4140
	TCGAGTAGCC	GGCCAAGAGC	GCCGCGGGTT	CCCAGGCAGC	TTCTCATTCC	CCTGTCCCTC	4200
30	CCATCCCGTC	TCTTGTTAAC	AGAAAACTG	CTTCACTTT	GAGATATGAG	TGGCCCGATA	4260
	CAGCCAGCTG	TGAGAGCTGT	ACTCCCTTCC	CTGCCCCACG	TGTTTTCTCT	TCTCAGGCGA	4320
35	CCCCCTCCCTG	AGCTGCTGGC	AGTGAGCTG	TTCTAAGCTC	CAGTGAGGGA	GGCATCCGCC	4380
	CACTTGGGGC	TTCTGTCCAA	GGTAAGGAGC	ACCTGTGAGT	CTAACTGCCA	GGCTCTGATG	4440
	GGGGTCTCGT	CTCTGTGGGA	CTAGAAAGTG	TCCCAACAAT	CTGACCAAGG	TAACAGGAAG	4500
40	TTAAGACAAA	GACAGAGACC	AAAGTCAGAA	TCAGAGCTGT	GCTGTGAGAC	AAAAAGATAA	4560
	AAAAAATAAA	ATGCTGGCCA	CAAAGTCAG	GAAACTAGA	AACTTAGAT	AGTACCTGGC	4620
45	AACAAAAGAA	AGCTTTTGGC	TAAAGATCAA	CGTGATACT	GTAAGAAAAA	TGAGCACTGG	4680
	GTGAGAGACT	GCCCCAACAA	AAAGAAGAGG	AGCCCCCTC	ATGACCAAAC	CCTTCACCTG	4740
	TTCGTGGCTA	AAAGTAAAGA	GATAACAAAA	GGGGTGCTAA	CACAGAAGCT	GAGTCCTTAA	4800
50	AAGAGTCCGG	TGGCCTACCT	GTTGAAGCAG	CTAAAAAAGA	GACTGTGTTT	CATACTCCTC	4860
	CACTGACCAG	TGCAAAACAA	GCTAAAAAGT	TCCTGGGCAC	TGCGGGCTTT	TGCAGATTGT	4920
55	GGATTCCAGG	TTTTGTGAG	TTAAAGAGAT	AAACAGCCCT	TCGTATAGAA	AAATAAAAAA	4980
	CAACCTTGGA	TGTCCTTGGA	TGCTATTGAG	ACTGCCCTAA	TGTTGTCCCC	AGCTATGGGA	5040
	CTCCTAGATG	TGACTGAGAA	CAAAGGTATT	GCCAAAGAAG	TTCTTACTCA	GAGATTGGGA	5100
60	CCCTGAAAAA	GACCTGTGGC	ATACTTGTA	GAAATTAGAC	CTGGTGGCTG	TAAGATGGCC	5160
	TGCTTGCTCG	CACATAGTGG	CTTCTGGTCA	AGGACGCAGA	TAAATTGACT	CTGAGACAAA	5220
65	ACTTGGCACA	TGTCCTAGAA	AGTGTGGTTC	AGCCCCATG	ACCGATGGCT	GAATAACGCT	5280
	CTTGAAAACA	TTATCCAAC	GTTCCCCTGA	CCGATGGACA	CATTGTCAGA	GCTTTTTTTG	5340

ACTGAACGAG TGACCTTCGC TCCCCCTGCT ATCCTCGATC TCACTACTGC CTGAGACTTC 5400
 ACCTACTCAT CATTGTGCTG ACATTCTGGC AGAAGAACT CATACTCGAA ATGATCTGAA 5460
 5 GGATCAGATC AGCCTTGGCC TGAGAGTTTG AGCTGGTACA CGGATGGCAG TAGCCTGGAG 5520
 GTTAAGGGTA AGCGGAAGGC GGGGACAGCA GTGCAGTGGT GGACAGAAAG CAAGTGATCT 5580
 10 AGGCCAGCAG CCTCCCTAAA GGGACTTCAG CCCACAAAGC CAAACTTGTG GCTTTAATAC 5640
 AAGCTCTGTA AATGGTAAAA AAAAAAAGT CTACACGGAC AGCAGGTATG CTCTTGCCAC 5700
 TGTACAGAGC AATATACAGA CAAAGAGAAC TGTTGACATC TGCAGAGAAA GACCTAAGAT 5760
 15 GCTGTGGCTA AAAGAAATCA GATGGCAAAT CTAACCGCCC AGGCATCCTA AAGAGCAATG 5820
 ATCCTGACAG TCTGAAGACT ATCAAGTTAT AGACAAATTA AGACTGGTAA AAAAAACCTT 5880
 GTATAAAATA GTAAAACTG AAAAAAGAAA ACTAGTCCTC TCATGAGAAG ACAGACCTGA 5940
 20 CATCTACTGA AAAATAGACT TTAAGGAAA AAATATGTGT ATGAATACCT TCTAGTTTTT 6000
 GTGAACGTTT TCAAGATGGA TAAAAGCTTT TCCTTGTAAG ACGAGACTGA TCAGATAGTC 6060
 25 ATCAAGAAGA TTGTAAAGA AAATTTTCCA AGGTTCCGAG TGCCAAAAGC AATAGTGTC 6120
 GATAATGGTC CTGCCTTTGT TGCCAGGTA AGTCAGGGTG TGGCCAAGTA TTTAGAGGTC 6180
 AAATGAAAAT TCCATTGTGT GTACAGACCT CAGAGCTCAG GAAAGATAAA AAAGAATAAA 6240
 30 TAAAACTCTA AACAGACCTT GACAAAATTA ATCCTAGAGA CTGGCACAGA CTTACTTGGT 6300
 ACTCCTTCCC CTGCCCCTAT TTAGAACTGA GAATACTCCC TCTTGATTCG GTTTTACTCT 6360
 35 TTTTAAGATC CTTTATGGGG CTCCTATGCC ATCACTGTCT TAAATGATGT GTTTAAACCT 6420
 ATGTTGTTAT AATAATGATC TATATGTTAA GTTAAAAGGC TTGCAGGTGG TGCAGAAAGA 6480
 AGTCTGGTCA CAACTGGCTA CAGTGAACAA GCTGGGTACC CCAAGGACAT CTTACCAGTT 6540
 40 CCAGCCAGAG ATCTGATCTA C 6561

(2) INFORMATION FOR SEQ ID NO:31:

45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 55 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 50 (ii) MOLECULE TYPE: DNA (genomic)

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

GACTAACCTT GATTCCACTG GAGCCGTATT ACCGCCATGC ATTAGTTATT AATAG

55

60 (2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 47 base pairs
 (B) TYPE: nucleic acid
 65 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
GACTAACCTT GATTCCACTG GAGTAATTGC GGCTAGCGGA TCTGACG

47

10 (2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 66 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15

(ii) MOLECULE TYPE: DNA (genomic)

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

25 GACTAACCTT GATTCCACTG GAGACACTTG ACCTCTACCG CGCCAGTCCT CCGATTGACT
GAGTCG

60

66

30 (2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

35

(ii) MOLECULE TYPE: DNA (genomic)

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

45 GACTAACCTT GATTCCACTG GAGGGATCCG CGCCCATGAT TATTATCG

48

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 55 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50

(ii) MOLECULE TYPE: DNA (genomic)

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

60 GACTAACCTT GATTCCAGCA ATGTCATGGC TACAGGCTCC CGGACGTCCC TGCTC

55

(2) INFORMATION FOR SEQ ID NO:36:

65 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

10 GACTAACCTT GATTCCAGCA ATGTTAGGAC AAGGCTGGTG GGCCTGG 48

(2) INFORMATION FOR SEQ ID NO:37:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 49 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
20 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GACTAACCTT GATTCCACTG GAGGGTCGAC CCTGTGGAAT GTGTGTCAG 49

30 (2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 48 base pairs
(B) TYPE: nucleic acid
35 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

45 GACTAACCTT GATTCCACTG GAGAATCTCG TGATGGCAGG TTGGGCGT 48

(2) INFORMATION FOR SEQ ID NO:39:

50 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 54 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

55 (ii) MOLECULE TYPE: DNA (genomic)

60 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

GACTAACCTT GATTCCACTG AAGAGATTTT ATTTAGTCTC CAGAAAAAGG GGGG 54

(2) INFORMATION FOR SEQ ID NO:40:

65 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 50 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: DNA (genomic)

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:
GACTAACCTT GATTCCACTG AAGCCCCCAA ATGAAAGACC CCCGCTGACG 50

15 (2) INFORMATION FOR SEQ ID NO:41:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 49 base pairs
(B) TYPE: nucleic acid
20 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:
30 GACTAACCTT GATTCCACTG GAGCCGGGAC GGAATTCGTA ATCTGCTGC 49

(2) INFORMATION FOR SEQ ID NO:42:
(i) SEQUENCE CHARACTERISTICS:
35 (A) LENGTH: 47 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
40 (ii) MOLECULE TYPE: DNA (genomic)

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:
GACTAACCTT GATTCCACTG GAGTTCTCGA GCGGCGCAT CTCGGCG 47

(2) INFORMATION FOR SEQ ID NO:43:
50 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
55 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

60 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
65 CGCTCTAGAA CTAGTGGATC 20

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
GTAATACGAC TCACTATAGG G

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
CGATCCACTG GAGCTCGGAG CCCACCCCT CCCATCTAGA GGT

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:
CGTCCTCCTG GAGAGCACAG GGTAGAGGAG TCTCGACGGT CAG

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:
CGCAACCCTG GAGACCTCTA GATGGGAGGG GGTGGGCTCC GAG

(2) INFORMATION FOR SEQ ID NO:48:

5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

GCAGGACCTG GAGCTGACCG TCGAGACTCC TCTACCCTGT GCT 43

20 (2) INFORMATION FOR SEQ ID NO:49:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
25 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

35 CGCTCTAGAA CTAGTGGATC 20

(2) INFORMATION FOR SEQ ID NO:50:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
40 (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: DNA (genomic)

50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

GTAATACGAC TCACTATAGG G 21

(2) INFORMATION FOR SEQ ID NO:51:

55 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
60 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

TACGTATCGA TGGATCCGA

19

(2) INFORMATION FOR SEQ ID NO:52:

- 5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: DNA (genomic)

- 15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

GGATCCATCG ATACGTAAG

19

- 20 (2) INFORMATION FOR SEQ ID NO:53:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 38 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 25 (ii) MOLECULE TYPE: DNA (genomic)
- 30

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

35 GGCCGCTAAC TAATAGCCCA TTCTCCAAGG TACGTAGC

38

(2) INFORMATION FOR SEQ ID NO:54:

- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 38 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 45 (ii) MOLECULE TYPE: DNA (genomic)

- 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

TACGTACCTT GGAGAATGGG CTATTAGTTA GCGGCCGC

38

(2) INFORMATION FOR SEQ ID NO:55:

- 55 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 55 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 60 (ii) MOLECULE TYPE: DNA (genomic)
- 65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:
GACTAACCTT GATTCCACTG GAGTTTCTC TATTCTTCAT TCCCCACTTC TTCTT 55

(2) INFORMATION FOR SEQ ID NO:56:

5

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 60 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10

(ii) MOLECULE TYPE: DNA (genomic)

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

GACTAACCTT GATTCCACTG GAGAATCTGG ACCAATTCTA TATAAGCCTG TGAAAAATTT 60

20

(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 46 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

GACTAACCTT GATTCCACTG GAGAAGAAGA AGTGGGGAAT GAAGAA 46

35

(2) INFORMATION FOR SEQ ID NO:58:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 51 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: DNA (genomic)

45

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

GACTAACCTT GATTCCACTG GAGATCTCTA GATGGGAGGG GGTCTGGGCT C 51

55

(2) INFORMATION FOR SEQ ID NO:59:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 47 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

60

(ii) MOLECULE TYPE: DNA (genomic)

65

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:
5 GACTAACCTT GATTCCACTG GAGCTCGGAG CCCACCCCT CCCATCT 47
(2) INFORMATION FOR SEQ ID NO:60:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 47 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: DNA (genomic)
15

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:
20 GACTAACCTT GATTCCACTG GAGGGAGGCC CTTATCTCAA AAATGTT 47
(2) INFORMATION FOR SEQ ID NO:61:
25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 51 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
30 (ii) MOLECULE TYPE: DNA (genomic)

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:
GACTAACCTT GATTCCACTG GAGTCTAAGA ACATTTTGA GATAAGGGCC T 51
40 (2) INFORMATION FOR SEQ ID NO:62:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 44 base pairs
 (B) TYPE: nucleic acid
45 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: DNA (genomic)
50

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:
55 GACTAACCTT GATTCCACTG GAGTCACAGG CTTATATAGT GAAA 44
(2) INFORMATION FOR SEQ ID NO:63:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 46 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
60
65 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

5 GACTAACCTT GATTCCCTGG AGACTGCACT GCTGTCCCCG CCTTCG 46

(2) INFORMATION FOR SEQ ID NO:64:

(i) SEQUENCE CHARACTERISTICS:

- 10 (A) LENGTH: 53 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA (genomic)

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

GAGTAACCTT GATTCCCTGG AGATTCTCA GACCCGGGTC GACCCTGTGG AAT 53

(2) INFORMATION FOR SEQ ID NO:65:

25 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 44 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
30 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

40 GACTAACCTT GATTCCCTGG AGCTCGAGGC GGCGCATCTC GGCG 44

(2) INFORMATION FOR SEQ ID NO:66:

(i) SEQUENCE CHARACTERISTICS:

- 45 (A) LENGTH: 47 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: DNA (genomic)

55 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

GACTAACCTT GATTCCCTGA AGACCTGCGT CATGCTGAGA CCCTCAA 47

(2) INFORMATION FOR SEQ ID NO:67:

60 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 50 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

65 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

5 GACTAACCTT GATTCCCTGA AGCGGCCAAT GCACCAAATG AAAGATTTTC 50

(2) INFORMATION FOR SEQ ID NO:68:

10 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 50 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA (genomic)

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

CGCATCTTTT AATTAAGTGG AGAATTTT TYACAGGCTT ATATAGKAAA 50

We claim:

1. A method for assembling a gene or gene vector comprising the steps of:
 - 5 a) designing at least 6 primers to produce at least three fragments in at least three separate polymerase chain reactions wherein each primer comprises at least one predetermined restriction endonuclease recognition site that recognizes a restriction endonuclease that cleaves at a distance from the recognition site, a sequence complementary to a template sequence for amplification, and bases positioned at the restriction endonuclease
10 cleavage site that are selected to be complementary to only one other overhanging created from enzymatic cleavage of the fragments;
 - b) combining the primers with template nucleic acid and performing a gene amplification reaction to produce multiple copies of an amplified template fragment incorporating the restriction endonuclease recognition site;
 - 15 c) digesting the amplified template fragments with one or more restriction endonucleases that recognize the restriction endonuclease recognition site of the primers to create overhanging termini wherein each overhanging termini is complementary to only one other overhanging termini on another fragment; and
 - d) combining the amplified and digested template fragments in a ligation
20 reaction to produce a directionally ordered gene, nucleic acid fragment or gene vector.
2. The method of claim 1 wherein the restriction endonuclease is at least one class IIS
25 restriction endonuclease.
3. The method of claim 2 wherein the class IIS restriction endonuclease is selected from the group consisting of: *AlwI*, *Alw26I*, *BbsI*, *BbvI*, *BbvII*, *BpmI*, *BsmAI*, *BsmI*, *BsmBI*, *BspMI*, *BsrI*, *BsrDI*, *Eco57I*, *EarI*, *FokI*, *GsuI*, *HgaI*, *HphI*, *MboII*, *MnII*, *PleI*, *SapI*, *SfaNI*, *TaqII*, *Tth111II*.
30
4. The method of claim 1 wherein class II restriction endonuclease recognition sites, linkers, or adapters are not used to create the gene or gene vector.

5. The method of claim 1 wherein the product of the ligation reaction is introduced into prokaryotic or eukaryotic cells.
- 5 6. The method of claim 1 wherein at least one target nucleic acid sequence is chosen from the group consisting of : transcriptional regulatory sequences; genetic vectors; introns and/or exons; viral encapsidation sequences; integration signals intended for introducing nucleic acid molecules into other nucleic acid molecules; retrotransposon(s); VL30 elements; or multiple allelic forms of a sequence.
- 10 7. The method of claim 1 wherein the method is used to generate combinatorial libraries of a target sequence.
8. The method of claim 7 wherein the target sequence is part or all of a gene.
- 15 9. The method of claim 8 wherein the gene encodes a protein.
10. The method of claim 8 wherein the primers amplify allelic variants of part or all of a gene.
- 20 11. The method of claim 1 wherein the product of the ligation reaction is passed between eukaryotic cells using a virus particle, by cell fusion, or by transfection.
12. The method of claim 1 wherein the product of the ligation reaction is not introduced
- 25 into prokaryotic cells.
13. The method of claim 1 further combining at least one screening or selection step to select the products of the ligation reaction.
- 30 14. The method of claim 1 wherein the product of the ligation reaction is mutated during passage in cells in order to generate genetic diversity.

15. The method of claim 14 wherein the product of the ligation reaction is mutated by-homologous recombination during passage in cells.
16. The method of claim 1, wherein the method is used to isolate and identify regulatory
5 sequences from a cell.
17. The method of claim 11, wherein cells containing the product of the ligation reaction are selected for enhanced biological activity.
- 10 18. The method of claim 17, wherein the cells containing the product of the ligation reaction are selected for tissue-specific, hormone-specific or developmental-specific gene expression.
19. The method of claim 1 wherein the product of the ligation reaction is a circularized
15 gene vector.
20. A nucleic acid primer having a 5' and a 3' end to amplify a nucleic acid fragment for the ligation of at least two fragments comprising:
- a restriction endonuclease recognition site that recognizes a restriction endonuclease,
 - 20 wherein the restriction endonuclease cleaves at a distance from the recognition site and creates overhanging termini;
 - a sequence complementary to a template sequence to be amplified to produce the nucleic acid fragment;
 - at least two nucleic acid bases positioned at the restriction endonuclease cleavage site
 - 25 and that form an overhanging terminus after cleavage by the restriction endonuclease, wherein the at least two nucleic acid bases are selected to be complementary to only one other overhanging terminus on another fragment of the ligation; and
 - an affinity handle on the 5' end of the primer.
- 30 21. The primer of claim 20 further comprising an anchor to provide stability to the restriction enzyme at the restriction enzyme recognition site.

22. A method for isolating and identifying promoters comprising the steps of:

a) obtaining a vector comprising at least a portion of a promoter region from a retrovirus transposon LTR and having two non-complementary overhanging termini;

b) designing at least two PCR primers to amplify at least one region of a retro-transposon LTR from template nucleic acid to produce at least one nucleic acid fragment wherein each primer comprises at least one predetermined restriction endonuclease recognition site that recognizes a restriction endonuclease that cleaves at a distance from the recognition site, a sequence complementary to a template sequence from a retrovirus transposon, and bases positioned at the restriction endonuclease cleavage site that are selected to be complementary to only one other overhanging terminus of the vector wherein the restriction endonuclease cleavage site is created from enzymatic cleavage of the fragments;

c) combining the primers with template nucleic acid and performing a gene amplification reaction to produce multiple copies of an amplified template fragment incorporating the restriction endonuclease recognition site;

d) digesting the amplified template fragments with one or more restriction endonuclease that recognize the restriction endonuclease recognition site of the primer to create overhanging termini; and

e) combining the amplified and digested template fragment in a ligation reaction with the vector to produce a gene vector with an intact LTR sequence.

20

23. The method of claim 22 wherein the template nucleic acid is DNA or RNA.

24. The method of claim 22 further comprising the step of sequencing the insert to identify the promoter sequence.

25

25. Promoter sequences of SEQ ID NOS:2-13 identified using the methods of claim 22.

26. The vector of SEQ ID NO:1.

30

Fig 1A

A.

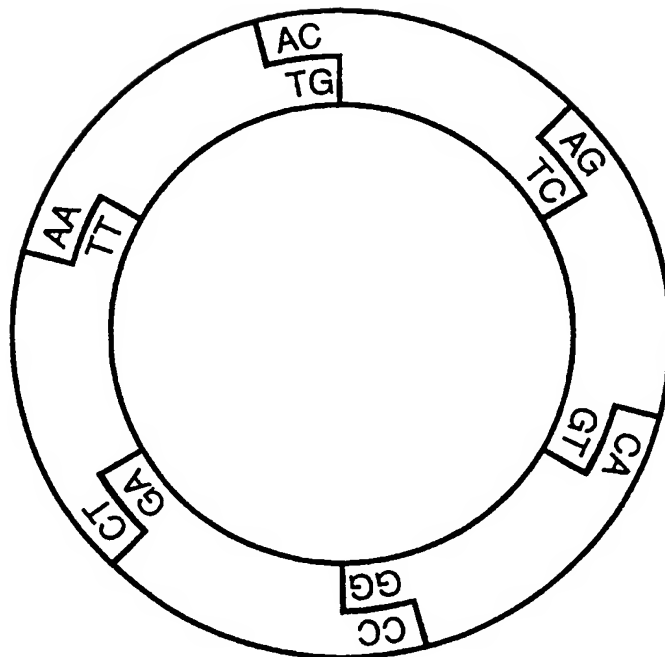
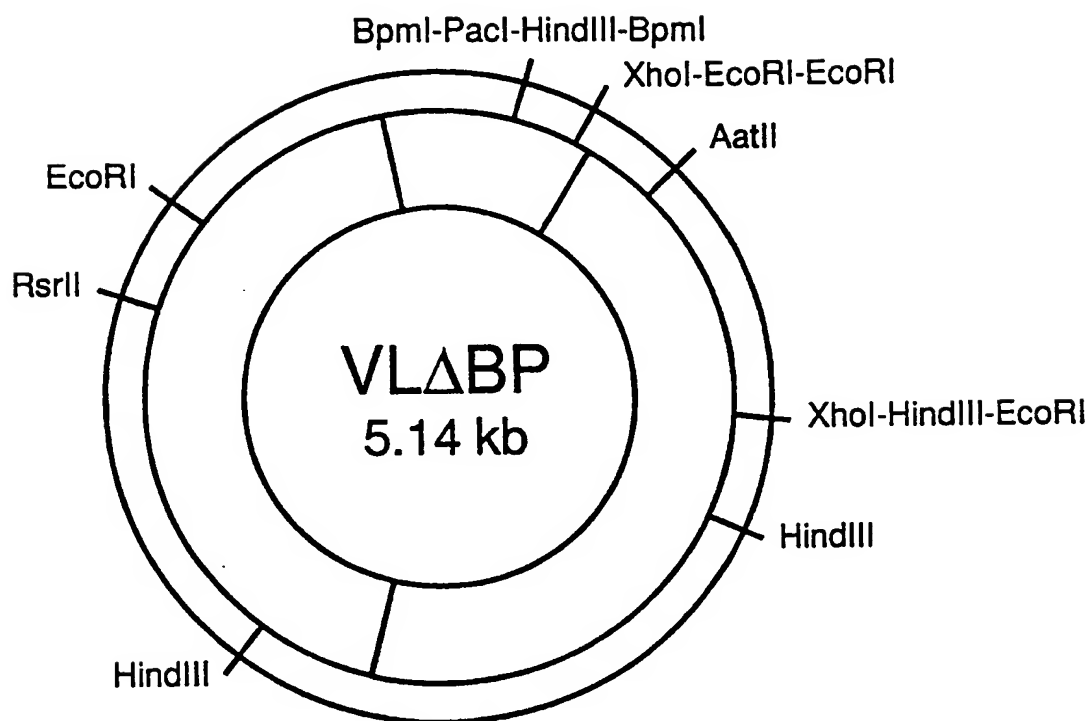


Fig 1B

B.



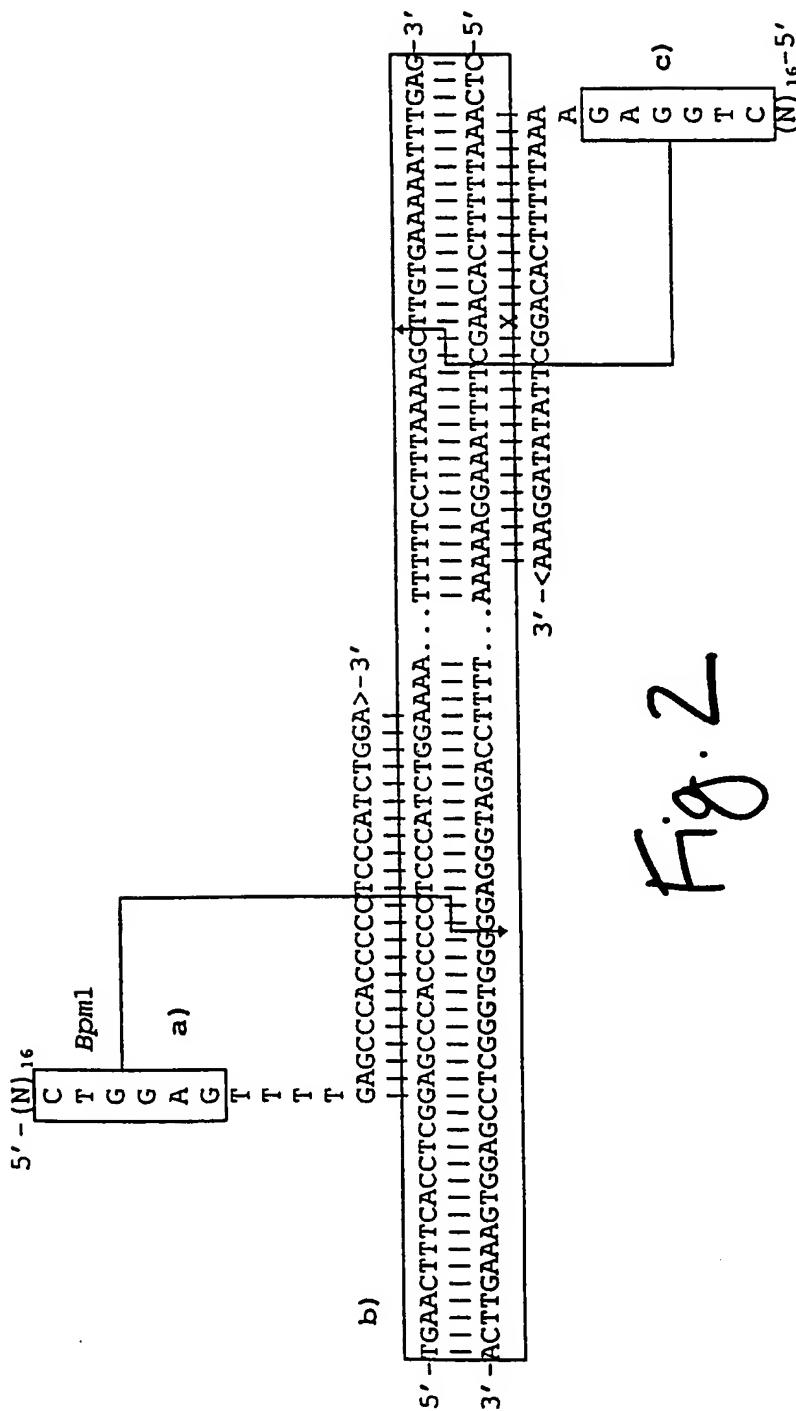
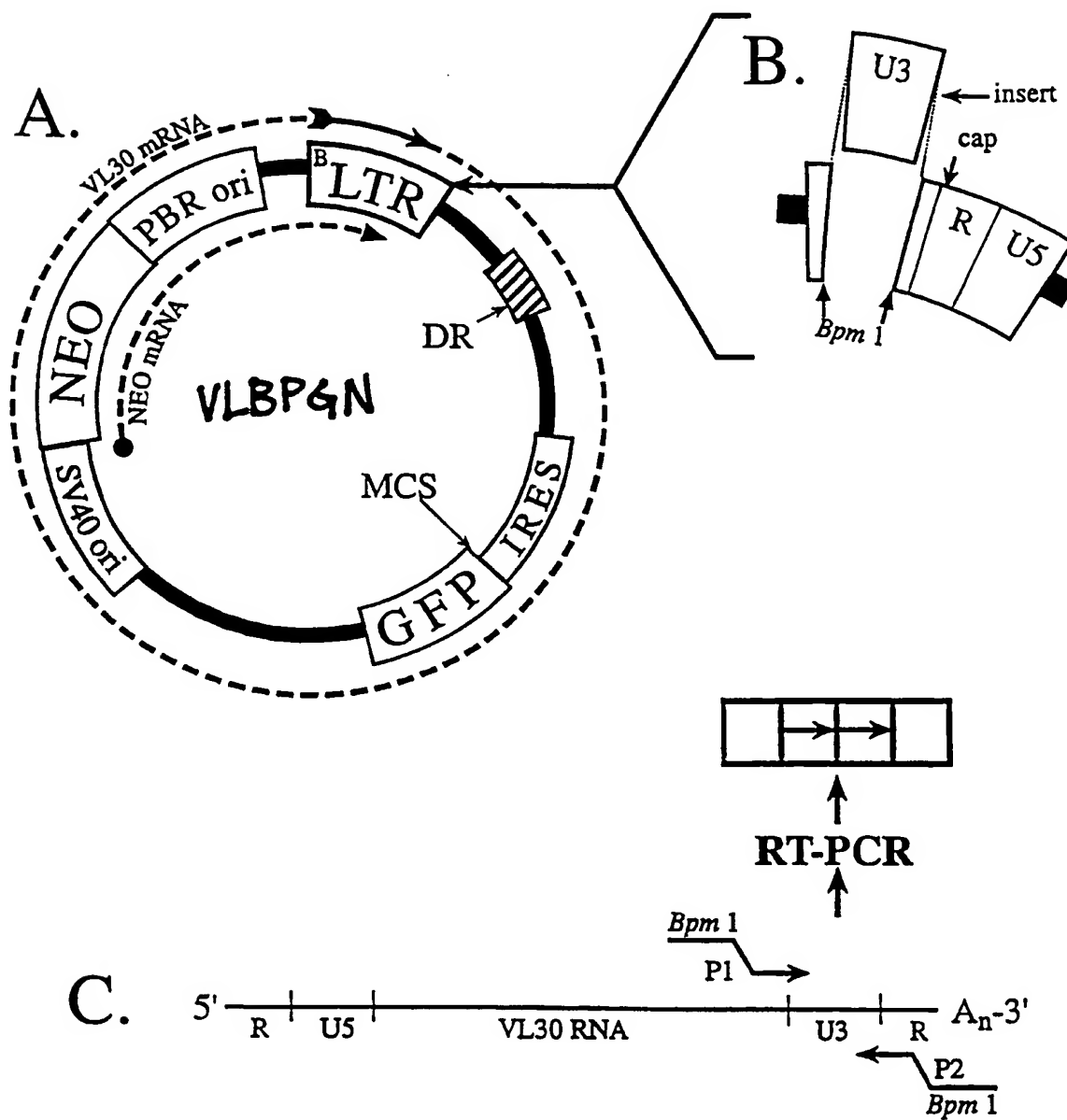


Fig. 3



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Fig. 4 A.

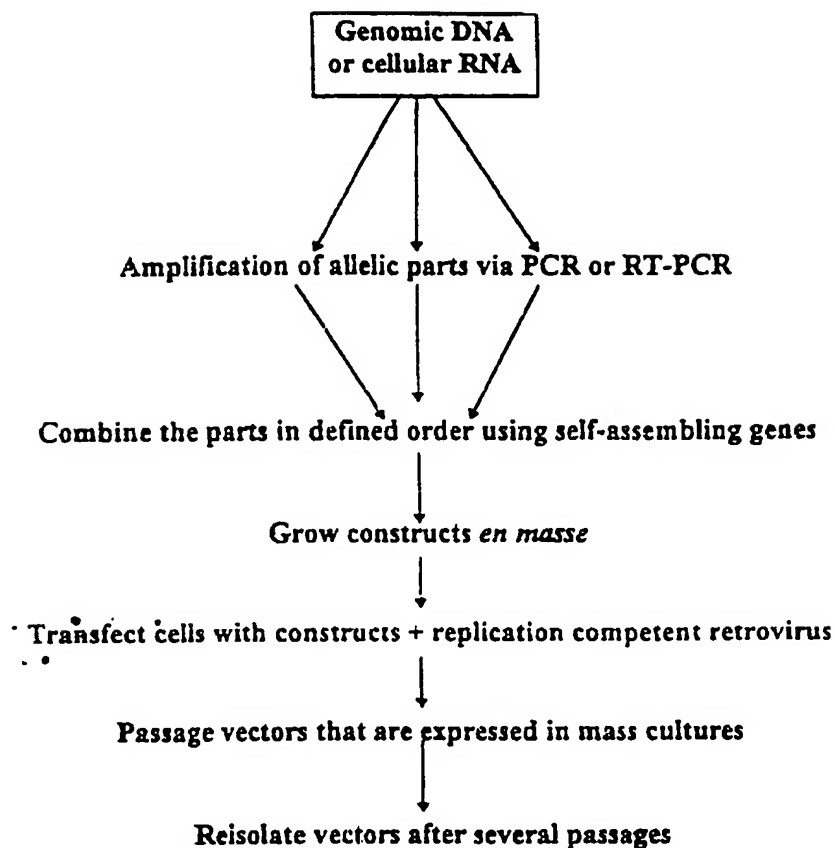


Fig 4 B.

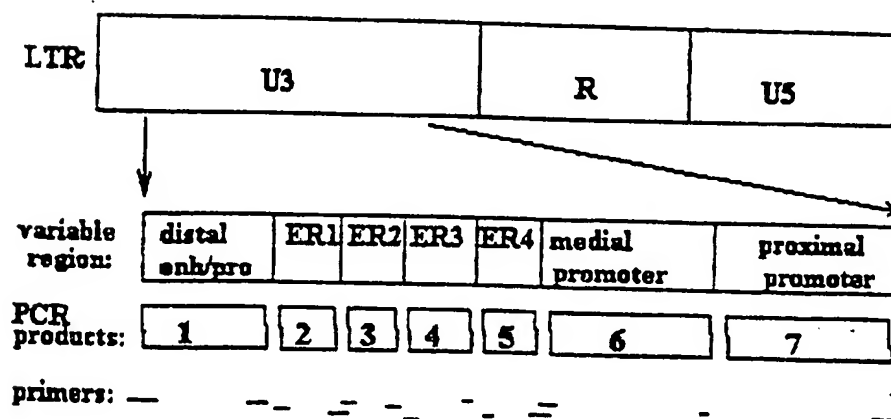


Fig. 5

```

1      1  CCTCCCATCT AGAGGTGTGT CTCGGAACAC TCCTAAACTT TTCACCCCAA AACTCCCTCAG CCTAAAGTTC GAAAJAACTG TTCCAAAGAAC      90
I.D.2
I.D.3

91      1  ATTTTTCAGA TAAAGGCCTC CTAGAACAAAC CTCAAJAATGA CATTGCCJAA TGAATAAGACA TGACTCCCTTA GTTACGTAGG TTCCCTTGATA      180
I.D.2
I.D.3

181     1  GGACATGACT CCTTAGCTTAC GTAGGTTTCT TGATAGGACA TGACTCCCTTA GTTACGTAGA TTCCCTTTGGT AGAAGCTCCCT AGTGATGTAA      270
I.D.2
I.D.3

271     1  ACCAGGTACT CCTTAGCTTAC GTAGGTTTCT TGATAGGACA TGACTCCCTTA GTTACGTAGA TTCCCTTTGGC AGAAGCTCCCT AGTGATGTAA      360
I.D.2
I.D.3

321     1  ACTTGACTTT TCCCTGCCCA GTTCTGCCCC TTGAGTGTCT ACTATATATAG C
I.D.2
I.D.3

```

Fig. 6

1 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC 90
4 5 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
6 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
7 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
8 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
9 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
10 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
11 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
12 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC
13 CCTCCCATCT AGAGATGTTT CCCAGAACAC TCCTGAACTC TTCACCCCCAG AATGCATGCC TGAACCTCCTC ACCCTAGAGT TCGAACCCCTC

91 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA 180
4 5 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
6 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
7 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
8 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
9 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
10 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
11 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
12 CCAACTAAAG ACTGTTCCAA GAACATTTTT GAGATAAGGG CCTCCTGGAA CAACCTCAGA ATGAAACCGG TACATTGCCA AATAATAGGA
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Fig. 7

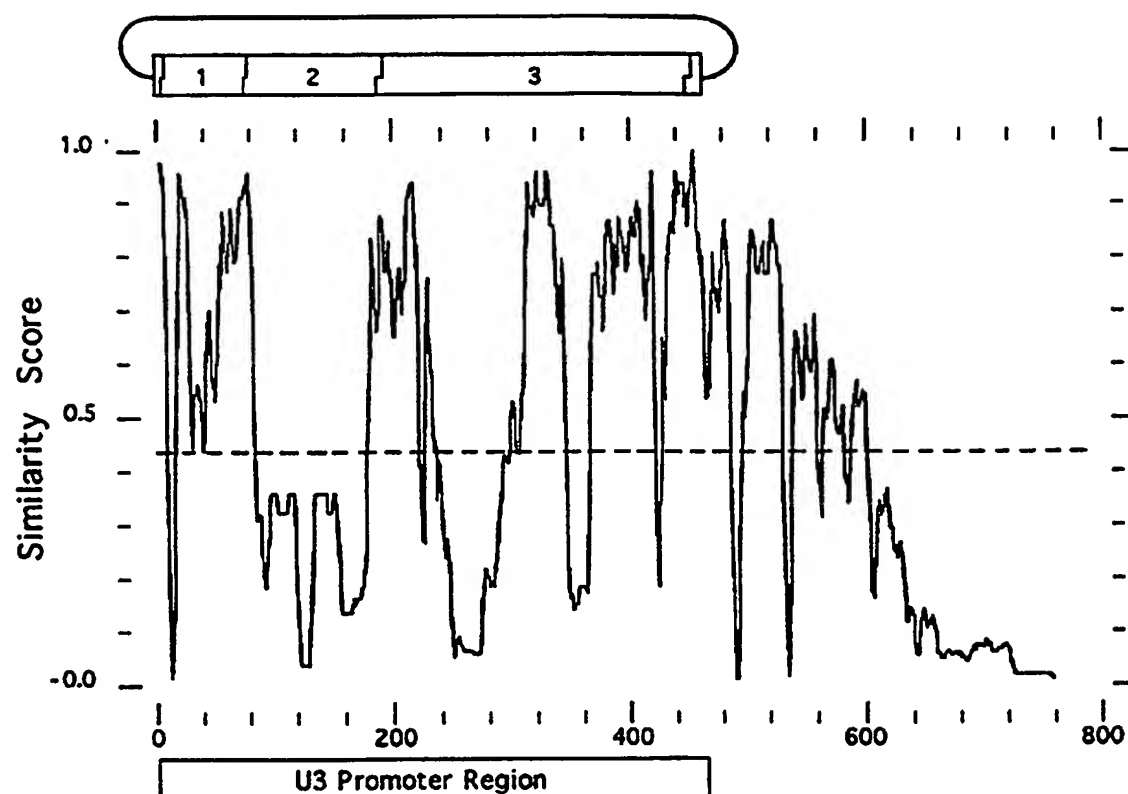
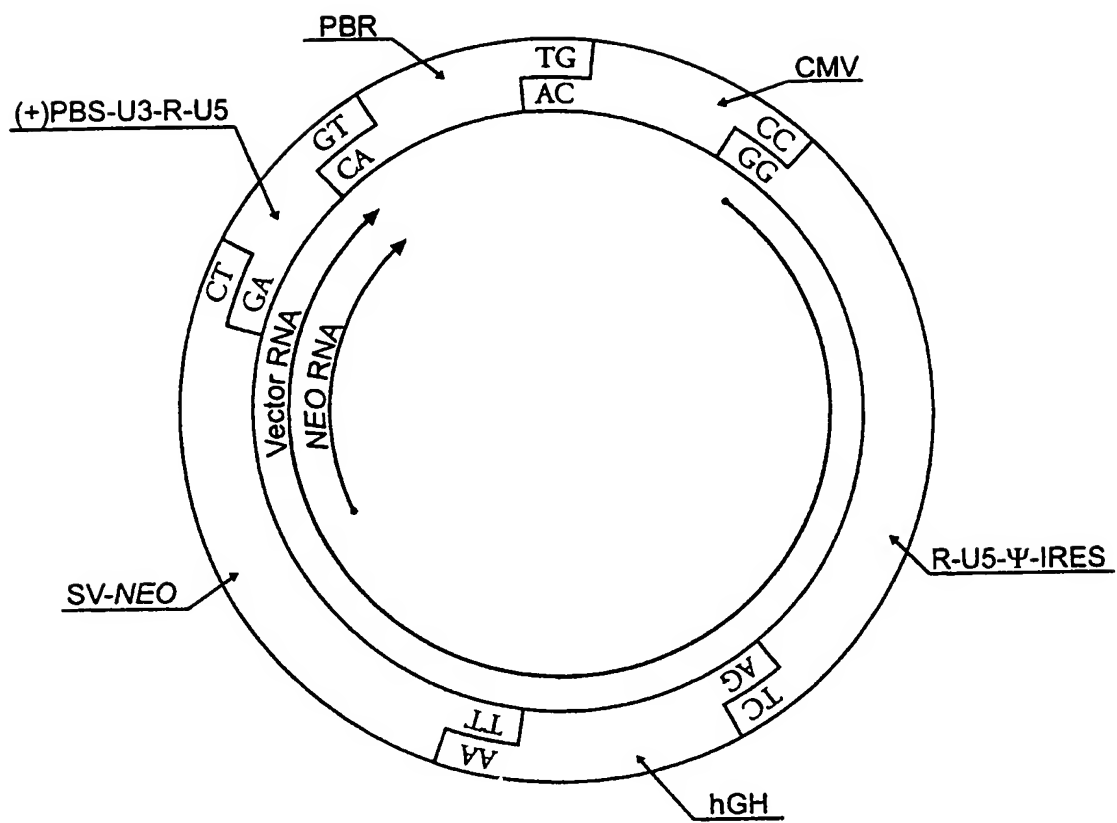


Fig. 8



INTERNATIONAL SEARCH REPORT

I. International Application No

PCT/US 98/03918

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/86 C12N15/10

According to International Patent Classification(IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	PADGETT K A ET AL: "Creating seamless junctions independent of restriction sites in PCR cloning" GENE, vol. 168, no. 1, 2 February 1996, page 31-35 XP004042930	1,2, 4-14, 19-21
Y	see the whole document ---	3
Y	TOMIC, M. ET AL.: "A rapid and simple method for introducing specific mutations into any position of DNA leaving all other positions unaltered" NUCLEIC ACIDS RESEARCH, vol. 18, no. 6, 1990, OXFORD GB. page 1656 XP002069445 cited in the application see the whole document ---	3
-/--		



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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- "S" document member of the same patent family

Date of the actual completion of the international search

26 June 1998

Date of mailing of the international search report

09/07/1998

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl.
Fax: (+31-70) 340-3016

Authorized officer

Chambonnet, F

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/03918

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>LEBEDENKO, E.N. ET AL.: "Method of artificial DNA splicing by directed ligation"</p> <p>NUCLEIC ACIDS RESEARCH, vol. 19, no. 24, 1991, OXFORD GB, pages 6757-6761, XP002069446 cited in the application see the whole document</p> <p style="text-align: center;">---</p>	1
A	<p>CHAKRABORTY, A.K. ET AL.: "Synthetic retrotransposon vectors for gene therapy"</p> <p>FASEB JOURNAL., vol. 7, no. 10, July 1993, FOR EXPERIMENTAL BIOLOGY, BETHESDA, MD US, pages 971-977, XP002029486 see the whole document</p> <p style="text-align: center;">---</p>	1
P,X	<p>WO 97 28282 A (STRATAGENE INC) 7 August 1997</p>	1.2, 4-14, 19-21
P,Y	<p>see the whole document</p> <p style="text-align: center;">---</p>	3
P,X	<p>HODGSON, C.P. ET AL.: "Self-assembling genes (SAGE): construction of complex vectors and combinatorial libraries without sub-cloning"</p> <p>CANCER GENE THERAPY, vol. 4, no. 6 conf. suppl., November 1997, page s27 XP002069448 see the whole document</p> <p style="text-align: center;">---</p>	1
P,X	<p>ZINK, A. M. ET AL.: "Transcriptional targeting with rescued LTRs: a hepatocyte promoter"</p> <p>CANCER GENE THERAPY, vol. 4, no. 6 conf. suppl., November 1997, page s28 XP002069449 see the whole document</p> <p style="text-align: center;">-----</p>	22

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 98/03918

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9728282 A	07-08-1997	NONE	